110 0501071517 050	0.0.00000000000000000000000000000000000	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \
U.S. DEPARTMENT OF C	OMMERCE PATENT AND TRADEMARK OFFICE	ATTORNEY'S DOCKET NUMBER UEMURA 7
		UEMURA /
TRANMITTAL LETTER	TO THE UNITED STATES	
DESIGNATED/ELECT	ED OFFICE (DO/EO/US)	U.S. APPLICATION NO. (If known, see 37 CFR 1.5)
CONCERNING A FILE	NG UNDER 35 U.S.C. 371	09/856371
INTERNATIONAL APPLICATION NO.	INTERNATIONAL FILING DATE	PRIORITY CLAIMED
PCT/JP99/06475	19 November 1999	20 November 1998
TITLE OF INVENTION		
NOVEL SERINE PROTEASE	BSSP2	
APPLICANT(S) FOR DO/EO/US		
Hidetoshi UEMURA et al.		
Applicant herewith submits to the United	d States Designated/Elected Office (DO/EO/	US) the following items and other information:
	of items concerning a filing under 35 U.S.C	
2. [] This is a SECOND or SUBS	EQUENT submission of items concerning	a filing under 35 U.S.C. 371.
3. [X] This is an express request to	begin national examination procedures (35	U.S.C. 371(f)) at any time rather than delay
examination until the expira	tion of the applicable time limit set in 35 U.	S.C. 371(b) and PCT Articles 22 and 39(1).
4. [X] The US has been elected in a	Demand by the expiration of 19 months from	m the priority date (PCT Article 31).
5. 🕍 A copy of the International A	pplication as filed (35 U.S.C. 371(c)(2))	
a llicattached hereto (re	quired only if not transmitted by the Interna	tional Bureau).
b. [X] has been communicated by the second interest of the second in	ted by the International Bureau.	
c. [] is not required, as the	application was filed in the United States I	Receiving Office (RO/US).
6. 🔀 An English language translati	on of the International Application as filed (35 U.S.C. 371(c)(2)).
7. [X] Amendments to the claims of	the International Application under PCT Ar	ticle 19 (35 U.S.C. 371(c)(3))
a. [] are transmitted herev	vith (required only if not transmitted by the	International Bureau).
b. [] have been communic	ated by the International Bureau.	
	however, the time limit for making such am-	endments has NOT expired.
d. [X] have not been made a		
- 5 - 90°	on of the amendments to the claims under P	CT Article 19 (35 U.S.C. 371(c)(3)).
9. [X] An oath or declaration of the i		
10. [An English language translati	on of the annexes to the International Prelin	ninary Examination Report under PCT Article 36
(35 U.S.C. 37I(c)(5)).		
Items 11. to 16. below concern docume	• •	
11. [] An Information Disclosure St		''41 27 OFF 2 00 - 12 21 ' ' 1 1 1
		iance with 37 CFR 3.28 and 3.31 is included.
13. [X] A FIRST preliminary amendm		
[] A SECOND or SUBSEQUEN 14. [] A substitute specification.	1 premimary amendment.	
15. [] A change of power of attorney	and/or addross latter	
16. [X] Other items or information:	and or address letter.	
	page of the International Publication (WO	00/31272).
	mational Preliminary Examination Report (I	n Japanese).
[X] Formal drawings, 07 shee		
[X] Courtesy Copy of the Inte	rnational Search Report.	
[X] Application Data Sheet	he) assigned to: FUSO PHADMACEUTICA	L INDUSTRIES, LTD. whose address is 7-10, Doshomachi
1-chome, Chuo-ku, Osaka-shi, Osaka, Jar		which contract, bib. whose address is /-10, Doshomach

Page 1 of 2

PCT09

RAW SEQUENCE LISTING DATE: 10/11/2001 PATENT APPLICATION: US/09/856,371 TIME: 09:51:46

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Output Set: N:\CRF3\10112001\I856371.raw

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9	<120>	· TITI	E OF	INVE	NTIO	N: N	OVEL	SER	INE	PROT	EASE	BSS	P2				
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40		u. 01	1 011	5	7114	, 41	711LG	001	10	**** 9		110		15	2114		
F 717 700F	agc g	to at	a ctt	-	too	caa	cac	aca		ααα	acc	tot	αtα		aca	96	
	Ser V															50	
44	DCI V	41 110	20	011	DCI	1119	1110	25	Cyb	GLY	niu	DCI	30	пси	ALG		
	cca c	ac to		ata	act	act	acc		tac	atσ	tac	ant		a a a	c+a	144	
	Pro H					-	_		_	_		-			_	144	
48	110 11	35		vul	1111	ALG	40	1115	Cys	Het	1 Y 1	45	rne	АТУ	Lieu		
	tcc c			agg	taa	caa		cat	αca	ααα	ata		agg	oat	aat	192	
	Ser A															192	
52		о О	u ser	Set	ттЪ	55	Val	птэ	нта	СТУ	60	Val	ser	птъ	GTA		
				~~~	a2a		20+	2+4	~+ ~	~~~		a+ a	~++	a a ±	+	240	
	gct g Ala V															240	
	65	ar Ar	g GIII	птэ	70	СТА	T 11T	Met	vaı		гуѕ	TTE	тте	PIO			
		+~ +>	~ ~~+	~~~			~~ t-	~~~	+-+	75 ~~ <b>+</b>		~~+	~		80	200	
	cct t															288	
	Pro L	eu ry	r ser		GIII	ASII	HIS	ASP	_	Asp	val	ALa	ьeu		GIN		
60	a+a =			85		++-	+		90	<b>.</b>				95		226	
	ctc c															336	
	Leu A	rg rn		тте	ASN	rne	ser	_	rnr	val	Asp	ата		Cys	ьeu		
64	~~~		100		<b>.</b>			105		<b>L</b>		4	110			20.	
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67	Pro A	та Гу	s Glu	GIn	${ t ryr}$	Phe	Pro	${\tt Trp}$	GLY	Ser	Gln	Cys	Trp	Val	Ser		

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= 102 = 104						, ър.												
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		1> L															
		2> T															
		3> 01			Mus	SD.											
		0> F			2240	J.											
		1> N			CDS												
		2> L(				71	1106	<b>5</b> ١									
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																gcacat	120
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= 186	gga																288
187				ilu <i>E</i>	Ата (			età i	Leu 1	Jeu :			Ser A	Ala A	Asn (	Cys	
188		_	L				5					10					
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191 192	15					20					25					30	
194																	384
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215	Lvs	Tle	Tle	Dro	Hic	Dro	T.011	Tur	Cor	λla	Cln	Agn	His	7 an	Trzz	yar Nan	024
216	цур	110	110	110	115	110	пси	тут	Der	120	GIII	ASII	птъ	кор	125	ASP	
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	a a a	aa+	ata		++~	000	aca	224		<b>~</b>	+	+++	<b>ac</b> -	140	~~~	+	700
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Input Set : A:\sequence listing.txt
Output Set: N:\CRF3\10112001\1856371.raw

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		\		TOTAL.	A												
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Input Set : A:\sequence listing.txt
Output Set: N:\CRF3\10112001\1856371.raw

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 314 Ile Pro His Pro Leu Tyr Ser Ala Gln Asn His Asp Tyr Asp Val Ala
 315
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                                  120
                                                       125
 318 Leu Leu Gln Leu Arg Thr Pro Ile Asn Phe Ser Asp Thr Val Asp Ala
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 322 Val Cys Leu Pro Ala Lys Glu Gln Tyr Phe Pro Trp Gly Ser Gln Cys
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 326 Trp Val Ser Gly Trp Gly His Thr Asp Pro Ser His Thr His Ser Ser
 327
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                                          170
 330 Asp Thr Leu Gln Asp Thr Met Val Pro Leu Leu Ser Thr His Leu Cys
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 334 Asn Ser Ser Cys Met Tyr Ser Gly Ala Leu Thr His Arg Met Leu Cys
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                                  200
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343 225 230 235 240
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                                             Met His Ile Cys Lys Ser
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 390 ctt ggg cat atc agg ctt act caa cac aag gcc gtg aat ctg tct gac
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 391 Leu Gly His Ile Arg Leu Thr Gln His Lys Ala Val Asn Leu Ser Asp
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Use of n and / or Xaa has been detected in the Sequence Listing. Review the Sequence Listing to ensure a corresponding explanation is present in the <220> to <223> fields of each sequence using n or Xaa.

### VERIFICATION SUMMARY

DATE: 10/11/2001

PATENT APPLICATION: US/09/856,371

TIME: 09:51:47

Input Set : A:\sequence listing.txt
Output Set: N:\CRF3\10112001\1856371.raw

L:1325 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:20 L:1355 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:21

THY

#### IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

ATTY.'S DOCKET: UEMURA7

In re Application of:

H. UEMURA, et al.

Serial No.: 09/856,371

Confirmation No.

Filed: May 21, 2001

Washington D.C.

For: NOVEL SERINE PROTEASE

BSSP2

September 4, 2001

RESPONSE TO NOTIFICATION TO COMPLY WITH REQUIREMENTS FOR PATENT APPLICATIONS CONTAINING NUCLEOTIDE SEQUENCE AND/OR AMINO ACID SEQUENCE DISCLOSURES

Honorable Commissioner for Patents Washington, D.C. 20231

In response to the Notice to Comply, dated July 3, 2001, and prior to the examination of the above-described application, please amend the present application as follows:

### IN THE SPECIFICATION

Please replace the paragraph beginning at page 19, line 10, with the following rewritten paragraph:

--The novel mouse serine protease can be divided into types 1, 2 and 3. It has been shown that type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids, and type 3 is composed of 445 amino acids. These amino acid sequences contain a common amino acid sequence of 238 amino acids whose N-terminus side starts with Ile-Val-Gly-Gly-Gln-Ala-Val (amino acid

1-7 of SEQ ID NO:2) as the mature serine protease. Further, the amino acid sequence of the mature serine protease contains a consensus sequence having serine protease activity. Since there are two or more amino acid sequences which are characteristic of sugar chain binding sites, the amino acid sequence is presumed to have at least two sugar chains.--

Please replace the paragraph beginning at the bottom of page 26, line 14, with the following rewritten paragraph:

-- The vector is not specifically limited in so far as it can express the protein of the present invention. Examples thereof include pBAD/His, pRSETA, pcDNA2.1, pTrcHis2A, pYES2, pBlueBac4.5, pcDNA3.1 and pSecTag2 manufacture by Invitrogen, pET and pBAC manufactured by Novagen, pGEM manufactured by Promega, pBluescriptII manufactured by Stratagene, pGEX and pUC18/19 manufactured by Pharmacia, PfastBAC1 manufactured by GIBCO and the like. Preferably, a protein expression vector (described in the specification of a patent application entitled "Protein expression vector and its use" and filed by the same applicant on the same day) is used. This expression vector is constructed by using pCRII-TOPO vector described in the Examples hereinafter, or a commercially available expression vector, for example pSecTag2A vector or pSecTag2B vector (Invitrogen) and integrating a secretory signal nucleotide sequence suitable for expression of the protein of the present invention, in the 3' downstream side

thereof, a Tag nucleotide sequence, a cleavable nucleotide sequence sequence and a cloning site, into which a nucleotide sequence encoding a target protein can be inserted, in this order. More specifically, it is preferred to use trypsin signal as the secretory signal, a nucleotide sequence encoding polyhistidine as the Tag nucleotide sequence, and a nucleotide sequence encoding an amino acid sequence which is susceptible to enzyme-specific cleavage, i.e., a nucleotide sequence encoding the amino acid sequence of Asp-Asp-Asp-Lys SEQ ID NO:42 (said amino acid sequence is recognized by enterokinase, and the recombinant fusion protein is cleaved at the C-terminus part thereof) as the cleavable nucleotide sequence.—

Please replace the paragraph beginning at page 49, line 22, with the following rewritten paragraph:

--The cloning was carried out by PCR using a mouse brain cDNA library (Clontech) as a template and nucleotide sequences corresponding to an amino acid sequence common to serine proteases represented by

Primer 1: GTG CTC ACN GCN GCB CAY TG (SEQ ID NO: 20)

Primer 2: CCV CTR WSD CCN CCN GGC GA (SEQ ID NO: 21) as primers. Namely, 5 µl of the template, 5 µl of 10 x ExTaq buffer, 5 µl of dNTP, 10 pmol of each of the above primers and 0.5 µl of ExTaq (TAKARA) were added and the total volume was adjusted to 50 µl with sterilized water. PCR was carried out by

repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for 1 minute, 30 times. The PCR product was mixed with pCR II-TOPO vector attached to TOPO TA cloning kit (Invitrogen) and the mixture was allowed to stand at room temperature for 5 minutes. Then, according to a conventional manner, E. coli Top 10 attached to the kit was transformed and applied to a LB  $(Amp^+)$  plate (containing 100 µg/ml of ampicillin). According to a conventional manner, a plasmid was extracted from each colony obtained and its nucleotide sequence was determined by cycle sequencing method with a fluorescence sequencer (ABI). Homology of the sequence of each clone was examined by means of GenBank. Regarding an unknown sequence, i.e., BSSP2 gene, the full length cDNA was obtained by 5' RACE and 3' RACE and, according to the same manner as described above, the nucleotide sequence was determined. Namely, BSSP2 clone specific primers, GSP1 primers [mBSSP2.2 (SEO ID NO: 27) or mBSSP2.0 (SEQ ID NO: 22)] and GSP2 primers [mBSSP2R2 (SEQ ID NO: 28) or mBSSP2.1 (SEQ ID NO: 23)] were prepared. PCR was carried out by using mouse brain Marathon-Ready cDNA (Clontech), AP1 primer attached to this reagent and either of the above GSP1 primers and heating at 94°C for 2 minutes once and repeating a cycle of heating at 94°C for 30 seconds, at 60°C for 30 seconds and then at 72°C for 30 seconds 35 times. Then, 5  $\mu$ l of the PCR product diluted to 1/100, 5  $\mu$ l of 10 x buffer, 5  $\mu l$  of dNTP, 10 pmol of either of 10  $\mu M$  of the

above GSP2 primer, 10 pmol of AP2 primer attached to the above reagent and 0.5 unit of ExTag were admixed and adjusted to 50 ul with sterilized water. Then, according to the same manner as the above, PCR was carried out. The PCR product was cloned by the above TOPO TA cloning kit and sequenced to obtain the upstream and downstream regions of the above clone. At this time, as for a clone which seemed not to cover the full length of a protein, the specific primers shown hereinafter were prepared based on the newly found nucleotide sequence. Further, based on this sequence, the primers capable of amplifying ORF as shown hereinafter [mBSSPF7 (SEQ ID NO: 26), mBSSP2R/E (SEQ ID NO: 29)] were prepared and PCR carried out using mouse brain Marathonready cDNA as a template to confirm that these clones were identical. This was cloned into pCR II-TOPO vector attached to TOPO TA cloning kit to obtain the plasmid pCR II/mBSSP2 containing the full length cDNA clone. The nucleotide sequence of DNA contained in this plasmid is shown in SEQ ID NO: 7 and the amino acid sequence of mSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 8. Further, two different types of clones were obtained. The nucleotide sequences of these DNA are shown in SEQ ID NOS: 3 and 5, respectively. The amino acid sequences of mBSSP2 proteins deduced from these nucleotide sequences are shown in SEQ ID NOS: 4 and 6. These novel proteases are divided into types 1, 2 and 3. Type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids and

type 3 is composed of 445 amino acids. These amino acid sequences contained the common amino acid sequence composed of 238 amino acids whose N-terminus side started with Ile-Val-Gly-Gly-Gln-Ala-Val (amino acid 1-7 of SEQ ID NO:2) as the mature serine protease. Further, in the amino acid sequence of the mature serine protease, a consensus sequence having a serine protease activity was contained. Furthermore, since there were two or more amino acid sequences specific for a sugar chain bonding site, it was presumed that the amino acid sequence had at least two sugar chains.--

Please replace the paragraph beginning at the bottom of page 56, line 24, with the following rewritten paragraph:

--Amplification was carried out by using the primers having the sequences represented by SEQ ID NOS: 15 and 16 so that the peptide of Leu-Val-His-Gly (SEQ ID NO:43) was present at the C-terminus of the part from trypsin signal to the enterokinase recognition site of pSecTrypHis/neurosin. This was inserted between NheI and HindIII sites of pSecTag2A to construct the plasmid pTrypSig.--

Please replace the paragraph beginning at page 61, line 3, with the following rewritten paragraph:

--Reverse transcription of 1 µg of mRNA of human fetus brain (Clontech) was carried out by using Superscript II (Gibco

BRL) and oligo dT-Not I primer (5' GGCCACGCGTCGACTAGTA C(T) $_{17}$  3' SEQ ID NO:44) to obtain cDNA. By using this as a template, PCR was carried out with primes prepared from mBSSP2 nucleotide sequence and represented by SEQ ID NOS: 30 and 31 to obtain a cDNA fragment of hBSSP2. Namely, 5 µl of the template, 5 µl of 10 x ExTag buffer (TAKARA), 5 µl of dNTPs, 10 pmol portions of the above primers and 0.5  $\mu$ l of ExTaq (TAKARA) were adjusted to 50 µl with sterilized water and PCR was carried out by repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for 1 minute, 35 times. The PCR reactions described hereinafter were carried out according to the same manner as the above composition and conditions except the template and primers. The PCR product was mixed with pGEM-T Easy vector (Promega) and Takara Ligation Solution I (TAKARA) and the reaction was carried out at 16°C for 2 hours. Then, according to the same manner, E. coli JM109 was transformed and applied to a LB (Amp⁺) plate. A plasmid was extracted from each colony formed according to a conventional manner and its nucleotide sequence was determined by dideoxy method. As for a clone having homology to mBSSP2, full length cDNA was obtained by 5' RACE and 3' RACE and its sequence was determined as described above. PCR was carried out by using the above cDNA as a template and primers having the sequences represented by SEQ ID NOS: 30 and 37. RACE was carried out by PCR using a 1/100 dilution of the above PCR product as a template and primers having the sequences

represented by SEQ ID NOS: 32 and 37. As for 5' RACE, cDNA for RACE was prepared from human fetal brain mRNA (Clontech) by using Superscript II and SMART RACE cDNA amplification kit (Clontech). PCR of this cDNA was carried out by using a primer of 10  $\times$ Universal Primer Mix (attached to the kit) and a primer having the sequence represented by SEQ ID NO: 33. Further, PCR was carried out by using the 1/100 dilution of the latter PCR product, a template, Nested PCR Primer (attached to the kit) and a primer having the sequence represented by SEQ ID NO: 34. finally obtained PCR product was subjected to TA cloning as described above and the nucleotide sequence was determined to obtain the upstream and downstream regions of the above clone. In addition, primers for amplifying the full length cDNA as represented by SEQ ID NOS: 35 and 36 were prepared based on the resultant nucleotide sequence and PCR was carried out by using the above synthetic cDNA as a template. This PCR product was cloned into pGEM-T Easy vector to obtain the plasmid pGEM-TE/hBSSP2 containing the full length cDNA clone. The DNA sequence contained in this plasmid is shown in SEQ ID NO: 9 and hBSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 10.--

### IN THE SEQUENCE LISTING

Please substitute the attached Sequence Listing for that originally filed.

#### REMARKS

Applicants have added into the present specification a substitute paper copy Sequence Listing section according to 37 C.F.R. §1.821(c) as new pages 1-28. Furthermore, attached hereto is a 3 1/2" disk containing the "Sequence Listing" in computer readable form in accordance with 37 C.F.R. §1.821(e).

Applicants have amended the specification to insert SEQ ID Nos, as supported in the present specification.

The following statement is provided to meet the requirements of 37 C.F.R. §1.825(a) and 1.825(b).

I hereby state, in accordance with 37 C.F.R. §1.825(a), that the amendments included in the substitute sheets of the sequence listing are believed to be supported in the application as filed and that the substitute sheets of the sequence listing are not believed to include new matter.

I hereby further state, in accordance with 37 C.F.R. \$1.825(b), that the attached copy of the computer readable form is the same as the attached substitute paper copy of the sequence listing.

Under U.S. rules, each sequence must be classified in <213> as an "Artificial Sequence", a sequence of "Unknown" origin, or a sequence originating in a particular organism, identified by its scientific name.

Neither the rules nor the MPEP clarify the nature of the relationship which must exist between a listed sequence and

an organism for that organism to be identified as the origin of the sequence under <213>.

Hence, counsel may choose to identify a listed sequence as associated with a particular organism even though that sequence does not occur in nature by itself in that organism (it may be, e.g., an epitopic fragment of a naturally occurring protein, or a cDNA of a naturally occurring mRNA, or even a substitution mutant of a naturally occurring sequence). Hence, the identification of an organism in <213> should not be construed as an admission that the sequence per se occurs in nature in said organism.

Similarly, designation of a sequence as "artificial" should not be construed as a representation that the sequence has no association with any organism. For example, a primer or probe may be designated as "artificial" even though it is necessarily complementary to some target sequence, which may occur in nature. Or an "artificial" sequence may be a substitution mutant of a natural sequence, or a chimera of two or more natural sequences, or a cDNA (i.e., intron-free sequence) corresponding to an intron-containing gene, or otherwise a fragment of a natural sequence.

The Examiner should be able to judge the relationship of the enumerated sequences to natural sequences by giving full consideration to the specification, the art cited therein, any further art cited in an IDS, and the results of his or her

In re Appln. No.: 09/856,371

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sequence search against a database containing known natural sequences.

Attached hereto is a marked-up version of the changes made to the specification by the current amendment. The attached page is captioned "Version with markings to show changes made".

Applicants submit that the present application contains patentable subject matter and therefore urge the examiner to pass the case to issuance.

If the examiner has any questions or comments concerning the above described application, the examiner is urged to contact the undersigned at the phone number below.

Respectfully submitted,

BROWDY AND NEIMARK, P.L.L.C. Attorneys for Applicant(s)

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F:\,A\Aoyb\Uemura 7\PTO\notice to comply with seq.wpd

#### VERSION WITH MARKINGS TO SHOW THE CHANGES MADE

The paragraph beginning at page 19, line 10, has been amended as follows:

The novel mouse serine protease can be divided into types 1, 2 and 3. It has been shown that type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids, and type 3 is composed of 445 amino acids. These amino acid sequences contain a common amino acid sequence of 238 amino acids whose N-terminus side starts with Ile-Val-Gly-Gly-Gln-Ala-Val (amino acid 1-7 of SEQ ID NO:2) as the mature serine protease. Further, the amino acid sequence of the mature serine protease contains a consensus sequence having serine protease activity. Since there are two or more amino acid sequences which are characteristic of sugar chain binding sites, the amino acid sequence is presumed to have at least two sugar chains.

The paragraph beginning at the bottom of page 26, line 14, has been amended as follows:

The vector is not specifically limited in so far as it can express the protein of the present invention. Examples thereof include pBAD/His, pRSETA, pcDNA2.1, pTrcHis2A, pYES2, pBlueBac4.5, pcDNA3.1 and pSecTag2 manufacture by Invitrogen, pET and pBAC manufactured by Novagen, pGEM manufactured by Promega, pBluescriptII manufactured by Stratagene, pGEX and pUC18/19 manufactured by Pharmacia, PfastBAC1 manufactured by GIBCO and the

like. Preferably, a protein expression vector (described in the specification of a patent application entitled "Protein expression vector and its use" and filed by the same applicant on the same day) is used. This expression vector is constructed by using pCRII-TOPO vector described in the Examples hereinafter, or a commercially available expression vector, for example pSecTag2A vector or pSecTag2B vector (Invitrogen) and integrating a secretory signal nucleotide sequence suitable for expression of the protein of the present invention, in the 3' downstream side thereof, a Tag nucleotide sequence, a cleavable nucleotide sequence and a cloning site, into which a nucleotide sequence encoding a target protein can be inserted, in this order. More specifically, it is preferred to use trypsin signal as the secretory signal, a nucleotide sequence encoding polyhistidine as the Tag nucleotide sequence, and a nucleotide sequence encoding an amino acid sequence which is susceptible to enzyme-specific cleavage, i.e., a nucleotide sequence encoding the amino acid sequence of Asp-Asp-Asp-Lys SEQ ID NO: 42 (said amino acid sequence is recognized by enterokinase, and the recombinant fusion protein is cleaved at the C-terminus part thereof) as the cleavable nucleotide sequence.

The paragraph beginning at page 49, line 22, has been amended as follows:

The cloning was carried out by PCR using a mouse brain cDNA library (Clontech) as a template and nucleotide sequences corresponding to an amino acid sequence common to serine proteases represented by

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is shown in SEQ ID NO: 7 and the amino acid sequence of mSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 8. Further, two different types of clones were obtained. The nucleotide sequences of these DNA are shown in SEQ ID NOS: 3 and 5, respectively. The amino acid sequences of mBSSP2 proteins deduced from these nucleotide sequences are shown in SEQ ID NOS: 4 and 6. These novel proteases are divided into types 1, 2 and 3. Type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids and type 3 is composed of 445 amino acids. These amino acid sequences contained the common amino acid sequence composed of 238 amino acids whose N-terminus side started with Ile-Val-Gly-Gly-Gln-Ala-Val (amino acid 1-7 of SEQ ID NO:2) as the mature serine protease. Further, in the amino acid sequence of the mature serine protease, a consensus sequence having a serine protease activity was contained. Furthermore, since there were two or more amino acid sequences specific for a sugar chain bonding site, it was presumed that the amino acid sequence had at least two sugar chains.

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C-terminus of the part from trypsin signal to the enterokinase recognition site of pSecTrypHis/neurosin. This was inserted between NheI and HindIII sites of pSecTag2A to construct the plasmid pTrypSig.

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plasmid was extracted from each colony formed according to a conventional manner and its nucleotide sequence was determined by dideoxy method. As for a clone having homology to mBSSP2, full length cDNA was obtained by 5' RACE and 3' RACE and its sequence was determined as described above. PCR was carried out by using the above cDNA as a template and primers having the sequences represented by SEQ ID NOS: 30 and 37. 3' RACE was carried out by PCR using a 1/100 dilution of the above PCR product as a template and primers having the sequences represented by SEQ ID NOS: 32 and 37. As for 5' RACE, cDNA for RACE was prepared from human fetal brain mRNA (Clontech) by using Superscript II and SMART RACE cDNA amplification kit (Clontech). PCR of this cDNA was carried out by using a primer of 10 x Universal Primer Mix (attached to the kit) and a primer having the sequence represented by SEQ ID NO: 33. Further, PCR was carried out by using the 1/100 dilution of the latter PCR product, a template, Nested PCR Primer (attached to the kit) and a primer having the sequence represented by SEQ ID NO: The finally obtained PCR product was subjected to TA cloning 34. as described above and the nucleotide sequence was determined to obtain the upstream and downstream regions of the above clone. In addition, primers for amplifying the full length cDNA as represented by SEQ ID NOS: 35 and 36 were prepared based on the resultant nucleotide sequence and PCR was carried out by using the above synthetic cDNA as a template. This PCR product was cloned into pGEM-T Easy vector to obtain the plasmid pGEM-TE/hBSSP2

containing the full length cDNA clone. The DNA sequence contained in this plasmid is shown in SEQ ID NO: 9 and hBSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 10.

## SEQUENCE LISTING

<110> UEMURA, Hidetoshi OKUI, Akira KOMINAMI, Katsuya YAMAGUCHI, Nozomi MITSUI, Shinichi	
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Pro	His	Trp 35	Val	Val	Thr	Ala	Ala 40	His	Cys	Met	Tyr	Ser 45	Phe	Arg	Leu	
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Ala 65	Val	Arg	Gln	His	Gln 70	Gly	Thr	Met	Val	Glu 75	Lys	Ile	: Ile	Pro	His 80	
Pro	Leu	Tyr	Ser	Ala	Gln	Asn	His	Asp	Tyr	Asp	val	Ala	Leu	Leu	Gln	

				85					90					95		
Leu	Arg	Thr	Pro 100	Ile	Asn	Phe	Ser	Asp 105	Thr	Val	Asp	Ala	Val 110	Cys	Leu	
Pro	Ala	Lys 115	Glu	Gln	Tyr	Phe	Pro 120	Trp	Gly	Ser	Gln	Cys 125	Trp	Val	Ser	
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Val	Cys	Pro 195	Ser	Gly	Asp	Thr	Trp 200	His	Leu	Val	Gly	Val 205	Val	Ser	Trp	
Gly	Arg 210	Gly	Cys	Ala	Glu	Pro 215	Asn	Arg	Pro	Gly	Val 220	Tyr	Ala	Lys	Val	
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In re Application of: ) Art Unit:

H. UEMURA, et al. ) Examiner:

Serial No.: 09/856,371 ) Confirmation No.

Filed: May 21, 2001 ) Washington D.C.

For: NOVEL SERINE PROTEASE ) August 14, 2001

BSSP2

### PRELIMINARY AMENDMENT

Honorable Commissioner for Patents Washington, D.C. 20231

Prior to examination of the above-identified application, please enter the following Preliminary Amendment:

#### IN THE SPECIFICATION

Page 1, please amend the second paragraph as follows:

In general, proteases are biosynthesized as inactive precursors. They undergo limited hydrolysis in molecules to be converted into activated type proteases. In so far as enzymes are proteases, they have an activity for hydrolyzing a peptide bond, while their actions vary according to kinds of proteases. According to a particular kind of catalytic site, proteases are divided into serine proteases, cysteine proteases, aspartate proteases, metal proteases and the like. Proteases of each kind have a variety of properties, ranging from a protease having general digestive properties to a protease having various regulatory domains and strict substrate specificity, thereby specifically hydrolyzing

only characteristic proteins.

Page 2, please amend the paragraph on page 2 as follows:

Further, proteins undergo various types of processing even after translation to produce active proteins. In many secretory proteins, a protein is first synthesized on the ribosome in cytoplasm as an inactive precursor (pro-form) which comprises an active protein bearing at the N-terminus thereof a peptide of about 15 to 60 amino acids responsible for secretion (secretory signal). This peptide region is concerned with the mechanism for passing through the cell membrane and is removed upon cleavage by a specific protease during the passage through the membrane, in almost all the cases, to produce the mature form. A secretory signal has a broad hydrophobic region comprising hydrophobic amino acids in the middle of the sequence, and basic amino acid residues at a site close to the N-terminus. A secretory signal is a synonym for a signal peptide. In addition, in some proteins, a peptide moiety which functions as a secretory signal is further attached to the N-terminus of the inactive precursor (pro-form). Such a protein is called a prepro-protein (prepro-form).

Page 3, please amend the first paragraph as follows:

For example, trypsin is present in the form of a prepro-form immediately after translation into amino acids.

After being secreted from cells, it is present in the form of a pro-form and is then converted into active trypsin in the duodenum upon limited hydrolysis by enteropeptidase or by trypsin itself.

Page 3, please amend the second paragraph as follows:

The optimal pH range of serine proteases is neutral to weak alkaline and, in general, many of them have a molecular weight of about 30,000 or lower. All proteases relating to blood coagulation, fibrinolysis and complement systems having a large molecular weight belong to the family of trypsin-like serine proteases. They have many regulator domains and form a protease cascade which is of very importance to reactions in a living body.

Page 4, please amend the first paragraph as follows:

Serine proteases expressed in a brain-nerve system

such as neurosin are considered to play various roles in the

brain-nerve system. Therefore, there is a possibility that

isolation of a gene encoding a novel protease expressed in a

brain-nerve system and production of a protein using the gene

would be useful for diagnosis or therapy of various diseases related to the brain-nerve system.

Page 4, please amend the second paragraph as follows:

Nowadays, in general, clinical diagnosis of Alzheimer's disease is based on the diagnosis standard of DSM-IIIR and NINCDS-ADRDA (Mckhann, G. et al., Neurology, 34. 939, 1994) or the diagnosis standard of DSM-IV (American Psychiatric Association; Diagnostic and statistical manuals of mental disorders, 4th ed., Washington DC, American Psychiatric Association, 1994). However, these standards are conditioned by a decline in recognition functions which causes a severe disability in daily life or social life. Then, it is pointed out that the diagnosis is less than scientifically objective because the diagnosis may be influenced by the level of an individual's social life and further the specialty and experience of a physician who diagnoses particular conditions. In addition, definite diagnosis of Alzheimer's disease is conducted by pathohistological analyses and, in this respect, substantial inconsistency between clinical diagnosis and autopsy diagnosis exists.

Page 5, please amend the paragraph on page 5 as follows:

At present, image diagnosis is employed as a supplemental means in clinical diagnosis of Alzheimer's diagnosis and it is possible to analyze brain functions, for example, decline of metabolism and atrophy in specific sites such as hippocampus, parietal lobe of cerebral cortex and the like which are specific for Alzheimer's disease by PET and SPECT. However, to define Alzheimer's disease based on lowering of a blood flow from parietal lobe to temporal lobe is very dangerous. In addition, there is a report showing that MRS test is useful for patients with dementia including those of Alzheimer's disease. Further, although CT-MRI image diagnosis is used, a lesion of white matter such as atrophy of brain, PVL or the like is not specific for Alzheimer type dementia. Since it has been reported that atrophy of brain proceeds with aging, the above observation is not necessarily found in Alzheimer type dementia. Furthermore, since an image obtained by MRI varies according to strength of a magnetic field, performance of the apparatus and imaging conditions, numerical data obtained in different facilities cannot be compared with each other except for atrophic change. addition, there is a limit to image measurement. Further, enlargement of the ventricle can be recognized in vascular dementia cases and there are cases wherein atrophy of the

hippocampus is observed after ischemia of the basilar artery.

Page 6, please amend the fourth paragraph as follows:

Further, data obtained in different facilities can be compared with each other by using the same diagnosis marker. Therefore, development of biological diagnosis markers is recognized to be a most important field among fields of Alzheimer's disease studies and its future prospects will be expected. Approaches to development of biological diagnosis markers up to now are divided into those based on constitute components of characteristic pathological changes of Alzheimer's disease such as senile plaque and neurofibril change, and an approach based on other measures. Examples of the former include cerebrospinal fluid tau protein,  $A\beta$  and its precursor,  $\beta$ APP. Examples of the latter include mydriasis test with cholilytic drug, Apo E and other genes relating to Alzheimer's disease. However, no good results have been obtained.

Page 7, please amend the paragraph on page 7 as follows:

Serine proteases are also considered to play an important role in cancer cells. The reason why extermination of cancer by surgical treatment or topical irradiation of

radioactive ray is difficult is the metastatic capability of cancer. To spread solid tumor cells in a body, they loosen their adhesion to original adjacent cells, followed by separating from original tissue, passing through other tissues to reach the blood vessels or lymph nodes, entering into the circulatory system through stratum basal and endothelial layer of the vessel, leave from the circulatory system at somewhere in the body, and surviving and proliferating in a new environment. While adhesion to adjacent epidermal cells is lost when expression of cadherin which is an intercellular adhesive molecule of epithelium is stopped, to break through tissues is considered to depend on proteolytic enzymes which decompose an extracellular matrix.

Page 8, please amend the first paragraph as follows:

As enzymes which decompose the matrix, mainly, metal proteases (Rha, S. Y. et al., Breast Cancer Research

Treatment, 43, 175, 1997) and serine proteases are known.

They cooperate to decompose matrix proteins such as collagen, laminin and fibronectin. Among the serine proteases known to be concerned in decomposition of the matrix, in particular, there is urokinase type plasminogen activator (U-PA). U-PA has a role as a trigger specific for a protein decomposition chain reaction. Its direct target is plasminogen. It is present in blood abundantly and is a precursor of an inactive

serine protease which accumulates in reconstructed sites of tissues such as injured sites and tumors as well as inflammatory sites. In addition, as proteases which are concerned in metastasis and infiltration of cancers, for example, a tissue factor, lysosomal type hydrolase and collagenase have been known.

Page 8, please amend the second paragraph as follows:

At present, cancer is the top cause of death in Japan and more than 200,000 people die per year. Accordingly, specific substances which can be used as markers for diagnosis and therapy or prophylaxis of cancer are studied intensively. Such specific substances are referred to as tumor markers or tumor marker relating biomarkers. They are utilized in aid of diagnosis before treatment of cancer, for presuming carcinogenic organ and pathological tissue type, for monitoring effect of treatment, for finding recurrence early, for presuming prognosis, and the like. At present, tumor markers are essential in clinical analyses. Among them, alpha fetoprotein (AFP) which has high specificity to hepatocellular carcinoma and yolk sac tumor (Taketa K. et al., Tumour Biol., 9, 110, 1988), and carcinoembronic antigen (CEA) are used worldwide. In the future, tumor markers will be required more and more, and it is desired to develop, for example, organ

specific markers and tumor cell specific markers which are highly reliable serologic diagnosis of cancer. Up to now, humunglandular kallikrein (hK2) which is a serine protease expressed at human prostatic epithelial cells has been reported as a marker for prostatic cancer. And, hK2 has 78% homology with the sequence of prostatic specific antigen (PSA) and PSA is also used widely as a biochemical marker of prostatic cancer (Mikolajczyk, S. d. et al., Prostate, 34, 44, 1998; Pannek, J. et al., Oncology, 11, 1273, 1997; Chu, T. M. et al., Tumour Biology, 18, 123, 1997; Hsieh, M. et al., Cancer Res., 57, 2651, 1997). Further, hK2 is reported to be useful as a marker for not only prostatic cancer but also stomach cancer (Cho, J. Y. et al.. Cancer, 79, 878, 1997). Moreover, CYFRA (CYFRA 211) for measuring cytokeratin 19 fragment in serum is reported to be useful for lung cancer (Sugiyama, Y. et al., Japan J. Cancer Res., 85, 1178, 1994). Gastrin release peptide precursor (ProGRP) is reported to be useful as a tumor marker (Yamaguchi, K. et al., Japan, J. Cancer Res., 86, 698, 1995).

Page 10, please amend the second paragraph as follows:

Under these circumstances, the present inventors have succeeded in cloning cDNA encoding novel human and mouse serine proteases.

Page 10, please amend the third paragraph as follows:

In summary, the 1st feature of the present invention is the amino acid sequences of biologically active mature serine proteases BSSP2 and nucleotide sequences encoding the amino acid sequences.

Page 18, please amend the second paragraph as follows:

In case of northern blotting analysis, mBSSP2 shows the expression in the head of a 15-20 days mouse fetus, and in the lung, prostate and testicle of a 3 month-old mouse. hBSSP2 shows the expression in brain, skeletal muscle and liver (see Figs. 1, 2 and 5). In case of RT-PCR analysis, mBSSP2 shows the expression in the brain and testicle of a 12 day-old mouse, and hBSSP2 shows the expression in the brain and skeletal muscle. Then, the novel proteases of the present invention are presumed to play various roles in the brain, prostate, lung, testicle, skeletal muscle and liver. For example, in the brain, there is a possibility that they can be used for treatment and diagnosis of brain diseases such as Alzheimer's disease (AD), epilepsy, brain tumor and the like. Further, in other tissues, there is a possibility that BSSP2 of the present invention and a gene encoding it can be used for treatment and diagnosis of various diseases such as

cancer, inflammation, infertility, prostatomegaly and the like. Further, it is presumed they may have a certain influence on blood coagulation, fibrinolysis and complement systems. Furthermore, there is a possibility that inhibitors of serine proteases can be used for treatment and diagnosis of Alzheimer's disease, epilepsy, cancer, inflammation, infertility, prostatomegaly and the like.

Page 22, please amend the paragraph on page 22 as follows:

In general, many genes of eucaryotes exhibit polymorphism and, sometimes, one or more amino acids are substituted by this phenomenon. Further, even in such a case, sometimes, a protein maintains its activity. Then, the present invention includes a gene encoding a protein obtained by modifying a gene encoding any one of the amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10, artificially, in so far as the protein has the characteristic function of the gene of the present invention. Further, the present invention includes a protein which is a modification of any one of amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10 in so far as the protein has the characteristics of the present invention. Modification is understood to include substitution, deletion, addition and/or insertion. In particular, the present inventors have shown

that, even when several amino acids are added to or deleted from the N-terminus amino acid of the BSSP2 mature protein represented by SEQ ID NO: 2, the resultant sequence maintains its activity.

Page 23, please amend the first paragraph as follows:

That is, the present invention includes a protein comprising any one of the amino acid sequences described in SEQ ID NOS: 2, 4, 6, 8 and 10; an amino acid sequence encoded 5 by any one of the nucleotide sequences represented by SEQ ID NOS: 1, 3, 5, 7 and 9; or one of these amino acid sequences wherein one to several amino acids have been substituted, deleted, added and/or inserted, and belonging to serine protease family.

Page 23, please amend the second paragraph as follows:

Each codon for the desired amino acid itself has been known and can be selected freely. For example, codons can be determined according to a conventional manner by taking into consideration the frequency of use of codons in a host to be utilized (Grantham, R. et al., Nucleic Acids Res., 9, r43, 1989). Therefore, the present invention also includes a nucleotide sequence appropriately modified by

taking into consideration the degeneracy of a codon. Further, these nucleotide sequences can be modified by a site directed mutagenesis using a primer composed of a synthetic oligonucleotide encoding the desired modification (Mark, D. F. et al., Proc. Natl. Acad. Sci. USA., 81, 5662, 1984), or the like.

Page 23, please amend the third paragraph as follows:

Furthermore, the DNA of the present invention includes DNA which is hybridizable to any one of the nucleotide sequences described in SEQ ID NOS: 1, 3, 5, 7 and 9 or nucleotide sequences complementary to these nucleotide sequences in so far as the protein encoded by the nucleotide sequence has the same properties as those of the BSSP2 of the present invention. It is considered that many of the sequences which are hybridizable to a given sequence under stringent conditions have a similar activity to that of a protein encoded by the given sequence. The stringent conditions according to the present invention includes, for example, incubation in a solution containing 5 x SSC, 5% Denhardt's solution (0.1% BSA, 0.1% Ficol 1400, 0.1% PVP), 0.5% SDS and 20 µg/ml denatured salmon sperm DNA at 37°C overnight, followed by washing with 2 x SSC containing 0.1% SDS at room temperature. Instead of SSC, SSPE can be

In re Appl. No. 09/856,371 appropriately used.

Page 25, please amend the second paragraph as follows:

The present invention also relates to a vector comprising the nucleotide sequence represented by SEQ ID NO: 1 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 2; the nucleotide sequence represented by SEQ ID NO: 3 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 4; the nucleotide sequence represented by SEQ ID NO: 5 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 6; the nucleotide sequence represented by SEQ ID NO: 7 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 8; or the nucleotide sequence represented by SEQ ID NO: 9 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 10; or a nucleotide sequence similar to them. A nucleotide sequence similar to a given nucleotide sequence used herein means a nucleotide sequence which is hybridizable to the given nucleotide sequence or its complementary nucleotide sequence under the above-described stringent conditions and which encodes a protein having the same properties as those of the protein encoded by the nucleotide sequence.

Page 28, please amend the first paragraph as follows:

The animal cells and insect cells used herein include cells derived from human beings and cells derived from flies or silk worms. For example there are CHO cells, COS cells, BHK cells, Vero cells, myeloma cells, HEK293 cells, HeLa cells, Jurkat cells, mouse L cells, mouse C127 cells, 10 mouse FM3A cells, mouse fibroblast, osteablast, cartilage cells, S2, Sf9, Sf21, High Five™ (registered trade mark) cells and the like.

Page 28, please amend the second paragraph as follows:

The protein of the present invention as such can be expressed as a recombinant fused protein so as to facilitate isolation, purification and recognition. The recombinant fused protein used herein means a protein expressed as an adduct wherein a suitable peptide chain is added to the N-terminus and/or C-terminus of the desired protein expressed by a nucleotide sequence encoding the desired protein. The recombinant protein used herein means that obtained by integrating a nucleotide sequence encoding the desired protein in the expression vector of the present invention and cut off an amino acid sequence which derived from nucleic acids other than those encoding the desired protein from the expressed recombinant

fused protein, and is substantially the same as the protein of the present invention.

Page 29, please amend the first paragraph as follows:

Introduction of the above vector into host cells can be carried out by, for example, transfection according to the lipopolyamine method, DEAE-dextran method, Hanahan method, lipofectin method or calcium phosphate method, microinjection, eletroporation and the like.

Page 33, please amend the first paragraph as follows:

For obtaining fertilized egg cells efficiently, ovulation may be induced with gonadotropin or the like.

Fertilized egg cells are recovered and a gene in an injection pipette is injected into male pronucleus of the egg cells by microinjection. For returning the injected egg cells to a fallopian tube, an animal (false pregnancy female mouse, etc.) is provided and about 10 to 15 eggs/mice are transplanted. Then, genomic DNA is extracted from the end part of the tail to confirm whether the transgene is introduced into newborn mouse or not. This confirmation can be carried out by detection of the transgene with southern blot technique or PCR technique, or by positive cloning

wherein a marker gene, which is activated only when homologous recombination is caused, has been introduced. Further, transcribed products derived from the transgene are detected by northern blot technique or RT-PCR technique to confirm expression of the transgene. Or, western blotting can be carried out with a specific antibody to a protein.

Page 33, please amend the second paragraph as follows:

The knockout mouse of the present invention is treated so that the function of mBSSP2 gene is lost. A knockout mouse means a transgenic mouse in which any one of its genes is destroyed by homologous recombination technique so that its function is deficient. A knockout mouse can be created by carrying out homologous recombination with ES cells and selecting embryonic stem cells wherein either of allele genes are modified or destroyed. For example, embryonic stem cells whose genes are manipulated at the blastocyte or morula stage of fertilized eggs are injected to obtain a chimeric mouse wherein cells derived from the embryonic stem cells are mixed with those derived from The chimeric mouse (chimeric means a single the embryo. individual formed by somatic cells based on two or more fertilized eggs) can be mated with a normal mouse to create a heterozygote mouse wherein all of the allele genes have been

modified or destroyed. Further, a homozygote mouse can be created by mating heterozygote mice.

Page 37, please amend the paragraph on page 37 as follows:

Examples of myeloma cells include X-63Ag8, NS-1, P3U1, SP2/0, AP-1 and the like with SP2/0 being preferred. preferred ratio of the number of the antibody producer cells (spleen cells) : the number of spleen cells are 1 : 20 to 20 : PEG (preferably PEG 1000 to PEG 6000) is added at a concentration of about 10 to 80% and the mixture is incubated at 20 to  $40\,^{\circ}\text{C}$ , preferably 30 to  $37\,^{\circ}\text{C}$  for 1 to 10 minutes to carry out the cell fusion efficiently. Screening of anti-hBSSP2 or mBSSP2 antibody producer hybridomas can be carried out by various For example, a supernatant of a hybridoma culture is methods. added to a solid phase to which hBSSP2 or mBSSP2 antigen is adsorbed directly or together with a carrier (e.g., microplate), followed by addition of an anti-immunoglobulin antibody (in case that the cells used in cell fusion are those of a mouse, antimouse immunoglobulin antibody is used) or protein A to detect the anti-hBSSP2 or mBSSP2 monoclonal antibody attached to the solid phase. Or, a supernatant of a hybridoma culture is added to a solid phase to which an anti-immunoglobulin antibody or protein A is adsorbed, followed by addition of hBSSP2 or mBSSP2 labeled with a radioactive substance, an enzyme, etc., to detect the

anti-hBSSP2 or mBSSP2 monoclonal antibody attached to the solid phase.

Page 38, please amend the first paragraph as follows: Selection and cloning of the anti-hBSSP or mBSSP monoclonal antibody can be carried out according to a per se known method or its modification. Normally, a HAT (hypoxanthine, aminopterin, thymidine)-added medium for culturing animal cells is used. Any culture medium can be used for selection, cloning and growing up in so far as the hybridoma can grow. For example, there can be used RPMI culture medium containing 1 to 20%, preferably 10 to 20% fetal bovine serum, a serum-free medium for culturing hybridomas. Preferably, the culture is carried out at a temperature of about 37°C. Normally, the culture time is 5 days to 3 weeks, preferably 1 week to 2 weeks. Normally, the culture is carried out under 5%  $CO_2$ . The antibody titer of a supernatant of a hybridoma culture can be measured according to the same manner as that of the above-described measurement of anti-BSSP2 antibody titer in an antiserum. That is, examples of the measurement to be used include radioimmunoassay (RIA), enzyme-linked immunosorbent assay (ELISA), FIA (fluorescence immunoassay), plaque assay, agglutination reaction method, and the like. Among them, ELISA as shown below is preferred.

Page 39, please amend the paragraph on page 39 as follows:

Normally, cloning is carried out by a per se known method such as semi-solid agar method, limiting dilution method and the like. Specifically, after confirming a well in which the desired antibody is produced by the above-described method, cloning is carried out to obtain a single clone. For cloning, it is preferred to employ limiting dilution method wherein hybridoma cells are diluted so that one colony is formed per one well of a culture plate. For cloning by limiting dilution method, feeder cells can be used, or a cell growth factor such as interleukin 6, etc. can be added to improve colony forming capability. In addition, cloning can be carried out by using FACS and single cell manipulation method. The cloned hybridoma is preferably cultured in a serum-free culture medium and an optimal amount of an antibody is added to its supernatant. The single hybridoma thus obtained can be cultured in a large amount by using a flask or a cell culture device, or cultured in the abdominal cavity of an animal (J. Immunol. Meth., 53, 313, 1982) to obtain a monoclonal antibody. When culturing in a flask, there can be used a cell culture medium (e.g., IMDM, DMEM, RPMI1640, etc.) containing 0 to 20% of FCS. When culturing in the abdominal cavity of an animal, the animal to be used is preferably the same species or the same line as that from which the myeloma cells used in the cell fusion are derived, a thymus

deficient nude mouse or the like, and the hybridoma is transplanted after administration of a mineral oil such as pristane, etc. After 1 to 2 weeks, myeloma cells are proliferated in the abdominal cavity to obtain ascites containing a monoclonal antibody.

Page 42, please amend the paragraph on page 42 as follows:

The polyclonal antibody of the present invention can be produced according to a per se known method or modification. For example, an immunogen (protein antigen) per se or a complex thereof with a carrier protein is prepared and, according to the same manner as that in the above monoclonal antibody production, a warm-blooded animal is immunized. material containing an antibody against the protein of the present invention or its fragment is collected from the immunized animal and the antibody is separated and purified to obtain the desired antibody. As for a complex of an immunogen and a carrier protein for immunizing a warm-blooded animal, the kind of a carrier protein and the mixing ratio of a carrier and a hapten are not specifically limited in so far as an antibody against hapten immunized by cross-linking with the carrier is efficiently For example, there can be used about 0.1 to 20, preferably about 1 to 5 parts by weight of bovine serum albumin, bovine cycloglobulin, hemocyanin, etc. coupled with one part by

weight of a hapten. For coupling a carrier and a hapten, various condensing agents can be used. Examples thereof include glutaraldehyde, carbodiimide or maleimide active ester, active ester agents having thiol group or dithiopyridyl group, and the like. The condensed product is administered as such or together with a carrier or diluent to a site of a warm-blooded animal where an antibody can be produced. For enhancing the antibody production, upon administration, Freund's complete adjuvant or Freund's incomplete adjuvant may be administered. Normally, the administration is carried out once every 2 to 6 weeks, 3 to 10 The polyclonal antibody can be collected from times in all. blood, ascites, or the like, preferably blood of the immunized The polyclonal antibody titer in an antiserum can be measured according to the same manner as measurement of the above monoclonal antibody titer in the antiserum. Separation and purification of the polyclonal antibody, like monoclonal antibody, can be carried out according to the same manner as those of immunoglobulins.

Page 44, please amend the second paragraph as follows:

As a sandwich method for determining hBSSP2 or mBSSP2
or a fragment thereof, there can be used a two step method, a one
step method and the like. In the two step method, first, the
immobilized antibody is reacted with hBSSP2 or mBSSP2 or a
fragment thereof and then unreacted materials are completely

removed by washing, followed by addition of the labeled antibody to form immobilized antibody-hBSSP2 or mBSSP2-labeled antibody. In the one step method, the immobilized antibody, labeled antibody and hBSSP2 or mBSSP2 or a fragment thereof are added at the same time.

Page 45, please amend the second paragraph as follows: For immobilization of the antibody, a known chemical bonding method or a physical adsorption can be used. Examples method chemical bonding method include а glutaraldehyde; maleimide method using N-succinimidyl-4-(Nmaleimidomethyl)cyclohexane-1-carboxylate, N-succinimdyl-2maleimide acetate or the like; carbodiimide method using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride; or the like. In addition, there are maleimidobenzoyl-N-hydroxysuccinimide ester method, N-succinimidyl-3-(2-pyridylthio)propionic acid method, bisdiazobenzidine method, and dipalmityllysine method. Or, it is possible to capture a complex formed beforehand by reacting a material to be tested with two antibodies, whose epitopes are different, with an immobilized a 3rd antibody against the antibody.

Page 46, please amend the paragraph on page 46 as follows:

For labeling, it is preferred to use an enzyme, fluorescent substance, luminous substance, radioactive substance, metal chelate, or the like. Examples of the enzyme include peroxidase, alkaline phosphatase,  $\beta$ -D-galactosidase, malate dehydrogenase, Staphylococcus nuclease,  $\delta$ -5-steroidisomerase,  $\alpha$ glycerol phosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, asparaginase, glucose oxidase, urease, catalase, glucose-6-phosphate ribonuclease, dehydrogenase, glucoamylase, acetylcholinesterase and the like. Examples of the fluorescent substance include fluorescein isothiocyanate, phycobiliprotein, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthalaldehyde, and the like. Examples of the luminous substance include isoluminol, lucigenin, luminol, aromatic acridinium ester, imidazole, acrdinium salt and its modified ester, luciferin, luciferase, aequorin and the like. Examples of the radioactive substance include 125I, 127I, 131I, 14C,  3 H,  32 P,  35 S and the like. The labeling material is not limited to them and any material which can be used for immunological determination can be used. Further, a low molecular weight hapten such as biotin, dinitrophenyl, pyridoxal or fluorescamine may be attached to the antibody. Preferably, horseradish peroxidase is used as a labeling enzyme. This enzyme can be reacted with various substrates and can readily be attached to

the antibody by periodate method.

Page 47, please amend the first paragraph as follows: When an enzyme is used as a labeling material, a substrate and, if necessary, a coloring enzyme is used for measuring its activity. In case of using peroxidase as the enzyme,  $H_2O_2$  is used as a substrate and, as a coloring agent, there can be used 2,2'-azino-di-[3-ethylbenzthiazoline sulfonic acidl ammonium salt (ABTS), 5'-aminosalicylic acid, ophenylenediamine, 4-aminoantipyrine, 3,3',5,5'tetramethylbenzidine and the like. In case of using alkaline phosphatase as the enzyme, o-nitrophenylphosphate, pnitrophenylphosphoric acid, or the like can be used as a substrate. In case of using  $\beta\text{-D-galactosidase}$  as the enzyme, fluorescein-d-( $\beta$ -D-galactopyranoside), 4-methylumbelliphenyl- $\beta$ -Dgalactopyranoside, or the like can be used as a substrate. present invention also includes a kit comprising the above monoclonal antibody, polyclonal antibody and reagents.

Page 47, please amend the second paragraph as follows:

As a cross-linking agent, a known cross-linking agent such as N,N'-o-phenylenedimaleimide, 4-(N-maleimidomethyl)cyclohexanoate-N-succinimide ester, 6-maleimidohexanoate-N-succineimide ester, 4,4'-dithiopyridine or the like can be utilized. The reaction of these cross-linking

agents with enzymes and antibodies can be carried out by a known method according to properties of a particular cross-linking agent. Further, as the antibody, a fragment thereof, for example, Fab', Fab, F(b'2) can be used as the case may be. A labeled enzyme can be obtained by the same treatment regardless of whether the antibody is polyclonal or monoclonal. When the above labeled enzyme obtained by using a cross-linking agent is purified by a known method such as affinity chromatography or the like, an immunoassay system having more higher sensitivity can be obtained. The enzyme labeled and purified antibody is stored in a dark cold place with addition of a stabilizer such as thimerosal, glycerin or after lyophilization.

Page 49, please amend the paragraph on page 49 as follows:

The cloning was carried out by PCR using a mouse brain cDNA library (Clontech) as a template and nucleotide sequences corresponding to an amino acid sequence common to serine proteases represented by

Primer 1: GTG CTC ACN GCN GCB CAY TG (SEO ID NO: 20)

Primer 2: CCV CTR WSD CCN CCN GGC GA (SEQ ID NO: 21)

as primers. Namely, 5  $\mu$ l of the template, 5  $\mu$ l of 10 x ExTaq buffer, 5  $\mu$ l of dNTP, 10 pmol of each of the above primers and 0.5  $\mu$ l of ExTaq (TAKARA) were added and the total volume was adjusted to 50  $\mu$ l with sterilized water. PCR was carried out by

repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for 1 minute, 30 times. product was mixed with pCR II-TOPO vector attached to TOPO TA cloning kit (Invitrogen) and the mixture was allowed to stand at temperature for 5 minutes. Then, according to conventional manner, E. coli Top 10 attached to the kit was transformed and applied to a LB (Amp+) plate (containing 100 ug/ml of ampicillin). According to a conventional manner, a plasmid was extracted from each colony obtained and its nucleotide sequence was determined by cycle sequencing method with a fluorescence sequencer (ABI). Homology of the sequence of each clone was examined by means of GenBank. Regarding an unknown sequence, i.e., BSSP2 gene, the full length cDNA was obtained by 5' RACE and 3' RACE and, according to the same manner as described above, the nucleotide sequence was determined. Namely, BSSP2 clone specific primers, GSP1 primers [mBSSP2.2 (SEQ ID NO: 27) or mBSSP2.0 (SEQ ID NO: 22)] and GSP2 primers [mBSSP2R2 (SEQ ID NO: 28) or mBSSP2.1 (SEQ ID NO: 23)] were PCR was carried out by using mouse brain Marathon-Ready cDNA (Clontech), AP1 primer attached to this reagent and either of the above GSP1 primers and heating at 94°C for 2 minutes once and repeating a cycle of heating at 94°C for 30 seconds, at 60°C for 30 seconds and then at 72°C for 30 seconds 35 times. Then, 5  $\mu$ l of the PCR product diluted to 1/100, 5  $\mu$ l of 10 x buffer, 5 µl of dNTP, 10 pmol of either of 10 µM of the

above GSP2 primer, 10 pmol of AP2 primer attached to the above reagent and 0.5 unit of ExTaq were admixed and adjusted to 50 µl with sterilized water. Then, according to the same manner as the above, PCR was carried out. The PCR product was cloned by the above TOPO TA cloning kit and sequenced to obtain the upstream and downstream regions of the above clone. At this time, as for a clone which seemed not to cover the full length of a protein, the specific primers shown hereinafter were prepared based on the newly found nucleotide sequence. Further, based on this sequence, the primers capable of amplifying ORF as hereinafter [mBSSPF7 (SEQ ID NO: 26), mBSSP2R/E (SEQ ID NO: 29)] were prepared and PCR carried out using mouse brain Marathonready cDNA as a template to confirm that these clones were identical. This was cloned into pCR II-TOPO vector attached to TOPO TA cloning kit to obtain the plasmid pCR II/mBSSP2 containing the full length cDNA clone. The nucleotide sequence of DNA contained in this plasmid is shown in SEQ ID NO: 7 and the amino acid sequence of mSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 8. Further, two different types of clones were obtained. The nucleotide sequences of these DNA are shown in SEQ ID NOS: 3 and 5, respectively. The amino acid sequences of mBSSP2 proteins deduced from these nucleotide sequences are shown in SEQ ID NOS: 4 and 6. These novel proteases are divided into types 1, 2 and 3. Type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids and

type 3 is composed of 445 amino acids. These amino acid sequences contained the common amino acid sequence composed of 238 amino acids whose N-terminus side started with Ile-Val-Gly-Gly-Gln-Ala-Val as the mature serine protease. Further, in the amino acid sequence of the mature serine protease, a consensus sequence having a serine protease activity was contained. Furthermore, since there were two or more amino acid sequences specific for a sugar chain bonding site, it was presumed that the amino acid sequence had at least two sugar chains.

Page 53, please amend the paragraph on page 53 as follows:

As seen from Figs. 1 and 2, in the case of northern blotting analysis, the expression of mBSSP2 was recognized in the head of 15th to 20th day fetuses of mice and, as to the 3-month-old mice, the expression was recognized in the prostate and testicle. Further, according to the results of RT-PCR, the expression was recognized in the head of 12-day-old mice and the testicle of 3-month-old mice.

## IN THE CLAIMS

Cancel claims 1-19 and 37-40 and enter the following new claims:

41. (New) A protein selected from the group consisting of:

- (a) a protein having the amino acid sequence of 238 amino acids represented by SEQ ID NO: 2;
- (b) a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 2 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 2;
- (c) a protein having the amino acid sequence of 273 amino acids represented by SEQ ID NO: 4;
- (d) a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 4 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 4;
- (e) a protein having the amino acid sequence of 311 amino acids represented by SEQ ID NO: 6;
- (f) a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 6 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 6;
- (g) a protein having the amino acid sequence of 455 amino acids represented by SEQ ID NO: 8;
- (h) a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 8 by

deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 8;

- (i) a protein having the amino acid sequence of 240 amino acids represented by the 1st to 240th amino acids of SEQ ID NO: 10;
- (j) a protein having an amino acid sequence derived from the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10;
- (k) a protein having the amino acid sequence of 457 amino acids represented by the -217th to 240th amino acids of SEQ ID NO: 10;
- (1) a protein having an amino acid sequence derived from the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10;
- (m) a protein having the amino acid sequence of 217 amino acids represented by the -217th to -1st amino acids of SEQ ID NO: 10;

- (n) a protein having an amino acid sequence derived from the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10; and
- (o) a modified derivative or fragment of these proteins (a) to (n).
- 42. (New) A nucleotide sequence selected from the group consisting of:
- (aa) a nucleotide sequence represented by the 1st to 714th bases of SEQ ID NO: 1;
- (bb) a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 2;
- (cc) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (aa) or (bb) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 2;
- (dd) a nucleotide sequence represented by the 247th to 1065th bases of SEQ ID NO: 3;

- (ee) a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 4;
- (ff) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (dd) or (ee) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 4;
- (gg) a nucleotide sequence represented by the 516th to 1448th bases of SEQ ID NO: 5;
- (hh) a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 6;
- (ii) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (gg) or (hh) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 6;
- (jj) a nucleotide sequence represented by the 116th to 1450th bases of SEQ ID NO: 7;
- $(\mbox{kk})$  a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 8;
- (11) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (jj) or (kk) under stringent conditions and

encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 8;

- (mm) a nucleotide sequence represented by the 807th to 1526th bases of SEQ ID NO: 9;
- (nn) a nucleotide sequence encoding the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10;
- (00) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (mm) or (nn) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10;
- (pp) a nucleotide sequence represented by the 156th to 1526th bases of SEQ ID NO: 9;
- $\,$  (qq) a nucleotide sequence encoding the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10;
- (rr) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (pp) or (qq) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10;

- (ss) a nucleotide sequence represented by the 156th to 806th bases of SEO ID NO: 9;
- (tt) a nucleotide sequence encoding the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10;
- (uu) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (ss) or (tt) under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10;
- (vv) a nucleotide sequence represented by SEQ ID NO:
  1;
- (ww) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (vv) under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 1;
- (xx) a nucleotide sequence represented by SEQ ID NO:
  3;
- inucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (xx) under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 3;

- (zz) a nucleotide sequence represented by SEQ ID NO: 5;
- (aaa) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (zz) under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 5;
- (bbb) A nucleotide sequence represented by SEQ ID NO:
  7;
- (ccc) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (bbb) under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 7;
- (ddd) a nucleotide sequence represented by SEQ ID NO:
  9;
- (eee) a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence (ddd) under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 9; and
- (fff) a fragment of these nucleotide sequences (aa) to (eee).
  - 43. (New) The process according to claim 23, wherein

the cells are E. coli cells, animal cells or insect cells.

- 44. (New) The method according to claim 33, wherein the specimen is a body fluid.
- 45. (New) The method according to claim 34, wherein the specimen is a body fluid.
- 46. (New) A method for screening for an inhibitor of serine protease comprising comparing the enzyme activity of the protein according to claim 41 upon bringing the protein into contact with a candidate compound with the enzyme activity of the protein without contact with the candidate compound.
- 47. (New) A pharmaceutical composition comprising the protein according to claim 41.
- 48. (New) A method for detecting a diagnostic marker for diseases in tissues comprising the protein according to claim 41, which comprises using an antibody against the protein according to claim 41.
- 49. (New) The method according to claim 48, wherein the marker is used for diagnosis of a cancer.

- 50. (New) A method for diagnosing Alzheimer's disease or epilepsy in the brain comprising using the marker according to claim 36.
- 51. (New) A method for diagnosing cancer or inflammation of the brain, prostate or testicle, comprising using the marker according to claim 36.
- 52. (New) A method for diagnosing sterility in semen or sperm comprising using the marker according to claim 36.
- 53. (New) A method for diagnosing prostatic hypertrophy comprising using the marker according to claim 36.

Please amend claim 20 as follows:

20. (Amended) A vector comprising the nucleotide sequence according to claim 42.

Please amend claim 21 as follows:

21. (Amended) Transformed cells having the nucleotide sequence according to claim 42 in an expressible state.

Please amend claim 22 as follows:

22. (Amended) A process for producing a protein which comprises culturing cells transformed with the nucleotide sequence (aa) to (ll), (vv) to (zz), (aaa), (bbb) or (ccc) of claim 42, and collecting mBSSP2 produced.

Please amend claim 23 as follows:

23. (Amended) A process for producing a protein which comprises culturing cells transformed with the nucleotide sequence (mm) to (uu), (ddd) or (eee) of claim 42, and collecting hBSSP2 produced.

Please amend claim 24 as follows:

24. (Amended) The process according to claim 22, wherein the cells are E. coli cells, animal cells or insect cells.

Please amend claim 29 as follows:

29. (Amended) An antibody against the protein according to claim 41 or a fragment thereof.

Please amend claim 31 as follows:

31. (Amended) A process for producing a monoclonal antibody against the protein according to claim 41 or a fragment thereof which comprises administering the protein according to

claim 41 or a fragment thereof to a warm-blooded animal other than a human being, selecting the animal whose antibody titer is recognized, collecting its spleen or lymph node, fusing the antibody producing cells contained therein with myeloma cells to prepare a monoclonal antibody producing hybridoma.

Please amend claim 32 as follows:

32. (Amended) A method for determining the protein according to claim 41 or a fragment thereof in a specimen which is based on immunological binding of an antigen against the protein or a fragment thereof to the protein or a fragment thereof in the specimen.

Please amend claim 33 as follows:

33. (Amended) A method for determining hBSSP2 or a fragment thereof in a specimen which comprises reacting a monoclonal antibody or a polyclonal antibody against the protein (i), (j), (k), (l), (m) or (n) of claim 41 or a modified derivative or fragment thereof and a labeled antibody with hBSSP2 or a fragment thereof in the specimen to detect a sandwich complex produced.

Please amend claim 34 as follows:

34. (Amended) A method for determining hBSSP2 or a fragment thereof in a specimen which comprises reacting a

monoclonal antibody or a polyclonal antibody against the protein (i), (j), (k), (l), (m) or (n) of claim 41 or a modified derivative thereof or a fragment thereof with labeled hBBSP2 and hBSSP2 or a fragment thereof in the specimen competitively to detect an amount of hBSSP2 or a fragment thereof in the specimen based on an amount of the labeled hBBSP2 reacted with the antibody.

Please amend claim 35 as follows:

35. (Amended) The method according to claim 32, wherein the specimen is a body fluid.

Please amend claim 36 as follows:

36. (Amended) A diagnostic marker for diseases in tissues comprising the protein according to claim 41.

## REMARKS

The present preliminary amendment is submitted in order to correct self-evident typographical and grammatical errors in the specification, as well as to define the invention better.

It is respectfully submitted that the claims are now in condition for examination, and prompt and favorable action is earnestly solicited.

Respectfully submitted,

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## "Version with markings to show changes"

Page 1, please amend the second paragraph as follows:

In general, proteases are biosynthesized as inactive precursors. They undergo limited hydrolysis in molecules to convert be converted into activated type proteases. In so far as enzymes are proteases, they have an activity for hydrolyzing a peptide bond, while their action modes are varied actions vary according to kinds of proteases. According to a particular kind of catalytic site, proteases are divided into serine proteases, cysteine proteases, aspartate proteases, metal proteases and the like. Proteases of each kind have a variety of properties, ranging from a protease having general digestive properties to a protease having various regulatory domains and strict substrate specificity, thereby specifically hydrolyzing only characteristic proteins.

Page 2, please amend the paragraph on page 2 as follows:

Further, proteins undergo various <u>types of</u> processing even after translation to produce active proteins. In many secretory proteins, a protein <u>areis</u> first synthesized on the ribosome in cytoplasm as an inactive precursor (pro-form) which comprises an active protein bearing at the N-terminus thereof a peptide of about 15 to 60 amino acids responsible for secretion (secretory signal). This peptide region is concerned with the

mechanism for passing through the cell membrane and is removed upon cleavage by a specific protease during the passage through the membrane, in almost all the cases, to produce the mature form. A secretory signal has a broad hydrophobic region comprising hydrophobic amino acids in the middle of the sequence, and basic amino acid residues at a site close to the N-terminus. A secretory signal is a synonym of for a signal peptide. In addition, in some proteins, a peptide moiety which functions as a secretory signal is further attached to the N-terminus of the inactive precursor (pro-form). Such a protein is called a prepro-protein (prepro-form).

Page 3, please amend the first paragraph as follows:

For example, trypsin is present as in the form of a prepro-form immediately after translation into amino acids.

After being secreted from cells, it is present as in the form of a pro-form and is then converted into active trypsin in the duodenum upon limited hydrolysis by enteropeptidase or by trypsin itself.

Page 3, please amend the second paragraph as follows:

The optimal pH range of serine proteases is neutral to
weak alkaline and, in general, many of them have a molecular

weight of about 30,000 or lower. All proteases of relating to blood coagulation, fibrinolysis and complement systems having a large molecular weight belong to the family of trypsin-like serine proteases. They have many regulator domains and form a protease cascade which is of very importance to reactions in a living body.

Page 4, please amend the first paragraph as follows:

Serine proteases expressed in a brain-nerve system such as neurosin are considered to play various roles in the brain-nerve system. Therefore, there is a possibility that isolation of a gene encoding a novel protease expressed in a brain-nerve system and production of a protein using the gene would be useful for diagnosis or therapeutic therapy of various diseases related to the brain-nerve system.

Page 4, please amend the second paragraph as follows:

Nowadays, in general, clinical diagnosis of

Alzheimer's disease is conducted based on the diagnosis

standard of DSM-IIIR and NINCDS-ADRDA (Mckhann, G. et al.,

Neurology, 34. 939, 1994) or the diagnosis standard of DSM-IV

(American Psychiatric Association; Diagnostic and statistical manuals of mental disorders, 4th ed., Washington DC, American Psychiatric Association, 1994). However, these standards are conditioned by a decline of in recognition functions which causes

a severe disability in a daily life or a social life. Then, it is pointed out that the diagnosis is less scientific objectivity than scientifically objective because the diagnosis may be influenced by the level of an individual's social life and further the specialty and experience of a physician who diagnoses particular conditions. In addition, definite diagnosis of Alzheimer's disease is conducted by pathohistological analyses and, in this respect, substantial inconsistency between clinical diagnosis and autopsy diagnosis is pointed out exists.

Page 5, please amend the paragraph on page 5 as follows:

image diagnosis is employed Αt present, а supplemental means in clinical diagnosis of Alzheimer's diagnosis and it is possible to analyze brain functions, for example, decline of metabolism and atrophy in specific sites such as hippocampus, parietal lobe of cerebral cortex and the like which are specific for Alzheimer's disease by PET and SPECT. However, to define Alzheimer's disease based on lowering of a blood flow from parietal lobe to temporal lobe is very dangerous. In addition, there is few a report showing that MRS testicle test is useful for patients with dementia including those of Alzheimer's disease. Further, although CT-MRI image diagnosis is used, a lesion of white matter such as atrophy of brain, PVL or the like is not specific for Alzheimer type dementia. Since older with aging, the above observation is not necessarily found in Alzheimer type dementia. Furthermore, since an image obtained by MRI varies according to strength of a magnetic field, performance of an the apparatus and imaging conditions, numerical data obtain obtained in different facilities cannot be compared with each other except for atrophic change. In addition, there is a limit to image measurement. Further, enlargement of the ventricle can be recognized in vascular dementia cases and there are cases wherein atrophy of the hippocampus is observed after ischemia of the basilar artery.

Page 6, please amend the fourth paragraph as follows:

Further, data obtained in different facilities

can be compared with each other by using the same diagnosis marker. Therefore, development of biological diagnosis markers is recognized to be a most important field among fields of Alzheimer's disease studies and its future prospects will be expected. Approaches to development of biological diagnosis markers up to now are divided into that those based on constitute components of characteristic pathological changes of Alzheimer's disease such as senile plaque and neurofibril change, and an approach based on other measures. Examples of the former include cerebrospinal fluid tau protein, Aβ and its precursor, βAPP.

Examples of the latter include mydriasis test with cholilytic drug, Apo E and other genes relating to Alzheimer's disease. However, no good results are have been obtained.

Page 7, please amend the paragraph on page 7 as follows:

Serine proteases are also considered to play an important role in cancer cells. The reason why extermination of cancer by surgical treatment or topical irradiation radioactive ray is difficult is metastasis the metastatic capability of cancer. For To spread of solid tumor cells in a body, they should loosen their adhesion to original adjacent cells, followed by separating from an original tissue, passing through other tissues to reach the blood vessel vessels or lymph node nodes, entering into the circulatory system through stratum basal and endothelial layer of the vessel, leave from the circulatory system at somewhere in the body, and surviving and proliferating in a new environment. While adhesion to adjacent epidermal cells is lost when expression of cadherin which is an intercellular adhesive molecule of epithelium is stopped, to break through tissues is considered to depend on proteolytic enzymes which decompose an extracellular matrix.

Page 8, please amend the first paragraph as follows:

As enzymes which decompose the matrix, mainly, metal proteases (Rha, S. Y. et al., Breast Cancer Research Treatment, 43, 175, 1997) and serine proteases are known. They cooperate to decompose matrix protein proteins such as collagen, laminin and fibronectin. Among the serine proteases known to be concerned in decomposition of the matrix, in particular, there is urokinase type plasminogen activator (U-PA). U-PA has a role as a trigger specific for a protein decomposition chain reaction. Its direct target is plasminogen. It is present in blood abundantly and is a precursor of an inactive serine protease which accumulates in reconstructed sites of tissues such as injured sites and tumors as well as inflammatory sites. addition, as proteases which are concerned in metastasis and infiltration of cancers, for example, a tissue factor, lysosomal type hydrolase and collagenase have been known.

Page 8, please amend the second paragraph as follows:

At present, cancer is the top cause of death in Japan and more than 200,000 people are died die per year. Then Accordingly, specific substances which can be used as markers for diagnosis and therapy or prophylaxis of cancer are studied intensively. Such specific substances are referred to as tumor markers or tumor marker relating biomarkers. They are utilized in aid of diagnosis before treatment of cancer, for presuming carcinogenic organ and pathological tissue type, for monitoring

effect of treatment, for finding recurrence early, for presuming prognosis, and the like. At present, tumor markers are essential in clinical analyses. Among them, alpha fetoprotein (AFP) which has high specificity to hepatocellular carcinoma and yolk sac tumor (Taketa K. et al., Tumour Biol., 9, 110, 1988), and carcinoembronic antigen (CEA) are used worldwide. In the future, tumor markers will be required more and more, and it is desired to develop, for example, organ specific markers and tumor cell specific markers which are highly reliable serologic diagnosis of cancer. Up to now, humunglandular kallikrein (hK2) which is a serine protease expressed at human prostatic epithelial cells has been reported as a marker for prostatic cancer. And, hK2 has 78% homology with the sequence of prostatic specific antigen (PSA) and PSA is also used widely as a biochemical marker of prostatic cancer (Mikolajczyk, S. d. et al., Prostate, 34, 44, 1998; Pannek, J. et al., Oncology, 11, 1273, 1997; Chu, T. M. et al., Tumour Biology, 18, 123, 1997; Hsieh, M. et al., Cancer Res., 57, 2651, 1997). Further, hK2 is reported to be useful as a marker for not only prostatic cancer but also stomach cancer (Cho, J. Y. et al.. Cancer, 79, 878, 1997). Moreover, CYFRA (CYFRA 211) for measuring cytokeratin 19 fragment in serum is reported to be useful for lung cancer (Sugiyama, Y. et al., Japan J. Cancer Res., 85, 1178, 1994). Gastrin release peptide precursor (ProGRP) is reported to be useful as a tumor marker (Yamaguchi, K. et al., Japan, J. Cancer Res., 86, 698, 1995).

Page 10, please amend the second paragraph as follows:

Under these circumstances, the present inventors

have succeeded in cloning of cDNA encoding novel human and

mouse serine proteases.

Page 10, please amend the third paragraph as follows:

In summary, the 1st feature of the present invention is <a href="mailto:the">the</a> amino acid sequences of <a href="mailto:biologically">biologically</a> active mature serine proteases BSSP2 and nucleotide sequences encoding the amino acid sequences.

Page 18, please amend the second paragraph as follows:

In case of northern blotting analysis, mBSSP2 shows the expression in the head of a 15-20 days mouse fetus, and in the lung, prostate and testicle of a 3 month-old mouse. hBSSP2 shows the expression in brain, skeletal muscle and liver (see Figs. 1, 2 and 5). In case of RT-PCR analysis, mBSSP2 shows the expression in the brain and testicle of a 12 day-old mouse, and hBSSP2 shows the expression in the brain and skeletal muscle. Then, the novel proteases of the present invention are presumed to play various roles in the brain, prostate, lung, testicle, skeletal muscle and liver. For example, in the brain, there is a possibility that they can be used for treatment and diagnosis of brain diseases such as Alzheimer's disease (AD), epilepsy, brain tumor and the like. Further, in other tissues, there is a possibility that BSSP2 of the present invention and a gene

encoding it can be used for treatment and diagnosis of various diseases such as cancer, inflammation, infertility, prostatomegaly and the like. Further, it is presumed they may have a certain influence on blood coagulation, fibrinolysis and complement systems. Furthermore, there is a possibility that inhibitors of serine proteases can be used for treatment and diagnosis of Alzheimer's disease, epilepsy, cancer, inflammation, infertility, prostatomegaly and the like.

Page 22, please amend the paragraph on page 22 as follows:

In general, many genes of eucaryote eucaryotes exhibit polymorphism and, sometimes, one or more amino acids are substituted by this phenomenon. Further, even in such a case, sometimes, a protein maintains its activity. Then, the present invention includes a gene encoding a protein obtained by modifying a gene encoding any one of the amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10, artificially, in so far as the protein has the characteristic function of the gene of the present invention. Further, the present invention includes a protein which is a modification of any one of amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10 in so far as the protein has the characteristics of the present invention. Modification is understood to include substitution, deletion, addition and/or insertion. In particular, the present inventors have shown that, even when several amino acids are

added to or deleted from the N-terminus amino acid of the BSSP2 mature protein

represented by SEQ ID NO: 2, the resultant sequence maintains its activity.

Page 23, please amend the first paragraph as follows:

That is, the present invention includes a protein

comprising any one of the amino acid sequences described in SEQ |

ID NOS: 2, 4, 6, 8 and 10; an amino acid sequence encoded

5 by any one of the nucleotide sequences represented by SEQ ID |

NOS: 1, 3, 5, 7 and 9; or one of these amino acid sequences

wherein one to several amino acids have been substituted,

deleted, added and/or inserted, and being belonging to serine |

protease family.

Page 23, please amend the second paragraph as follows:

Each codon for the desired amino acid itself has

been known and it can be selected freely. For example,

codons can be determined according to a conventional manner

by taking into consideration of the frequency of use of codons

in a host to be utilized (Grantham, R. et al., Nucleic Acids

Res., 9, r43, 1989). Therefore, the present invention also

includes a nucleotide sequence appropriately modified by taking into consideration of the degeneracy of a codon. Further, these nucleotide sequences can be modified by a site directed mutagenesis using a primer composed of a synthetic oligonucleotide encoding the desired modification (Mark, D. F. et al., Proc. Natl. Acad. Sci. USA., 81, 5662, 1984), or the like.

Page 23, please amend the third paragraph as follows: Furthermore, the DNA of the present invention includes DNA which is hybridizable to any one of the nucleotide sequences described in SEQ ID NOS: 1, 3, 5, 7 and 9 or nucleotide sequences complementary to these nucleotide sequences in so far as the protein encoded by the nucleotide sequence has the same properties as those of the BSSP2 of the present invention. is considered that many of the sequences which are hybridizable to a given sequence under stringent conditions have a similar activity to that of a protein encoded by the given sequence. The stringent conditions according to the present invention includes, for example, incubation in a solution containing 5 x SSC, 5%Denhardt's solution (0.1% BSA, 0.1% Ficol 1400, 0.1% PVP), 0.5% SDS and 20  $\mu g/ml$  denatured salmon sperm DNA at 37°C overnight, followed by washing with 2 x SSC containing 0.1% SDS at room temperature. Instead of SSC, SSPE can be appropriately used.

Page 25, please amend the second paragraph as follows: The present invention also relates to a vector comprising the nucleotide sequence represented by SEQ ID NO: 1 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 2; the nucleotide sequence represented by SEQ ID NO: 3 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 4; the nucleotide sequence represented by SEQ ID NO: 5 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 6; the nucleotide sequence represented by SEQ ID NO: 7 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 8; or the nucleotide sequence represented by SEQ ID NO: 9 or nucleotide sequence encoding the amino acid represented by SEQ ID NO: 10; or a nucleotide sequence similar them. A nucleotide sequence similar to a give given nucleotide sequence used herein means a nucleotide sequence which hybridizable to the given nucleotide sequence or complementary nucleotide sequence under the above-described stringent conditions and which encodes a protein having the same properties as those of the protein encoded by the nucleotide sequence.

Page 28, please amend the first paragraph as follows:

The animal cells and insect cells used herein

include cells derived from human being beings and cells derived

from fly flies or silk worm worms. For example there are CHO

cell cells, COS cell cells, BHK cell cells, Vero cell cells,

myeloma cell cells, HEK293 cells, HeLa cell cells, Jurkat cell

cells, mouse L cell cells, mouse C127 cell cells, 10 mouse FM3A

cell cells, mouse fibroblast, osteablast, cartilage cell cells,

S2, Sf9, Sf21, High Five (registered trade mark) cell cells and

the like.

Page 28, please amend the second paragraph as follows: The protein of the present invention as such can be expressed as a recombinant fused protein so as to facilitate isolation, purification and recognition. The recombinant fused protein used herein means a protein expressed as an adduct wherein a suitable peptide chain are is added to the N-terminus and/or C-terminus of the desired protein expressed by a nucleotide sequence encoding the desired protein. The recombinant protein used herein means that integrating a nucleotide sequence encoding the desired protein in the expression vector of the present invention and cut off an amino acid sequence which derived from nucleic acids other than those encoding the desired protein from the expressed recombinant fused protein, and is substantially the same as the protein of

the present invention.

Page 29, please amend the first paragraph as follows:

Introduction of the above vector into host cells can
be carried out by, for example, transfection according to <a href="the-">the</a>
lipopolyamine method, DEAE-dextran method, Hanahan method,
lipofectin method or calcium phosphate method, microinjection,
eletroporation and the like.

Page 33, please amend the first paragraph as follows: For obtaining fertilized egg cells efficiently, ovulation may be induced with gonadotropin or the like. Fertilized egg cells are recovered and a gene in an injection pipette is injected into male pronucleus of the egg cells by microinjection. For returning the injected egg cells to a fallopian tube, an animal (false pregnancy female mouse, etc.) is provided and about 10 to 15 eggs/mouse eggs/mice are transplanted. Then, genomic DNA is extracted from the end part of the tail to confirm whether the transgene is introduced into newborn mouse or not. This confirmation can be carried out by detection of the transgene with southern blot technique or PCR technique, or by positive cloning wherein a marker gene, activated only which is when homologous recombination is caused, has been introduced. transcribed products derived from the transgene are detected by

northern blot technique or RT-PCR technique to confirm expression of the transgene. Or, western blotting can be carried out with a specific antibody to a protein.

Page 33, please amend the second paragraph as follows: The knockout mouse of the present invention is treated so that the function of mBSSP2 gene is lost. A knockout mouse means a transgenic mouse in which any one of whose gene its genes is destroyed by homologous recombination technique so that its function is deficient. A knockout mouse can be created by carrying out homologous recombination with ES cells and selecting embryonic stem cells wherein either of allele genes are modified or destroyed. For example, embryonic stem cells whose genes are manipulated at blstocyte the blastocyte or morula stage of fertilized eggs are injected to obtain a chimera chimeric mouse wherein cells derived from the embryonic stem cells are mixed with those derived from the embryo. The chimera chimeric mouse (chimera chimeric means a single individual formed by somatic cells based on two or more fertilized eggs) can be mated with a normal mouse to create a heterozygote mouse wherein all of the allele genes have been modified or destroyed. Further, a homozygote mouse can be created by mating heterozygote mice.

Page 37, please amend the paragraph on page 37 as follows:

Examples of myeloma cells include X-63Ag8, NS-1, P3U1, SP2/0, AP-1 and the like with SP2/0 being preferred. preferred ratio of the number of the antibody producer cells (spleen cells): the number of spleen cells are 1:20 to 20:PEG (preferably PEG 1000 to PEG 6000) is added at a concentration of about 10 to 80% and the mixture is incubated at 20 to 40°C, preferably 30 to 37°C for 1 to 10 minutes to carry out the cell fusion efficiently. Screening of anti-hBSSP2 or mBSSP2 antibody producer hybridomas can be carried out by various For example, a supernatant of a hybridoma culture is added to a solid phase to which hBSSP2 or mBSSP2 antigen is adsorbed directly or together with a carrier (e.g., microplate), followed by addition of an anti-immunoglobulin antibody (in case that the cells used in cell fusion is are those of a mouse, antimouse immunoglobulin antibody is used) or protein A to detect the anti-hBSSP2 or mBSSP2 monoclonal antibody attached to the solid phase. Or, a supernatant of a hybridoma culture is added to a solid phase to which an anti-immunoglobulin antibody or protein A is adsorbed, followed by addition of hBSSP2 or mBSSP2 labeled with a radioactive substance, an enzyme, etc., to detect the anti-hBSSP2 or mBSSP2 monoclonal antibody attached to the solid phase.

Page 38, please amend the first paragraph as follows: Selection and cloning of the anti-hBSSP or mBSSP

monoclonal antibody can be carried out according to a per se known method or its modification. Normally, a HAT (hypoxanthine, aminopterin, thymidine)-added medium for culturing animal cells is used. Any culture medium can be used for selection, cloning and growing up in so far as the hybridoma can grow. For example, there can be used RPMI culture medium containing 1 to 20%, preferably 10 to 20% fetal bovine serum, a serum-free medium for culturing hybridomas. Preferably, the culture is carried out at a temperature of about 37°C. Normally, the culture time is 5days to 3 weeks, preferably 1 weeks week to 2 weeks. Normally, the culture is carried out under 5%  ${\rm CO}_2$ . The antibody titer of a supernatant of a hybridoma culture can be measured according to the same manner as that of the above-described measurement of anti-BSSP2 antibody titer in an antiserum. That is, examples of the measurement to be used include radioimmunoassay (RIA), enzyme-linked immunosorbent assay (ELISA), FIA (fluorescence immunoassay), plaque assay, agglutination reaction method, and the like. Among them, ELISA as shown blew below is preferred.

Page 39, please amend the paragraph on page 39 as follows:

Normally, cloning is carried out by a per se known method such as semi-solid agar method, limiting dilution method and the like. Specifically, after confirming a well in which the desired antibody is produced by the above-described method,

cloning is carried out to obtain a single clone. For cloning, it is preferred to employ limiting dilution method wherein hybridoma cells are diluted so that one colony is formed per one well of a culture plate. For cloning by limiting dilution method, feeder cells can be used, or a cell growth factor such as interleukin 6, etc. can be added to improve colony forming capability. In addition, cloning can be carried out by using FACS and single cell manipulation method. The cloned hybridoma is preferably cultured in a serum-free culture medium and an optimal amount of an antibody is added to its supernatant. The single hybridoma thus obtained can be cultured in a large about amount by using a flask or a cell culture device, or cultured in the abdominal cavity of an animal (J. Immunol. Meth., 53, 313, 1982) to obtain a monoclonal antibody. When culturing in a flask, there can be used a cell culture medium (e.g., IMDM, DMEM, RPMI1640, etc.) containing 0 to 20% of FCS. When culturing in the abdominal cavity of an animal, the animal to be used is preferably the same species or the same line as that from which the myeloma cells used in the cell fusion are derived, a thymus deficient nude mouse or the like, and the hybridoma transplanted after administration of a mineral oil such as pristane, etc. After 1 to 2 weeks, myeloma cells are proliferated in the abdominal cavity to obtain ascites containing a monoclonal antibody.

Page 42, please amend the paragraph on page 42 as follows:

The polyclonal antibody of the present invention can be produced according to a per se known method or modification. For example, an immunogen (protein antigen) per se or a complex thereof with a carrier protein is prepared and, according to the same manner as that in the above monoclonal antibody production, a warm-blooded animal is immunized. material containing an antibody against the protein of the present invention or its fragment is collected from the immunized animal and the antibody is separated and purified to obtain the desired antibody. As for a complex of an immunogen and a carrier protein for immunizing a warm-blooded animal, the kind of a carrier protein and the mixing ratio of a carrier and a hapten are not specifically limited in so far as an antibody against hapten immunized by cross-linking with the carrier is efficiently produced. For example, there can be used about 0.1 to 20, preferably about 1 to 5 parts by weight of bovine serum albumin, bovine cycloglobulin, hemocyanin, etc. coupled with one part by weight of a hapten. For coupling a carrier and a hapten, various condensing agents can be used. Examples thereof include glutaraldehyde, carbodiimide or maleimide active ester, active ester agents having thiol group or dithiopyridyl group, and the like. The condensed product is administered as such or together with a carrier or diluent to a site of a warm-blooded animal

where an antibody can be produced. For enhancing the antibody production, upon administration, Freund's complete adjuvant or Freund's incomplete adjuvant may be administrated administered. Normally, the administration is carried out once every 2 to 6 weeks, 3 to 10 times in all. The polyclonal antibody can be collected from blood, ascites, or the like, preferably blood of the immunized animal. The polyclonal antibody titer in an antiserum can be measured according to the same manner as measurement of the above monoclonal antibody titer in the antiserum. Separation and purification of the polyclonal antibody, like the above monoclonal antibody, can be carried out according to the same manner as those of immunoglobulins.

Page 44, please amend the second paragraph as follows:

As a sandwich method for determining hBSSP2 or mBSSP2
or a fragment thereof, there can be used a two step method, a one
step method and the like. In the two step method, first, the
immobilized antibody is reacted with hBSSP2 or mBSSP2 or a
fragment thereof and then unreacted materials are completely
removed by washing, followed by addition of the labeled antibody
to form immobilized antibody-hBSSP2 or mBSSP2-labeled antibody.
In the one step method, the immobilized antibody, labeled
antibody and hBSSP2 or mBSSP2 or a fragment thereof are added at
the same time.

Page 45, please amend the second paragraph as follows: For immobilization of the antibody, a known chemical bonding method or a physical adsorption can be used. Examples chemical bonding method include method glutaraldehyde; maleimide method using  $\frac{N}{2}$  succusinimidyl  $\frac{N}{2}$  $\underline{\textbf{succinimidyl-4-}}$  (N-maleimidomethyl) cyclohexane-1-carboxylate, Nsuccusinimidyl-2-maleimide N-succinimdyl-2-maleimide acetate or the like; carbodiimide method using 1-ethvl-3-(3dimethylaminopropyl)carbodiimide hydrochloride; or the like. In addition, there are maleimidobenzoyl-N-hydroxysuccinimide ester method, N-succinimidyl-3-(2-pyridylthio)propionic acid method, bisdiazobenzidine method, and dipalmityllysine method. Or, it is possible to capture a complex formed beforehand by reacting a material material to be tested with two antibodies, whose epitopes are different, with an immobilized a 3rd antibody against the antibody.

Page 46, please amend the paragraph on page 46 as follows:

For labeling, it is preferred to use an enzyme, fluorescent substance, luminous substance, radioactive substance, metal chelate, or the like. Examples of the enzyme include peroxidase, alkaline phosphatase,  $\beta$ -D-galactosidase, malate dehydrogenase, Staphylococcus nuclease,  $\delta$ -5-steroidisomerase,  $\alpha$ -glycerol phosphate dehydrogenase, triose phosphate isomerase,

horseradish peroxidase, asparaginase, glucose oxidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, acetylcholinesterase and the like. Examples of the fluorescent substance include fluorescein isothiocyanate, phycobiliprotein, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthalaldehyde, and the like. Examples of the luminous substance include isoluminol, lucigenin, luminol, aromatic acridinium ester, imidazole, acrdinium salt and its modified ester, luciferin, luciferase, aequorin and the like. Examples of the radioactive substance include  $^{125}\text{I}$ ,  $^{127}\text{I}$ ,  $^{131}\text{I}$ ,  $^{14}\text{C}$ ,  $^3\mathrm{H},~^{32}\mathrm{P},~^{35}\mathrm{S}$  and the like. The labeling material is not limited to them and any material which can be used for immunological determination can be used. Further, a low molecular weight hapten such as biotin, dinitrophenyl, pyridoxal or fluorescamine may be attached to the antibody. Preferably, horseradish peroxidase is used as a labeling enzyme. This enzyme can be reacted with various substrates and can readily be attached to the antibody by periodate method.

Page 47, please amend the first paragraph as follows: When an enzyme is used as a labeling material, a substrate and, if necessary, a coloring enzyme is used for measuring its activity. In case of using peroxidase as the enzyme,  $\rm H_2O_2$  is used as a substrate and, as a coloring agent, there can be used 2,2'-azino-di-[3-ethylbenzthiazoline sulfonic

ammonium salt (ABTS), 5'-aminosalicylic acid, oacid phenylenediamine, 4-aminoantipyrine, 3,3',5,5'tetramethylbenzidine and the like. In case of using alkaline phosphatase as the enzyme, o-nitorphenylphosphate onitrophenylphosphate, p-nitrophenylphosphoric acid, or the like can be used as a substrate. In case of using  $\beta\text{-}D\text{-}galactosidase$ fluorescein-d-( $\beta$ -D-galactopyranoside), as the enzyme, methylumbelliphenyl- $\beta$ -D-galactopyranoside, or the like can be used as a substrate. The present invention also includes a kit comprising the above monoclonal antibody, polyclonal antibody and reagents.

Page 47, please amend the second paragraph as follows: As a cross-linking agent, a known cross-linking agent such N,N'-o-phenylenedimaleimide, 4-(Nas maleimidomethyl)cyclohexanoate-N-succinimide ester, maleimidohexanoate-N-succineimide ester, 4,4'-dithiopyridine or the like can be utilized. The reaction of these cross-linking agents with enzymes and antibodies can be carried out by a known method according to properties of a particular cross-linking Further, as the antibody, a fragment thereof, for agent. example, Fab', Fab, F(b'2) can be used as the case may be. A labeled enzyme can be obtained by the same treatment regardless of whether the antibody is polyclonal or monoclonal. above labeled enzyme obtained by using a cross-linking agent is

purified by a known method such as affinity chromatography or the like, a an immunoassay system having more higher sensitivity can be obtained. The enzyme labeled and purified antibody is stored at in a dark cold place with addition of a stabilizer such as thimerosal, glycerin or after lyophilization.

Page 49, please amend the paragraph on page 49 as follows:

The cloning was carried out by PCR using a mouse brain cDNA library (Clontech) as a template and nucleotide sequences corresponding to an amino acid sequence common to serine proteases represented by

Primer 1: GTG CTC ACN GCN GCB CAY TG (SEQ ID NO: 20)

Primer 2: CCV CTR WSD CCN CCN GGC GA (SEQ ID NO: 21)

as primers. Namely, 5 µl of the template, 5 µl of 10 x ExTaq buffer, 5 µl of dNTP, 10 pmol of each of the above primers and 0.5 µl of ExTaq (TAKARA) were added and the total volume was adjusted to 50 µl with sterilized water. PCR was carried out by repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for 1 minutes minute, 30 times. The PCR product was mixed with pCR II-TOPO vector attached to TOPO TA cloning kit (Invitrogen) and the mixture was allowed to stand at room temperature for 5 minutes. Then, according to a conventional manner, E. coli Top 10 attached to the kit was transformed and applied to a LB (Amp⁺) plate (containing 100

 $\mu g/ml$  of ampicillin). According to a conventional manner, a plasmid was extracted from each colony obtained and its nucleotide sequence was determined by cycle sequencing method with a fluorescence sequencer (ABI). Homology of the sequence of each clone was examined by means of GenBank. Regarding an unknown sequence, i.e., BSSP2 gene, the full length cDNA was obtained by 5' RACE and 3' RACE and, according to the same manner as described above, the nucleotide sequence was determined. Namely, BSSP2 clone specific primers, GSP1 primers [mBSSP2.2 (SEQ ID NO: 27) or mBSSP2.0 (SEQ ID NO: 22)] and GSP2 primers [mBSSP2R2 (SEQ ID NO: 28) or mBSSP2.1 (SEQ ID NO: 23)] were prepared. PCR was carried out by using mouse brain Marathon-Ready cDNA (Clontech), AP1 primer attached to this reagent and either of the above GSP1 primers and heating at  $94^{\circ}\text{C}$  for 2 minutes once and repeating a cycle of heating at  $94^{\circ}\text{C}$  for 30seconds, at 60°C for 30 seconds and then at 72°C for 30 seconds 35 times. Then, 5  $\mu l$  of the PCR product diluted to 1/100, 5  $\mu l$ of 10 x buffer, 5  $\mu l$  of dNTP, 10 pmol of either of 10  $\mu M$  of the above GSP2 primer, 10 pmol of AP2 primer attached to the above reagent and 0.5 unit of ExTaq were admixed and adjusted to 50  $\mu\text{l}$ with sterilized water. Then, according to the same manner as the above, PCR was carried out. The PCR product was cloned by the above TOPO TA cloning kit and sequenced to obtain the upstream and downstream regions of the above clone. At this time, as for a clone which seemed not to cover the full length of a protein,

the specific primers shown hereinafter were prepared based on the newly found nucleotide sequence. Further, based on this sequence, the primers capable of amplifying ORF as shown hereinafter [mBSSPF7 (SEQ ID NO: 26), mBSSP2R/E (SEQ ID NO: 29)] were prepared and PCR carried out using mouse brain Marathonready cDNA as a template to confirm that these clones were This was cloned into pCR II-TOPO vector attached to identical. TOPO TA cloning kit to obtain the plasmid pCR II/mBSSP2 containing the full length cDNA clone. The nucleotide sequence of DNA contained in this plasmid is shown in SEQ ID NO: 7 and the amino acid sequence of mSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 8. Further, two different types of clones were obtained. The nucleotide sequences of these DNA are shown in SEQ ID NOS: 3 and 5, respectively. The amino acid sequences of mBSSP2 proteins deduced from these nucleotide sequences are shown in SEQ ID NOS: 4 and 6. These novel proteases are divided into types 1, 2 and 3. Type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids and type 3 is composed of 445 amino acids. These amino acid sequences contained the common amino acid sequence composed of 238 amino acids whose N-terminus side started with Ile-Val-Gly-Gly-Gln-Ala-Val as the mature serine protease. Further, in the amino acid sequence of the mature serine protease, a consensus sequence having a serine protease activity was contained. Furthermore, since there were two or more amino acid sequence

<u>sequences</u> specific for a sugar chain bonding site, it was presumed that the amino acid sequence had at least two sugar chains.

Page 53, please amend the paragraph on page 53 as follows:

As seen <u>form from</u> Figs. 1 and 2, in the case of prostate and testicle. Further, according to the results of RT-PCR, the expression was recognized in the testicle of 3-month-old mice.

# IN THE CLAIMS

- 20. (Amended) A vector comprising the nucleotide sequence according to any one of claims 2, 4, 6, 8, 10, 12 and 14-19 claim 42.
- 21. (Amended) Transformed cells having the nucleotide sequence according to any one of claims 2, 4, 6, 8, 10, 12 and 14-19 claim 42 in an expressible state.
  - 22. (Amended) A process for producing a protein which

comprises culturing cells transformed with the nucleotide sequence according(aa) to any one (11), (vv) to (zz), (aaa), | (bbb) or (ccc) of claims 2, 4, 6, 8, 15-18 claim 42, and | collecting mBSSP2 produced.

- 23. (Amended) A process for producing a protein which comprises culturing cells transformed with the nucleotide sequence according(mm) to any one(uu), (ddd) or (eee) of claims | 10, 12, 14 or 19 claim 42, and collecting hBSSP2 produced.
- 24. (Amended) The process according to claim  $22 \frac{1}{100} = 23$ , wherein the cells are  $E.\ coli$  cells, animal cells or insect cells.
- 29. (Amended) An antibody against the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 claim 41 or a fragment thereof.
- 31. (Amended) A process for producing a monoclonal antibody against the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 claim 41 or a fragment thereof which comprises administering the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 claim 41 or a fragment thereof to a warm-blooded animal other than a human being, selecting the

animal whose antibody titer is recognized, collecting its spleen or lymph node, fusing the antibody producing cells contained therein with myeloma cells to prepare a monoclonal antibody producing hybridoma.

- 32. (Amended) A method for determining the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 claim 41 or a fragment thereof in a specimen which is based on immunological binding of an antigen against the protein or a fragment thereof to the protein or a fragment thereof in the specimen.
- fragment thereof in a specimen which comprises reacting a monoclonal antibody or a polyclonal antibody against the protein according to any one (i), (j), (k), (l), (m) or (n) of claims 9, 11 and 13 claim 41 or a modified derivative or fragment thereof and a labeled antibody with hBSSP2 or a fragment thereof in the specimen to detect a sandwich complex produced.
- 34. (Amended) A method for determining hBSSP2 or a fragment thereof in a specimen which comprises reacting a monoclonal antibody or a polyclonal antibody against the protein according to any one of claims 9, 11 and 13 and (i), (j), (k), (l), (m) or (n) of claim 41 or a modified derivative thereof or

a fragment thereof with labeled hBBSP2 and hBSSP2 or a fragment thereof in the specimen competitively to detect an amount of hBSSP2 or a fragment thereof in the specimen based on an amount of the labeled hBBSP2 reacted with the antibody.

- 35. (Amended) The method according to any one of claims 32-34 claim 32, wherein the specimen is a body fluid.
- 36. (Amended) A diagnostic marker for diseases in tissues comprising the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 claim 41.

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

ATTY.'S DOCKET: UEMURA 7

In re Application of:
Hidetoshi UEMURA et al.
)
Examiner:
)

I.A. No.: PCT/JP99/06475
)
Washington, D.C.
)

Filed: 19 November 1999
)
May 21, 2001
)
For: NOVEL SERINE PROTEASE BSSP2)

# PRELIMINARY AMENDMENT

Honorable Commissioner for Patents and Trademarks Washington, D.C. 20231

Sir:

Contemporaneous with the filing of this case and prior to calculation of the filing fee, kindly amend as follows:

### IN THE SPECIFICATION

After the title please insert the following paragraph:

## REFERENCE TO RELATED APPLICATIONS

--The present application is the national stage under 35 U.S.C. §371 of international application PCT/JP99/06475, filed 19 November 1999 which designated the United States, and which application was not published in the English language.--

# REMARKS

The above amendment to the specification is being made to insert reference to the PCT application of which the present case is a U.S. national stage.

Favorable consideration is earnestly solicited.

Respectfully submitted, BROWDY AND NEIMARK, P.L.L.C. Attorneys for Applicant

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### FIELD OF THE INVENTION

NOVEL SERINE PROTEASE BSSP2

The present invention relates isolated to polynucleotides of human and mouse serine proteases referred (hereinafter to "hBSSP2" "mBSSP2", as and respectively, and, in case no differentiation thereof from each other is needed, simply referred to as "BSSP2"), and their homologous forms, mature forms, precursors polymorphic variants as well as a method for detecting thereof. Further, it relates to hBSSP2 and mBSSP2 proteins, compositions containing hBSSP2 and mBSSP2 polynucleotides and proteins, as well as their production and use.

### BACKGROUND OF THE INVENTION

In general, proteases are biosynthesized as inactive precursors. They undergo limited hydrolysis in molecules to convert into activated type proteases. In so far as enzymes are proteases, they have an activity for hydrolyzing a peptide bond, while their action modes are varied according to kinds of proteases. According to a particular kind of catalytic site, proteases are divided into serine proteases, cysteine proteases, aspartate

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proteases, metal proteases and the like. Proteases of each kind have a variety of properties, ranging from a protease having general digestive properties to a protease having various regulatory domains and strict substrate specificity, thereby specifically hydrolyzing only characteristic proteins.

Further, proteins undergo various processing even after translation to produce active proteins. In many secretory proteins, a protein are first synthesized on the ribosome in cytoplasm as an inactive precursor (pro-form) which comprises an active protein bearing at the N-terminus thereof a peptide of about 15 to 60 amino acids responsible for secretion (secretory signal). This peptide region is concerned with the mechanism for passing through the cell and is removed upon cleavage by a specific membrane protease during the passage through the membrane, in almost all the cases, to produce the mature form. A secretory signal a broad has hydrophobic region comprising hydrophobic amino acids in the middle of the sequence, and basic amino acid residues at a site close to the N-terminus. A secretory signal is a synonym of a signal peptide. addition, in some proteins, a peptide moiety which functions as a secretory signal is further attached to the N-terminus of the inactive precursor (pro-form). protein is called a prepro-protein (prepro-form).

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For example, trypsin is present as a prepro-form immediately after translation into amino acids. After being secreted from cells, it is present as a pro-form and is converted into active trypsin in duodenum upon limited hydrolysis by enteropeptidase or by trypsin itself.

The optimal pH range of serine proteases is neutral to weak alkaline and, in general, many of them have a molecular weight of about 30,000 or lower. All proteases of blood coagulation, fibrinolysis and complement systems having a large molecular weight belong to trypsin-like serine proteases. They have many regulator domains and form a protease cascade which is of very importance to reactions in a living body.

Recently, cDNAs and amino acid sequences of many novel proteases have been determined by PCR for consensus sequences of serine proteases using oligonucleotide primers. According to this method, novel proteases have been found by various researchers such as Yamamura et al. (Yamanura, Y et al., Biochem. Biophys. Res. Commun., 239, 386, 1997), Gschwend, et al. (Gschwend, T. P. et al., Mol. Cell. Neurosci., 9. 207, 1997), Chen et al. (Chen, Z-L, et al., J. Neurosci., 15, 5088, 1995) and others.

SEQ ID NO: 3 of JP 9-149790 A discloses neurosin as a novel serine protease. Neurosin has also been reported in Biochimica et Byophysica Acta, 1350, 11-14,

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1997. By this, there is provided a method for mass production of neurosin using the serine protease gene and a method for screening specific inhibitors using the enzyme. In addition, the screening method has been shown to be useful for screening medicines for treating various diseases.

Serine proteases expressed in a brain-nerve system such as neurosin are considered to play various roles in the brain-nerve system. Therefore, there is a possibility that isolation of a gene encoding a novel protease expressed in a brain-nerve system and production of a protein using the gene would be useful for diagnosis or therapeutic of various diseases related to the brain-nerve system.

in general, Nowadays, clinical diagnosis Alzheimer's disease is conducted based on the diagnosis standard of DSM-IIIR and NINCDS-ADRDA (Mckhann, G. et al., Neurology, 34. 939, 1994) or the diagnosis standard of DSM-(American Psychiatric Association; Diagnostic and statistical manuals of mental disorders, 4th Washington DC, American Psychiatric Association, 1994). However, these standards are conditioned by decline of recognition functions which causes a severe disability in a daily life or a social life. Then, it is pointed out that the diagnosis is less scientific objectivity because the

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diagnosis may be influenced by the level of an individual's social life and further the specialty and experience of a physician who diagnoses particular conditions. In addition, definite diagnosis of Alzheimer's disease is conducted by pathohistological analyses and, in this respect, substantial inconsistency between clinical diagnosis and autopsy diagnosis is pointed out.

At present, image diagnosis is employed as supplemental means in clinical diagnosis of Alzheimer's diagnosis and it is possible to analyze brain functions, for example, decline of metabolism and atrophy in specific sites such as hippocampus, parietal lobe of cerebral cortex and the like which are specific for Alzheimer's disease by PET and SPECT. However, to define Alzheimer's disease based on lowering of a blood flow from parietal lobe to temporal lobe is very dangerous. In addition, there is few report showing that MRS testicle useful for patients with dementia including those of Alzheimer's disease. although CT-MRI image diagnosis is used, a lesion of white matter such as atrophy of brain, PVL or the like is not specific for Alzheimer type dementia. Since it has been reported that atrophy of brain proceeds as getting older, the above observation is not necessarily found in Alzheimer Furthermore, since an image obtained by MRI type dementia. varies according to strength of a magnetic

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performance of an apparatus and imaging conditions, numerical data obtain in different facilities cannot be compared with each other except atrophic change. In addition, there is a limit to image measurement. Further, enlargement of ventricle can be recognized in vascular dementia cases and there are cases wherein atrophy of hippocampus is observed after ischemia of basilar artery.

Under these circumstances, many researchers have requested to develop biological diagnosis markers as a means for providing better precision and objectivity for clinical diagnosis of Alzheimer's disease. At the same time, the following important roles in the future will be expected.

- 1) Objective judgment system of effect of medicaments for treating Alzheimer's disease.
- 2) Detection of Alzheimer's disease before a diagnosis standard is met, or disease conditions are manifested.

can be compared with each other by using the same diagnosis marker. Therefore, development of biological diagnosis markers is recognized to be a most important field among fields of Alzheimer's disease studies and its future prospects will be expected. Approaches to development of biological diagnosis markers up to now are divided into

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that based on constitute components of characteristic pathological changes of Alzheimer's disease such as senile plaque and neurofibril change, and an approach based on other measures. Examples of the former include cerebrospinal fluid tau protein,  $A\beta$  and its precursor,  $\beta APP$ . Examples of the latter include mydriasis test cholilytic drug, Apo E and other genes relating Alzheimer's disease. However, no good results are obtained.

Serine proteases are also considered to play important role in cancer cells. The extermination of cancer by surgical treatment or topical irradiation of radioactive ray is difficult is metastasis capability of cancer. For spread of solid tumor cells in a they should loosen their adhesion to body, original adjacent cells, followed by separating from an original tissue, passing through other tissues to reach blood vessel or lymph node, entering into the circulatory system through stratum basal and endothelial layer of the vessel, leave from the circulatory system at somewhere in the body, and surviving and proliferating in a new environment. While adhesion to adjacent epidermal cells is lost expression of cadherin which is an intercellular adhesive molecule of epithelium is stopped, to break through tissues considered to depend on proteolytic enzymes decompose an extracellular matrix.

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As enzymes which decompose the matrix, mainly, metal proteases (Rha, S. Y. et al., Breast Cancer Research Treatment, 43, 175, 1997) and serine proteases are known. They cooperate to decompose matrix protein such as collagen, laminin and fibronectin. Among serine proteases known to be concerned in decomposition of the matrix, in particular, there is urokinase type plasminogen activator (U-PA). U-PA has role as trigger specific for а protein decomposition chain reaction. Its direct target plasminogen. It is present in blood abundantly and is a precursor of an inactive serine protease which accumulates in reconstructed sites of tissues such as injured sites and tumors as well as inflammatory sites. In addition, as proteases which are concerned in metastasis and infiltration of cancers, for example, a tissue factor, lysosomal type hydrolase and collagenase have been known.

At present, cancer is the top cause of death in Japan and more than 200,000 people are died per year. Then, specific substances which can be used as markers for diagnosis and therapy or prophylaxis of cancer are studied intensively. Such specific substances are referred to as tumor markers or tumor marker relating biomarkers. They are utilized in aid of diagnosis before treatment of cancer, for presuming carcinogenic organ and pathological tissue type, for monitoring effect of treatment, for finding

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recurrence early, for presuming prognosis, and the like. present, tumor markers are essential in Among them, alpha fetoprotein (AFP) which has analyses. high specificity to hepatocellular carcinoma and yolk sac tumor (Taketa K. et al., Tumour Biol., 9, 110, 1988), and carcinoembronic antigen (CEA) are used worldwide. future, tumor markers will be required more and more, and it is desired to develop, for example, organ specific markers and tumor cell specific markers which are highly reliable serologic diagnosis of cancer. ФŪ humunglandular kallikrein (hK2) which is a serine protease expressed at human prostatic epithelial cells has been reported as a marker for prostatic cancer. And, hK2 has 78% homology with the sequence of prostatic specific antigen (PSA) and PSA is also used widely as a biochemical marker of prostatic cancer (Mikolajczyk, S. d. et al., Prostate, 34, 44, 1998; Pannek, J. et al., Oncology, 11, 1273, 1997; Chu, T. M. et al., Tumour Biology, 18, 123, 1997; Hsieh, M. et al., Cancer Res., 57, 2651, 1997). Further, hK2 is reported to be useful as a marker for not only prostatic cancer but also stomach cancer (Cho, J. Y. et al.. Cancer, 79, 878, 1997). Moreover, CYFRA (CYFRA 21for measuring cytokeratin 19 fragment in serum is reported to be useful for lung cancer (Sugiyama, Y. et al., Japan J. Cancer Res., 85, 1178, 1994). Gastrin release

peptide precursor (ProGRP) is reported to be useful as a tumor marker (Yamaguchi, K. et al., Japan, J. Cancer Res., 86, 698, 1995).

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## OBJECTS OF THE INVENTION

Thus, the main object of the present invention is to provide a novel serine protease which can be used for treating or diagnosing various diseases such as Alzheimer's disease (AD), epilepsy, cancer, inflammation, infertility, prostatomegaly and the like in various tissues such as brain, lung, prostate, testicle, skeletal muscle, liver and the like, and can be used as an excellent marker instead of that presently used.

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## SUMMARY OF THE INVENTION

Under these circumstances, the present inventors have succeeded in cloning of cDNA encoding novel human and mouse serine proteases.

In summary, the 1st feature of the present invention is amino acid sequences of biological active mature serine proteases BSSP2 and nucleotide sequences encoding the amino acid sequences.

That is, they are the amino acid sequence composed of 238 amino acids (mature type BSSP2 (SEQ ID NO: 2)) and a nucleotide sequence encoding the amino acid

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sequence (the 1st to 714th bases of SEQ ID NO: 1). addition, they include amino acid sequences substantially similar to SEQ ID NO: 2 and nucleotide sequences encoding such similar amino acid sequences. Further, they include modified derivatives of proteins having these amino acid sequences. An amino acid sequence substantially similar to a given amino acid sequence used herein means an amino acid sequence derived from the given amino acid sequence by modification such as substitution, deletion, addition and/or insertion of one to several amino acids with maintaining the same property as that of the protein having the given amino acid sequence. The modified derivative of the proteins includes, for example, phosphate adduct, sugar chain adduct, metal adduct (e.g., calcium adduct), the protein fused to another protein such as albumin etc., dimer of the protein, and the like.

In the nucleotide sequences in the Sequence Listing hereinafter, the symbol "n" represents that any of the normal bases of a nucleic acid, i.e., adenine (a), cytosine (c), guanine (g) and thymine (t) is present at that position.

The 2nd feature of the present invention is an amino acid sequence composed of 273 amino acids [type 1 BSSP2 (SEQ ID NO: 4)] wherein 35 amino acids of -35th to -1st amino acids represented by SEQ ID NO: 4 are added to

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the N-terminus side of the mature BSSP2 amino acid sequence (SEQ ID NO: 2) and a nucleotide sequence encoding the amino acid sequence (247th to 1065th bases of SEQ ID NO: 3). feature addition. this includes amino acid sequences substantially similar to SEQ ID NO: 4 and nucleotide sequences encoding these substantially similar amino acid sequences. Further, this feature includes modified derivatives of proteins having these amino acid sequences.

The 3rd feature of the present invention is an amino acid sequence composed of 311 amino acids [type 2 BSSP2 (SEQ ID NO: 6)] wherein 73 amino acids of -73rd to -1st amino acids represented by SEQ ID NO: 6 are added to the N-terminus side of the mature BSSP2 amino acid sequence (SEQ ID NO: 2) and a nucleotide sequence encoding the amino acid sequence (516th to 1448th bases of SEO ID NO: 5). addition. this feature includes amino acid sequences substantially similar to SEQ ID NO: 6 and nucleotide sequences encoding these substantially similar amino acid sequences. Further, this feature includes derivatives of proteins having there amino acid sequences.

The 4th feature of the present invention is an amino acid sequence composed of 445 amino acids [type 3 BSSP2 (SEQ ID NO: 8)] wherein 207 amino acids of -207th to -1st amino acids represented by SEQ ID NO: 8 are added to the N-terminus side of the mature BSSP2 amino acid sequence

(SEQ ID NO: 2) and a nucleotide sequence encoding the amino acid sequence (116th to 1450th bases of SEQ ID NO: 7). In addition, this feature includes amino acid sequences substantially similar to SEQ ID NO: 8 and nucleotide sequences encoding these substantially similar amino acid sequences. Further, this feature includes modified derivatives of proteins having there amino acid sequences.

The 5th feature of the present invention is an amino acid sequence of a biologically active, mature human serine protease, hBSSP2, and a nucleotide sequence encoding the amino acid sequence. That is, they are an amino acid sequence [mature type hBSSP2 (SEQ ID NO: 10) composed of 240 amino acids represented by SEQ ID NO: 10 (1st to 240th amino acids) and a nucleotide sequence encoding the amino acid sequence (807th to 1526th bases of SEQ ID NO: 9). addition, this feature includes amino acid sequences substantially similar to SEQ ID NO: 10 (1st to 240th amino acids) and nucleotide sequences encoding these substantially similar amino acid sequences. Further, this feature includes modified derivatives of proteins having there amino acid sequences.

The 6th feature of the present invention is an amino acid sequence composed of 457 amino acids (-217th to 240th amino acids of SEQ ID NO: 10) wherein 217 amino acids of -217th to -1st amino acids represented by SEQ ID NO: 10

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are added to the N-terminus side of the mature human serine protease hBSSP2 amino acid sequence (1st to 240 amino acids of SEQ ID NO: 10) and a nucleotide sequence encoding the amino acid sequence (156th to 1526th bases of SEQ ID NO: 9). In addition, this feature includes amino acid sequences substantially similar to SEQ ID NO: 10 and nucleotide sequences encoding these substantially similar amino acid sequences. Further, this feature includes modified derivatives of proteins having there amino acid sequences.

The 7th feature of the present invention is an amino acid sequence composed of 217 amino acids of -217th to -1st amino acids of SEQ ID NO: 10 and a nucleotide sequence encoding the amino acid sequence (156th to 806th bases of SEQ ID NO: 9). In addition, this feature includes amino acid sequences substantially similar to the amino acid composed of 217 amino acids of -217th to -1st SEQ ID NO: 10 and nucleotide sequences encoding substantially similar amino acid sequences. Further, this feature includes modified derivatives of proteins having there amino acid sequences.

The present invention also relates to the nucleotide sequences represented by SEQ ID NOS: 1, 3, 5, 7 and 9 as well as nucleotide sequences similar to them.

The 8th feature of the present invention is a vector comprising the nucleotide sequence according to any

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of the above 1st to the 7th feature, and transformant cells transformed with the vector.

The 9th feature of the present invention is a process for producing BSSP2 protein from the transformed cells of the 8th feature.

The 10th feature of the present invention is a transgenic non-human animal, wherein the expression level of BSSP2 gene has been altered.

The 11th feature of the present invention is an antibody against BSSP2 protein or its fragment and a process for producing thereof.

The 12th feature of the present invention is a method for determining BSSP2 protein or its fragment in a specimen using the antibody of the 11th feature.

The 13th feature is a diagnostic marker of diseases comprising BSSP2 protein.

Hereinafter, unless otherwise stated, the nucleotide sequence represented by each SEQ ID NO: includes the above-described various fragments thereof, and similar nucleotide sequences and their fragments. Likewise, the amino acid sequence represented by each SEQ ID NO: includes the above-described various fragments thereof, similar nucleotide sequences and their fragments, and modified derivatives thereof. In addition, unless otherwise stated, BSSP2, hBSSP2, and mBSSP2 include proteins having the

above-described respective amino acid sequences.

## BRIEF DESCRIPTION OF THE DRAWINGS

- Fig. 1 illustrates the results of northern blotting using mRNAs prepared from mice in Example 2 hereinafter.
  - Fig. 2 illustrates the results of northern blotting using mRNAs prepared from mice in Example 2 hereinafter.
  - Fig. 3 is a plasmid constructed by the method of Example 4 hereinafter.
  - Fig. 4 illustrates the construction of plasmid pFBTrypSigTag/BSSP2 according to the method of Example 4 hereinafter.
  - Fig. 5 illustrates the detection of hBSSP2 mRNA by northern hybridization.
  - Fig. 6 illustrates the detection of hBSSP2 mRNA by RT-PCR.
- Fig. 7 illustrates the expression of hBSSP2 by a 20 baculovirus system.

## DETAILED DESCRIPTION OF THE INVENTION

The nucleotide sequences encoding hBSSP2 or mBSSP2 of the present invention can be obtained by preparing mRNAs from cells expressing the protein and

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converting it into double stranded DNAs according to a conventional manner. For preparing mRNA, isothiocyanate-calcium chloride method (Chirwin, et al., Biochemistry, 18, 5294, 1979) or the like can be used. preparing poly (A) + RNA from total RNAs, there can be used affinity chromatography using a carrier, for example, Sepharose, latex particles, etc., to which oligo (dT) is attached, and the like. The above-obtained RNA can be used as a template and treated with reverse transcriptase by using, as a primer, oligo (dT) which is complementary to the poly (A) strand at the 3'-terminus, or a random primer, or a synthesized oligonucleotide corresponding to a part of the amino acid sequence of hBSSP2 or mBSSP2 to obtain a hybrid mRNA strand comprising DNA or cDNA complementary to the mRNA. The double stranded DNA can be obtained by treating the above-obtained hybrid mRNA strand with E. coli RNase, E. coli DNA polymerase and E. coli DNA ligase to convert into a DNA strand.

It is also possible to carry out cloning by RT-PCR method using primers synthesized on the basis of the nucleotide sequence of hBSSP2 or mBSSP2 gene and using hBSSP2 or mBSSP2 expressing cell poly (A) + RNA as a template. Alternatively, the desired cDNA can be obtained without using PCR by preparing or synthesizing a probe on the basis of the nucleotide sequence of hBSSP2 or mBSSP2

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gene and screening a cDNA library directly. Among genes obtained by these methods, the gene of the invention can be selected by confirming a nucleotide sequence thereof. The gene of the present invention can also be prepared according to a conventional method using chemical syntheses of nucleic acids, for example, phosphoamidite method (Mattencci, M. D. et al., J. Am. Chem. Soc., 130, 3185, 1981) and the like.

By using the thus-obtained hBSSP2 or mBSSP2 gene, their expression in various tissues can be examined.

In case of northern blotting analysis, mBSSP2 shows the expression in head of 15-20 days mouse fetus, and lung, prostate and testicle of 3 month-old mouse. hBSSP2 shows the expression in brain, skeletal muscle and liver (see Figs. 1, 2 and 5). In case of RT-PCR analysis, mBSSP2 shows the expression in brain and testicle of 12 day-old mouse, and hBSSP2 shows the expression in brain and skeletal muscle. Then, the novel proteases of the present invention are presumed to play various roles in brain, prostate, lung, testicle, skeletal muscle and liver. example, in brain, there is a possibility that they can be used for treatment and diagnosis of brain diseases such as Alzheimer's disease (AD), epilepsy, brain tumor and the Further, in other tissues, there is a possibility that BSSP2 of the present invention and a gene encoding it

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can be used for treatment and diagnosis of various diseases such as cancer, inflammation, infertility, prostatomegaly and the like. Further, it is presumed they may have a certain influence on blood coagulation, fibrinolysis and complement systems. Furthermore, there is a possibility that inhibitors of serine proteases can be used for treatment and diagnosis of Alzheimer's disease, epilepsy, cancer, inflammation, infertility, prostatomegaly and the like.

The novel mouse serine protease can be divided into types 1, 2 and 3. It has been shown that type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids, and type 3 is composed of 445 amino acids. These amino acid sequences contain a common amino acid sequence of 238 amino acids whose N-terminus side starts Ile-Val-Gly-Gly-Gln-Ala-Val as the mature protease. Further, the amino acid sequence of the mature serine protease contains a consensus sequence having serine protease activity. Since there are two or more amino acid sequences which are characteristic of sugar chain binding sites, the amino acid sequence is presumed to have at least two sugar chains.

Furthermore, in the novel human serine protease (hBSSP2), there are a transmembrane region and a scavenger receptor cysteine rich-like domain in the N-terminus side

of hBSSP2 mature protein as seen from SEQ ID NO: 10.

The term "pro part" used herein means a part of a pro-form, i.e., the pro-form from which the corresponding active type protein part is removed. The term "pre part" used herein means a part of a prepro-form, i.e., the prepro-form from which the corresponding pro-form is removed. The term "prepro part" used herein means a part of a prepro-form, i.e., the prepro-form from which the corresponding active type protein part is removed.

The amino acid sequence represented by SEQ ID NO: 2 is the BSSP2 mature or active type protein composed of 238 amino acids, and the nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 1 is composed of 714 bases. The present inventors have shown that the serine protease activity is maintained even when one to several amino acids of the N-terminus of the mature type protein of the present invention is deleted or added, while the sequence represented by SEQ ID NO: 2 is preferred.

The amino acid sequence represented by SEQ ID NO:

4 is type 1 BSSP2 protein composed of 273 amino acids, and
the nucleotide sequence encoding the amino acid sequence
represented SEQ ID NO: 3 is composed of 1685 bases. The
sequence of the -35th to -1st amino acids is the prepro or
pro part and the amino acid sequence represented by SEQ ID
NO: 4 is considered to be a precursor type of the BSSP2

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protein.

The amino acid sequence represented by SEQ ID NO: 6 is type 2 BSSP 2 protein composed of 311 amino acids and the nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 5 is composed of 2068 bases. The sequence of the -73rd to -1st amino acids is the prepro or pro part and the amino acid sequence represented by SEQ ID NO: 6 is considered to be a precursor type of BSSP2 protein.

The amino acid sequence represented by SEQ ID NO: 8 is type 3 BSSP2 protein composed of 445 amino acids and the nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 7 is composed of 2070 bases. The amino acid sequence of the -207th to -1st amino acids is the prepro or pro part and the amino acid sequence represented by SEQ ID NO: 8 is considered to be a precursor type of BSSP2 protein.

SEQ ID NOS: 4, 6 and 8 contain the common amino acid sequence represented by SEQ ID NO: 2 as the mature BSSP2 protein. Further, each of amino acid sequences of -25th to 238th amino acids in SEQ ID NOS: 4, 6 and 8 is the consensus sequence.

The amino acid sequence represented by SEQ ID NO: 10 is hBSSP2 protein composed of 457 amino acids and the nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 9 is composed of 1371 bases.

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Since a transmembrane region and a scavenger receptor cysteine rich-like domain are present in the amino acid sequence of the -217th to -1st amino acids of SEQ ID NO: 10, it is considered that hBSSP2 exhibits its activity not only in the form of the mature protein but also in the form of an adduct of the -217th to -1st amino acids.

general, many genes of eucaryote exhibit polymorphism and, sometimes, one or more amino acids are substituted by this phenomenon. Further, even in such case, sometimes, a protein maintains its activity. Then, the present invention includes a gene encoding a protein obtained by modifying a gene encoding any one of the amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10, artificially, in so far as the protein has characteristic function of the gene of the present invention. Further, the present invention includes a protein which is a modification of any one of amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10 in so far as the protein has the characteristics of present invention. Modification is understood to include substitution, deletion, addition and/or insertion. In particular, the present inventors have shown that, even when several amino acids are added to or deleted from the N-terminus amino acid οf the BSSP2 mature protein represented by SEQ ID NO: 2, the resultant sequence

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maintains its activity.

That is, the present invention includes a protein comprising any one of amino acid sequences described in SEQ ID NOS: 2, 4, 6, 8 and 10; an amino acid sequence encoded by any one of nucleotide sequences represented by SEQ ID NOS: 1, 3, 5, 7 and 9; or one of these amino acid sequences wherein one to several amino acids have been substituted, deleted, added and/or inserted, and being belonging to serine protease family.

Each codon for the desired amino acid itself has been known and it can be selected freely. For example, codons can be determined according to a conventional manner by taking into consideration of frequency of use of codons in a host to be utilized (Grantham, R. et al., Nucleic Acids Res., 9, r43, 1989). Therefore, the present invention also includes a nucleotide sequence appropriately modified by taking into consideration of degeneracy of a codon. Further, these nucleotide sequences can be modified by a site directed mutagenesis using a primer composed of a synthetic oligonucleotide encoding the desired modification (Mark, D. F. et al., Proc. Natl. Acad. Sci. USA., 81, 5662, 1984), or the like.

Furthermore, the DNA of the present invention includes DNA which is hybridizable to any one of nucleotide sequences described in SEQ ID NOS: 1, 3, 5, 7 and 9 or

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nucleotide sequences complementary to these nucleotide sequences in so far the protein encoded by as nucleotide sequence has the same properties as those of the BSSP2 of the present invention. It is considered that many of sequences which are hybridizable to a given sequence under stringent conditions have a similar activity to that of a protein encoded by the given sequence. The stringent conditions according to the present invention includes, for example, incubation in a solution containing 5 x SSC, 5% Denhardt's solution (0.1% BSA, 0.1% Ficol 1400, 0.1% PVP), 0.5% SDS and 20 µg/ml denatured salmon sperm DNA at 37°C overnight, followed by washing with 2 x SSC containing 0.1% SDS at room temperature. Instead of SSC, SSPE can be appropriately used.

Probes for detecting a BSSP2 gene can be designed based on any one of nucleotide sequences described in SEQ ID NOS: 1, 3, 5, 7 and 9. Or, primers can be designed for amplifying DNA or RNA containing the nucleotide sequence. To design probes or primers is carried out routinely by a person skilled in the art. An oligonucleotide having a designed nucleotide sequence can be synthesized chemically. And, when a suitable label is added to the oligonucleotide, the resultant oligonucleotide can be utilized in various hybridization assay. Or, it can be utilized in nucleic acid synthesis reactions such as PCR. An oligonucleotide

to be utilized as a primer has, preferably, at least 10 bases, more preferably 15 to 50 bases in length. An oligonucleotide to be utilized as a probe has, preferably, 100 bases to full length.

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Moreover, it is possible to obtain a promoter region and an enhancer region of a BSSP2 gene present in the genome based on the cDNA nucleotide sequence of BSSP2 provided by the present invention. Specifically, these control regions can be obtained according to the same manner as described in JP 6-181767 A; J. Immunol., 155, 2477, 1995; Proc. Natl. Acad. Sci., USA, 92, 3561, 1995 and the like. The promoter region used herein means a DNA region which is present upstream from a transcription initiation site and controls expression of a gene. The enhancer region used herein means a DNA region which is present in an intron, a 5'-non-translated region or a 3'-non-translated region and enhances expression of a gene.

The present invention also relates to a vector comprising the nucleotide sequence represented by SEQ ID NO: 1 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 2; the nucleotide sequence represented by SEQ ID NO: 3 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 4; the nucleotide sequence represented by SEQ ID NO: 4; the nucleotide sequence represented by SEQ ID NO: 5 or a nucleotide sequence encoding the amino acid

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sequence represented by SEQ ID NO: 6; the nucleotide sequence represented by SEQ ID NO: 7 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 8; or the nucleotide sequence represented by SEO ID NO: 9 or a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 10; or a nucleotide sequence similar to them. A nucleotide sequence similar to a give nucleotide sequence used herein means a nucleotide sequence which is hybridizable to the given nucleotide sequence or its complementary nucleotide sequence under the above-described stringent conditions and encodes a protein having the same properties as those of the protein encoded by the nucleotide sequence.

The vector is not specifically limited in so far as it can express the protein of the present invention. Examples thereof include pBAD/His, pRSETA, pcDNA2.1, pBlueBac4.5, pcDNA3.1 pTrcHis2A, pYES2, and manufacture by Invitrogen, pET and pBAC manufactured by pGEM manufactured by Promega, pBluescriptII manufactured by Stratagene, pGEX and pUC18/19 manufactured by Pharmacia, PfastBAC1 manufactured by GIBCO and the like. Preferably, a protein expression vector (described in the specification of a patent application entitled "Protein expression vector and its use" and filed by the same applicant on the same day) is used. This expression vector

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is constructed by using pCRII-TOPO vector described in the hereinafter, Examples a commercially available or expression vector, for example pSecTag2A vector pSecTag2B vector (Invitrogen) and integrating a secretory signal nucleotide sequence suitable for expression of the protein of the present invention, in the 3' downstream side thereof, a Tag nucleotide sequence, a cleavable nucleotide sequence and a cloning site, into which a nucleotide sequence encoding a target protein can be inserted, in this More specifically, it is preferred to use trypsin order. signal as the secretory signal, a nucleotide sequence encoding polyhistidine as the Tag nucleotide sequence, and a nucleotide sequence encoding an amino acid sequence which is susceptible to enzyme-specific cleavage, i.e., nucleotide sequence encoding the amino acid sequence of Asp-Asp-Asp-Lys (said amino acid sequence is recognized by enterokinase, and the recombinant fusion protein is cleaved at the C-terminus part thereof) as the cleavable nucleotide sequence.

Furthermore, the present invention provides transformed cells having the nucleotide sequence of the present invention in an expressible state by means of the above vector. Preferably, host cells to be used for the transformed cells of the present invention are animal cells and insect cells. However, host cells include any cells

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(including those of microorganisms) which can express a nucleotide sequence encoding the desired protein in the expression vector of the present invention and can secrete extracellularly.

The animal cells and insect cells used herein include cells derived from human being and cells derived from fly or silk worm. For example, there are CHO cell, COS cell, BHK cell, Vero cell, myeloma cell, HEK293 cells, HeLa cell, Jurkat cell, mouse L cell, mouse C127 cell, mouse FM3A cell, mouse fibroblast, osteoblast, cartilage cell, S2, Sf9, Sf21, High Five[™] (registered trade mark) cell and the like.

The protein of the present invention as such can be expressed as a recombinant fused protein so as to facilitate isolation, purification and recognition. The recombinant fused protein used herein means a protein expressed as an adduct wherein a suitable peptide chain are added to the N-terminus and/or C-terminus of the desired protein expressed by a nucleotide sequence encoding the desired protein. The recombinant protein used herein means that obtained by integrating a nucleotide sequence encoding the desired protein in the expression vector of the present invention and cut off an amino acid sequence which derived from nucleic acids other than those encoding the desired protein from the expressed recombinant fused protein, and

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is substantially the same as the protein of the present invention.

Introduction of the above vector into host cells can be carried out by, for example, transfection according to lipopolyamine method, DEAE-dextran method, Hanahan method, lipofectin method or calcium phosphate method, microinjection, eletroporation and the like.

As described above, the present invention also relates to a process for producing hBSSP2 of mBSSP2 comprising culturing cells transformed with the above nucleotide sequence of the present invention and collecting the produced hBSSP2 of mBSSP2. The culture of cells and separation and purification of the protein can be carried out by a per se known method.

The present invention also relates to an inhibitor of the novel serine protease of the present invention. Screening of the inhibitor can be carried out according to a per se known method such as comparing the enzyme activity upon bringing into contact with a candidate compound with that without contact with the candidate compound, or the like

The present invention relates to a non-human transgenic animal whose expression level of hBSSP2 or mBSSP2 gene has been altered. The hBSSP2 or mBSSP2 gene used herein includes cDNA, genomic DNA or synthetic DNA

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encoding hBSSP2 or mBSSP2. In addition, expression of a gene includes any steps of transcription and translation. The non-human transgenic animal of the present invention is useful for studies of functions or expression control of hBSSP2 or mBSSP2, elucidation of mechanisms of diseases in which hBSSP2 or mBSSP2 is presumed to be involved, and development of disease model animals for screening and safety test of medicine.

In the present invention, expression of a gene can be modified artificially by mutagenizing at a part of important sites which control normal expression (enhancer, promoter, intron, etc.) such deletion, substitution, addition and/or insertion increase or decrease an expression level of the gene in comparison with its inherent expression level. mutagenesis can be carried out according to a known method to obtain the transgenic animal.

In a narrow sense, the transgenic animal means an animal wherein a foreign gene is artificially introduced into reproductive cells by gene recombinant techniques. In a broad sense, the transgenic animal includes an antisense transgenic animal the function of whose specific gene is inhibited by using antisense RNA, an animal whose specific gene is knocked out by using embryonic stem cells (ES cells), and an animal into which point mutation DNA is

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introduced, and the transgenic animal means an animal into which a foreign gene is stably introduced into a chromosome at an initial stage of ontogeny and the genetic character can be transmitted to the progeny.

The transgenic animal used herein should be understood in a broad sense and includes any vertebrates other than a human being. The transgenic animal of the present invention is useful for studies of functions or expression control of BSSP2, elucidation of mechanisms of diseases associated with cells expressing in a human being, and development of disease model animals for screening and safety test of medicine.

As a technique for creating the transgenic animal, a gene is introduced into a nucleus in a pronucleus stage of egg cells with a micropipette directly under a phase-contrast microscope (microinjection, U.S. Patent 4,873,191). Further, there are a method using embryonic stem cell (ES cell), and the like. In addition, there are newly developed methods such as a method wherein a gene is introduced into a retroviral vector or adenoviral vector to infect egg cells, a sperm vector method wherein a gene is introduced into egg cells through sperms, and the like.

A sperm vector method is a gene recombinant technique wherein a foreign gene is incorporated into sperm cells by adhesion, electroporation, etc., followed by

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fertilization of egg cells to introduce the foreign gene into the egg cells (M. Lavitranoet et al., Cell, 57, 717, 1989). Alternatively, an in vivo site specific gene recombinant technique such as that using cre/loxP recombinase system of bacteriophage P1, FLP recombinase system of Saccharomyces cerevisiae, etc. can be used. Furthermore, introduction of a transgene of the desired protein into a non-human animal using a retroviral vector has been reported.

For example, a method for creating a transgenic animal by microinjection can be carried out as follows.

First, a transgene primarily composed of promoter responsible for expression control, encoding a specific protein and a poly A signal is required. It is necessary to confirm expression modes and amounts between respective systems because an expression mode and amount of a specific molecule is influenced by a promoter activity, and transgenic animals differ from each other according to a particular system due to the difference in a copy number of an introduced transgene and a introduction site on a chromosome. An intron sequence which is spliced may be previously introduced before the poly A signal because it has been found that an expression amount varies due to a non-translation region and splicing. Purity of a gene to be used for introduction into fertilized egg cells

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should be as high as possible. This is of importance. Animals to be used include a mouse for collecting fertilized eggs (5 to 6 week old), a male mouse for mating, a false pregnancy female mouse, a seminiferous tubule-ligated mouse, and the like.

For obtaining fertilized egg cells efficiently, ovulation may be induced with gonadotropin or the like. Fertilized egg cells are recovered and a gene injection pipette is injected into male pronucleus of the egg cells by microinjection. For returning the injected egg cells to a fallopian tube, an animal (false pregnancy female mouse, etc.) is provided and about 10 15 eggs/mouse are transplanted. Then, genomic extracted from the end part of the tail to confirm whether the transgene is introduced into newborn mouse or not. This confirmation can be carried out by detection of the transgene with southern blot technique or PCR technique, or by positive cloning wherein a marker gene, which activated only when homologous recombination is caused, has been introduced. Further, transcribed products derived from the transgene are detected by northern blot technique or RT-PCR technique to confirm expression of the transgene. Or, western blotting can be carried out with a specific antibody to a protein.

The knockout mouse of the present invention is

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treated so that the function of mBSSP2 gene is lost. knockout mouse means a transgenic mouse any of whose gene is destroyed by homologous recombination technique so that its function is deficient. A knockout mouse can be created by carrying out homologous recombination with ES cells and selecting embryonic stem cells wherein either of allele genes are modified or destroyed. For example, embryonic stem cells whose genes are manipulated at blstocyte or morula stage of fertilized eggs are injected to obtain a chimera mouse wherein cells derived from the embryonic stem cells are mixed with those derived from the embryo. chimera mouse (chimera means a single individual formed by somatic cells based on two or more fertilized eggs) can be mated with a normal mouse to create a heterozygote mouse wherein all of the allele genes have been modified or destroyed. Further, a homozygote mouse can be created by mating heterozygote mice.

recombination Homologous means recombination between two genes whose nucleotide sequences are the same verv similar to each other in terms of recombination mechanism. PCR can be employed to select homologous recombinant cells. A PCR reaction can be carried out by using a part of a gene to be inserted and a part of a region where the insertion is expected as primers to find out occurrence of homologous recombination in cells

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which give an amplification product. Further, for causing homologous recombination in a gene expressed in embryonic stem cells, homologous recombinant cells can readily be selected by using a known method or its modification. For example, cells can be selected by joining a neomycin resistant gene to a gene to be introduced to impart neomycin resistance to cells after introduction.

The present invention also provide an antibody recognizing hBSSP2 or mBSSP2 or a fragment thereof. The antibody of the present invention includes an antibody against a protein having the amino acid sequence described in any of SEQ ID NOS: 2, 4, 6, 8 and 10 or its fragment. An antibody against hBSSP2 or mBSSP2 or a fragment thereof (e.g., polyclonal antibody, monoclonal antibody, peptide antibody) or an antiserum can be produced by using hBSSP2 or mBSSP2 or a fragment thereof, etc. as an antigen according to a per se known process for producing an antibody or an antiserum.

The hBSSP2 or mBSSP2 of a fragment thereof is administered to a site of a warm-blooded animal where an antibody can be produced by administration thereof as such or together with a diluent or carrier. For enhancing the antibody production, upon administration, Freund's complete adjuvant or Freund's incomplete adjuvant may be administrated. Normally, the administration is carried out

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once every 1 to 6 weeks, 2 to 10 times in all. Examples of the warm-blooded to be used include monkey, rabbit, dog, quinea pig, mouse, rat, sheep, goat, chicken and the like with mouse and rat being preferred. As rats, for example, Wistar and SD rats are preferred. As mice, for example, BALB/c, C57BL/6 and ICR mice are preferred.

For producing monoclonal antibody producer cells, individuals whose antibody titer have been recognized are selected from warm-blooded animals, e.g., a mouse immunized with an antigen. Two to 5 days after the last immunization, the spleen or lymph node of the immunized animal collected and antibody producer cells contained therein are subjected to cell fusion with myeloma cells to prepare a monoclonal antibody producer hybridoma. The antibody titer in an antiserum can be determined by, for example, reacting the antiserum with a labeled hBSSP2 or mBSSP2 as described hereinafter, followed by measurement of the activity bound The cell fusion can be carried out to the antibody. according to a known method, for example, that described by Koehler and Milstein (Nature, 256, 495, 1975) or its modifications (J. Immunol. Method, 39, 285, 1980; Eur. J. biochem, 118, 437, 1981; Nature, 285, 446, 1980). fusion promoting agent, there are polyethylene glycol (PEG), Sendai virus and the like. Preferably, PEG is used. Further, for improving fusion efficiency, lectin, poly-L-

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lysine or DMSO can be appropriately added.

Examples of myeloma cells include X-63Ag8, NS-1, P3U1, SP2/0, AP-1 and the like with SP2/0 being preferred. The preferred ratio of the number of the antibody producer cells (spleen cells) : the number of spleen cells are 1 : 20 to 20 : 1. PEG (preferably PEG 1000 to PEG 6000) is added at a concentration of about 10 to 80% and the mixture is incubated at 20 to 40°C, preferably 30 to 37°C for 1 to 10 minutes to carry out the cell fusion efficiently. Screening of anti-hBSSP2 or mBSSP2 antibody producer hybridomas can be carried out by various methods. example, a supernatant of a hybridoma culture is added to a solid phase to which hBSSP2 or mBSSP2 antigen is adsorbed directly or together with a carrier (e.g., microplate), followed by addition of an anti-immunoglobulin antibody (in case that the cells used in cell fusion is those of a mouse, anti-mouse immunoglobulin antibody is used) or protein A to anti-hBSSP2 detect the or mBSSP2 monoclonal antibody attached to the solid phase. Or, a supernatant of a hybridoma culture is added to a solid phase to which an anti-immunoglobulin antibody or protein A is adsorbed, followed by addition of hBSSP2 or mBSSP2 labeled with a radioactive substance, an enzyme, etc., to detect the antihBSSP2 or mBSSP2 monoclonal antibody attached to the solid phase.

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Selection and cloning of the anti-hBSSP or mBSSP monoclonal antibody can be carried out according to a per se known method or its modification. Normally, a HAT (hypoxanthine, aminopterin, thymidine)-added medium culturing animal cells is used. Any culture medium can be used for selection, cloning and growing up in so far as the hybridoma can grow. For example, there can be used RPMI culture medium containing 1 to 20%, preferably 10 to 20% fetal bovine serum, a serum-free medium for culturing hybridomas. Preferably, the culture is carried out at a temperature of about 37°C. Normally, the culture time is 5 days to 3 weeks, preferably 1 weeks to 2 weeks. Normally, the culture is carried out under 5% CO2. The antibody titer of a supernatant of a hybridoma culture can be measured according to the same manner as that of the abovedescribed measurement of anti-BSSP2 antibody titer in an That is, examples of the measurement to be used include radioimmunoassay (RIA), enzyme-linked immunosorbent assay (ELISA), FIA (fluorescence immunoassay), plaque assay, agglutination reaction method, and the like. Among them, ELISA as shown blew is preferred.

Screening by ELISA

A protein prepared according to the same operation as that for an immunogen is immobilized on the surface of each well of an ELISA plate. Next, BSA, MSA,

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OVA, KLH, gelatin, skimmed milk, or the like is immobilized each well to prevent non-specific adsorption. supernatant of a hybridoma culture is added to each well and is allowed to stand for a given time so that an immunological reaction proceeds. Each well is washed with a washing solution such as PBS or the like. Preferably, a surfactant is added to this washing solution. An enzyme labeled secondary antibody is added and allowed to stand for a given time. As the enzyme to be used for the label, there can be used  $\beta$ -galactosidase, alkaline phosphatase, peroxidase and the like. After washing each well with the same washing solution, a substrate solution of the labeled enzyme used is added so that an enzymatic reaction proceeds. When the desired antibody is present in the supernatant of a hybridoma culture, the enzymatic reaction proceeds and the color of the substrate solution is changed.

Normally, cloning is carried out by a per known method such as semi-solid agar method, limiting dilution method and the like. Specifically, confirming a well in which the desired antibody is produced by the above-described method, cloning is carried out to obtain a single clone. For cloning, it is preferred to employ limiting dilution method wherein hybridoma cells are diluted so that one colony is formed per one well of a culture plate. For cloning by limiting dilution method,

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feeder cells can be used, or a cell growth factor such as interleukin 6, etc. can be added to improve colony forming In addition, cloning can be carried out by capability. using FACS and single cell manipulation method. The cloned hybridoma is preferably cultured in a serum-free culture medium and an optimal amount of an antibody is added to its The single hybridoma thus obtained can be supernatant. cultured in a large about by using a flask or a cell culture device, or cultured in the abdominal cavity of an animal (J. Immunol. Meth., 53, 313, 1982) to obtain a monoclonal antibody. When culturing in a flask, there can be used a cell culture medium (e.g., IMDM, DMEM, RPMI1640, etc.) containing 0 to 20% of FCS. When culturing in the abdominal cavity of an animal, the animal to be used is preferably the same species or the same line as that from which the myeloma cells used in the cell fusion are derived, a thymus deficient nude mouse or the like, and the hybridoma is transplanted after administration of a mineral oil such as pristane, etc. After 1 to 2 weeks, myeloma cells are proliferated in the abdominal cavity to obtain ascites containing a monoclonal antibody.

The monoclonal antibody of the present invention which does not cross-react with other proteins can be obtained by selecting a monoclonal antibody which recognizes an epitope specific to hBSSP2 or mBSSP2. In

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general, an epitope presented by an amino acid sequence composed of at least 3, preferably 7 to 20 successive amino acid residues in an amino acid sequence which constitutes a particular protein is said to be an inherent epitope of the protein. Then, a monoclonal antibody recognizing epitope constituted by a peptide having an amino acid sequence composed of at least 3 successive amino acid residue selected from the amino acid residues disclosed in any of SEQ ID NOS: 2, 4, 6 and 8 can be said to be the monoclonal antibody specific for BSSP2 of the present An epitope common to BSSP2 family can be invention. selected by selecting an amino acid sequence conservative among the amino acid sequences described in SEQ ID NOS: 2, 4, 6, 8 and 10. Or, in case of a region containing an acid sequence specific for each sequence, antibody which can differentiate respective monoclonal proteins can be selected.

Separation and purification of the anti-hBSSP2 or mBSSP2 monoclonal antibody, like a conventional polyclonal antibody, can be carried out according to the same manner as those of immunoglobulins. As a known purification method, there can be used a technique, for example, salting out, alcohol precipitation, isoelectric precipitation, electrophoresis, ammonium sulfate precipitation, absorption and desorption with an ion exchange material (e.g., DEAE),

ultrafiltration, gel filtration, or specific purification by collecting only an antibody with an antibody-binding solid phase or an active adsorber such as protein A or protein G, etc., and dissociating the binding to obtain the antibody. For preventing formation of aggregates during purification or decrease in the antibody titer, for example, human serum albumin is added at a concentration of 0.05 to 2%. Alternatively, amino acids such as glycine,  $\alpha$ -alanine, etc., in particular, basic amino acids such as lysine, arginine, histidine, etc., saccharides such as glucose, mannitol, etc., or salts such as sodium chloride, etc. can be added. In case of IgM antibody, since it is very liable to be aggregated, it may be treated with  $\beta$ -propionilactone and acetic anhydride.

The polyclonal antibody of the present invention can be produced according to a per se known method or its modification. For example, an immunogen (protein antigen) per se or a complex thereof with a carrier protein is prepared and, according to the same manner as that in the above monoclonal antibody production, a warm-blooded animal is immunized. A material containing an antibody against the protein of the present invention or its fragment is collected from the immunized animal and the antibody is separated and purified to obtain the desired antibody. As for a complex of an immunogen and a carrier protein for

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immunizing a warm-blooded animal, the kind of a carrier protein and the mixing ratio of a carrier and a hapten are not specifically limited in so far as an antibody against hapten immunized by cross-linking with the carrier efficiently produced. For example, there can be used about 0.1 to 20, preferably about 1 to 5 parts by weight of bovine serum albumin, bovine cycloglobulin, hemocyanin, etc. coupled with one part by weight of a hapten. For coupling a carrier and a hapten, various condensing agents can be Examples thereof include used. glutaraldehyde, carbodiimide or maleimide active ester, active ester agents having thiol group or dithiopyridyl group, and the like. The condensed product is administered as such or together with a carrier or diluent to a site of a warm-blooded animal where an antibody can be produced. For enhancing antibody production, upon administration, Freund's complete adjuvant or Freund's incomplete adjuvant may be administrated. Normally, the administration is carried out once every 2 to 6 weeks, 3 to 10 times in all. The polyclonal antibody can be collected from blood, ascites, or the like, preferably blood of the immunized animal. polyclonal antibody titer in an antiserum can be measured according to the same manner as measurement of the above monoclonal antibody titer in the antiserum. Separation and purification of the polyclonal antibody, like the above

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monoclonal antibody, can be carried out according to the same manner as those of immunoglobulins.

The monoclonal antibody and polyclonal antibody against hBSSP2 or mBSSP2 or a fragment thereof can be utilized for diagnosis and treatment of diseases associated with cells expressing hBSSP2 or mBSSP2. By using these antibodies, hBSSP2 or mBSSP2 or a fragment thereof can be determined based on their immunological binding to hBSSP2 or mBSSP2 or a fragment thereof of the present invention. Specifically, examples of a method for determining hBSSP2 or mBSSP2 or a fragment thereof in a specimen by using these antibodies include a sandwich method wherein the antibody attached to an insoluble carrier and the labeled antibody are reacted with hBSSP2 or mBSSP2 or a fragment thereof to form a sandwich complex and the sandwich complex detected, as well as a competitive method wherein labeled hBSSP2 or mBSSP2, and hBSSP2 or mBSSP2 or fragment thereof in the specimen are competitively reacted with the antibody and hBSSP2 or mBSSP2 or a fragment thereof in the specimen is determined based on the amount of the labeled antigen reacted with the antibody.

As a sandwich method for determining hBSSP2 or mBSSP2 or a fragment thereof, there can be used two step method, one step method and the like. In two step method, first, the immobilized antibody is reacted with hBSSP2 or

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mBSSP2 or a fragment thereof and then unreacted materials are completely removed by washing, followed by addition of the labeled antibody to form immobilized antibody-hBSSP2 or mBSSP2-labeled antibody. In one step method, the immobilized antibody, labeled antibody and hBSSP2 or mBSSP2 or a fragment thereof are added at the same time.

Examples of an insoluble carrier used for the determination include synthetic resins such as polystyrene, polyethylene, polypropylene, polyvinyl chloride, polyester, polyacrylate, nylon, polyacetal, fluorine plastic, etc.; polysaccharides such as cellulose, agarose, etc.; glass; metal; and the like. An insoluble carrier may be shaped in various forms, for example, tray, sphere, fiber, rod plate, container, cell, test tube, and the like. The antibody adsorbed by a carrier is stored at a cold place in the presence of an appropriate preservative such as sodium azide or the like.

For immobilization of the antibody, a known chemical bonding method or a physical adsorption can be used. Examples of a chemical bonding method include a method using glutaraldehyde; maleimide method using N-succusinimidyl-4-(N-maleimidomethyl)cyclohexane-1-carboxylate, N-succusinimidyl-2-maleimide acetate or the like; carbodiimide method using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride; or the like.

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In addition, there are maleimidobenzoyl-N-hydroxysuccinimide ester method, N-succinimidyl-3-(2-pyridylthio)propionic acid method, bisdiazobenzidine method, and dipalmityllysine method. Or, it is possible to capture a complex formed beforehand by reacting a material to be tested with two antibodies, whose epitopes are different, with an immobilized a 3rd antibody against the antibody.

For labeling, it is preferred to use enzyme, fluorescent substance, luminous substance, radioactive substance, metal chelate, or the like. Examples of the include peroxidase, alkaline phosphatase, enzyme galactosidase, malate dehydrogenase, Staphylococcus nuclease,  $\delta$ -5-steroidisomerase,  $\alpha$ -glycerol phosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, asparaginase, glucose oxidase, ribonuclease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, acetylcholinesterase and the like. Examples of fluorescent the substance include fluorescein isothiocyanate, phycobiliprotein, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthalaldehyde, and the like. Examples of the luminous substance include isoluminol, lucigenin, luminol, aromatic acridinium ester, imidazole, acrdinium salt and its modified ester, luciferin, luciferase, aequorin and the like. Examples of radioactive substance include 125I, 127I, 131I, 14C, 3H, 32P, 35S

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and the like. The labeling material is not limited to them and any material which can be used for immunological determination can be used. Further, a low molecular weight hapten such as biotin, dinitrophenyl, pyridoxal or fluorescamine may be attached to the antibody. Preferably, horseradish peroxidase is used as a labeling enzyme. This enzyme can be reacted with various substrates and can readily be attached to the antibody by periodate method.

When an enzyme is used as a labeling material, a substrate and, if necessary, a coloring enzyme is used for measuring its activity. In case of using peroxidase as the enzyme,  $H_2O_2$  is used as a substrate and, as a coloring agent, there can be used 2,2'-azino-di-[3ethylbenzthiazoline sulfonic acid] ammonium salt (ABTS), 5'-aminosalicylic acid, o-phenylenediamine, aminoantipyrine, 3,3',5,5'-tetramethylbenzidine and like. In case of using alkaline phosphatase as the enzyme, o-nitorphenylphosphate, p-nitrophenylphosphoric acid, or the like can be used as a substrate. In case of using  $\beta\text{-}D\text{-}$ galactosidase enzyme, as the fluorescein-d-(B-Dgalactopyranoside), 4-methylumbelliphenyl-β-Dgalactopyranoside, or the like can be used as a substrate. The present invention also include a kit comprising the above monoclonal antibody, polyclonal antibody and reagents.

As a cross-linking agent, a known cross-linking

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N, N'-o-phenylenedimaleimide, agent such as 4-(Nmaleimidomethyl) cyclohexanoate-N-succinimide ester, 6maleimidohexanoate-N-succineimide ester, dithiopyridine or the like can be utilized. The reaction of these cross-linking agents with enzymes and antibodies be carried out by a known method according properties of a particular cross-linking agent. as the antibody, a fragment thereof, for example, Fab', Fab, F(b'2) can be used as the case may be. A labeled enzyme can be obtained by the same treatment regardless of whether the antibody is polyclonal or monoclonal. When the above labeled enzyme obtained by using a cross-linking agent is purified by a known method such as affinity chromatography or the like, a immunoassay system having more higher sensitivity can be obtained. The enzyme labeled and purified antibody is stored at a dark cold place with addition of a stabilizer such as thimerosal, glycerin or after lyophilization.

An objective to be determined is not specifically limited in so far as it is a sample containing BSSP2 or a fragment thereof, or a sample containing a precursor of BSSP2 or a fragment thereof and includes body fluids such as plasma, serum, blood, serum, urine, tissue fluid, cerebrospinal fluid and the like.

The following Examples further illustrate the

present invention in detail but are not construed to limit the scope thereof.

Example 1

Cloning of novel serine protease mBSSP2 gene

The cloning was carried out by PCR using a mouse brain cDNA library (Clontech) as a template and nucleotide sequences corresponding to an amino acid sequence common to serine proteases represented by

Primer 1: GTG CTC ACN GCN GCB CAY TG (SEQ ID NO: 20)

Primer 2: CCV CTR WSD CCN CCN GGC GA (SEQ ID NO: 21) Namely, 5  $\mu$ l of the template, 5  $\mu$ l of 10 xas primers. ExTaq buffer, 5 µl of dNTP, 10 pmol of each of the above primers and 0.5 µl of ExTaq (TAKARA) were added and the total volume was adjusted to 50 µl with sterilized water. PCR was carried out by repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for The PCR product was mixed with pCR 1 minutes, 30 times. II-TOPO vector attached to TOPO TA cloning kit (Invitrogen) and the mixture was allowed to stand at room temperature for 5 minutes. Then, according to a conventional manner, E. coli Top 10 attached to the kit was transformed and applied to a LB (Amp⁺) plate (containing 100 µg/ml of ampicillin). According to a conventional manner, a plasmid was extracted from each colony obtained and its nucleotide sequence was determined by cycle sequencing method with a fluorescence

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sequencer (ABI). Homology of the sequence of each clone was examined by means of GenBank. Regarding an unknown i.e., BSSP2 gene, the full length cDNA was obtained by 5' RACE and 3' RACE and, according to the same manner as described above, the nucleotide sequence was Namely, BSSP2 clone specific primers, GSP1 determined. primers [mBSSP2.2 (SEQ ID NO: 27) or mBSSP2.0 (SEQ ID NO: 22)] and GSP2 primers [mBSSP2R2 (SEQ ID NO: 28) or mBSSP2.1 (SEQ ID NO: 23)] were prepared. PCR was carried out by using mouse brain Marathon-Ready cDNA (Clontech), AP1 primer attached to this reagent and either of the above GSP1 primers and heating at 94°C for 2 minutes once and repeating a cycle of heating at 94°C for 30 seconds, at 60°C for 30 seconds and then at 72°C for 30 seconds 35 Then, 5 µl of the PCR product diluted to 1/100, 5 μl of 10 x buffer, 5 μl of dNTP, 10 pmol of either of 10 μM of the above GSP2 primer, 10 pmol of AP2 primer attached to the above reagent and 0.5 unit of ExTag were admixed and adjusted to 50 µl with sterilized water. Then, according to the same manner as the above, PCR was carried out. PCR product was cloned by the above TOPO TA cloning kit and sequenced to obtain the upstream and downstream regions of the above clone. At this time, as for a clone which seemed not to cover the full length of a protein, the specific primers shown hereinafter were prepared based on the newly

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found nucleotide sequence. Further, based on this sequence, the primers capable of amplifying ORF as shown hereinafter [mBSSPF7 (SEQ ID NO: 26), mBSSP2R/E (SEQ ID NO: 29)] were prepared and PCR carried out using mouse brain Marathonready cDNA as a template to confirm that these clones were was cloned into pCR II-TOPO vector identical. This attached to TOPO TA cloning kit to obtain the plasmid pCR II/mBSSP2 containing the full length cDNA clone. The nucleotide sequence of DNA contained in this plasmid is shown in SEQ ID NO: 7 and the amino acid sequence of mSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 8. Further, two different types of clones were The nucleotide sequences of these DNA are shown obtained. in SEQ ID NOS: 3 and 5, respectively. The amino acid sequences of mBSSP2 proteins deduced from these nucleotide sequences are shown in SEQ ID NOS: 4 and 6. These novel proteases are divided into types 1, 2 and 3. Type 1 is composed of 273 amino acids, type 2 is composed of 311 amino acids and type 3 is composed of 445 amino acids. These amino acid sequences contained the common amino acid sequence composed of 238 amino acids whose N-terminus side started with Ile-Val-Gly-Gly-Gln-Ala-Val as the serine protease. Further, in the amino acid sequence of the mature serine protease, a consensus sequence having a serine protease activity was contained. Furthermore, since

there were two or more amino acid sequence specific for a sugar chain bonding site, it was presumed that the amino acid sequence had at least two sugar chains.

Table 1

	5	SEQ	Name of	Direc-	Se	quence	Use
		ID	primer	tion			
		NO:					
g#: 51 <u>L</u>	10	22	mBSSP2.0	Forward	ATGGTGG.	AGAAGATCATTCCT	RACE
And And Mr. H.		23	mBSSP2.1	Forward	TACAGTG	CCCAGAACCATG	RACE
		24	mBSSPF4	Forward	CTCAACT	CTCTGCTAGACCG	RACE
The way of the		25	mBSSP2F5	Forward	ATAGTTG	GCGGCCAAGCTGT	mature
		26	mBSSPF7	Forward	CCCAGCA	GAACTTACTGCCT	FL*
		27	mBSSP2.2	Reverse	TGTTGCA	GAGGTGGGTGCTG	RACE
The state of the state of		28	mBSSP2R2	Reverse	TACCATT	GTGTCCTGCAGTGT	RACE
	15	29	mBSSP2R5/E	Reverse	TGAATTC	TGCTGCTTCTTCGGCTAGC	G FL*
\$c.m		*: f	or full ler	nath			

': for full length

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## Example 2

Expression mBSSP2 gene in mice internal organs

According to the protocol of QuickPrep Micro mRNA

purification Kit (Amersham-Pharmacia), mRNAs were isolated

from various internal organs of Balb/c mice or their

fetuses. They were subjected to electrophoresis according

to a conventional manner and transcribed to a nylon

membrane. A probe was prepared separately by isolating a

part of a nucleotide sequence encoding the mature protein

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of mBSSP2 from pCR II/mBSSP2, purifying it and labeling it with  $\alpha^{-32}P$  dCTP. The probe was diluted with  $5 \times SSC$  and reacted with the above membrane filter at 65°C for a whole day and night. Then, the filter was washed twice each with  $2 \times SSC/0.1\%$  SDS at room temperature for 30 minutes,  $1 \times C$ SSC/0.1% SDS at room temperature for 30 minutes and 0.1 x SSC/0.1% SDS at 65°C for 30 minutes. The filter was exposed to an imaging plate for FLA2000 (Fuji Film) for one day to analyze the expression. The results shown in the drawings are those obtained by using mRNAs prepared from head of fetuses of mice and mRNAs prepared from brain of 5day-, 10-day-, 14-day-, 18-day-, 30-day-, 3-month-, 7-month and 1-year-old mice (Fig. 1) and mRNAs prepared from various internal organs of 3-month-old mice (Fig. 2). addition, the mRNAs of mice prepared above were subjected to RT-PCR by using Ready To Go RT-PCR Beads (Amersham-Pharmacia) and mBSSP2 gene specific primers (SEQ ID NOS: 25 and 29) according to the protocol attached to the kit.

As seen form Figs. 1 and 2, in case of northern blotting analysis, the expression of mBSSP2 was recognized in head of 15th to 20th day fetuses of mice and, as to the 3-month-old mice, the expression was recognized in prostate and testicle. Further, according to the results of RT-PCR, the expression was recognized in head of 12-day-old mice and testicle of 3-month-old mice.

Example 3

Expression of novel serine protease mature protein encoded by mBSSP2 gene

(1) Construction of expression plasmid

A cDNA region encoding the mature protein of BSSP2 protein was amplified by PCR using the plasmid pCR II/mBSSP2 as a template (the sequence of the 1st to 717th bases of SEQ ID NO: 1 was amplified by using the primers having the sequences represented by SEQ ID NOS: 25 and 29). The PCR product was ligated to pTrc-HisB (Invitrogen) which had been digested with BamHI and blunted with mung bean nuclease. E. coli JM109 was transformed by the resultant and colonies formed were analyzed by PCR to obtain E. coli containing the desired serine protease expressing plasmid pTrcHis/mBBSP2.

The resultant *E. coli* was designated E. coli pTrcHis/mBSSP2 and deposited at National Institute of Bioscience and Human-Technology (NIBH), Agency of Industrial Science & Technology of 1-1-3 Higashi, Tsukubashi, Ibaraki-ken, Japan on October 29, 1998 under the accession numbers of FERM P-17033.

(2) Expression of protein by  $E.\ coli$  containing expression plasmid

A single colony of  $E.\ coli$  having the expression plasmid was inoculated in 10 ml of LB (Amp $^+$ ) culture medium

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and incubated at 37°C overnight. This was inoculated in 250 ml of LB (Amp⁺) culture medium and incubated at 37°C. When the absorbance at 600 nm became 0.5, 250  $\mu l$  of 0.1 M (isopropyl- $\beta$ -D-(-)-thiogalactopyranoside) was and the incubation was continued for additional 5 hours. The E. coli was centrifuged and suspended in a cell disruption buffer (10 mM phosphate buffer pH 7.5, 1 mM EDTA) and sonicated on ice to disrupt E. coli. This was centrifuged at 14,000 r.p.m. for 20 minutes to obtain a precipitate. The precipitate was washed twice with a cell disruption buffer containing 0.5% Triton  $X-100^{\text{TM}}$  and washed with water to remove Triton  $X-100^{TM}$ . Then, the resultant mixture was dissolved by soaking in a denaturation buffer containing 8 M urea (8M urea, 50 mM Tris pH8.5, 20 mM 2ME) at 37°C for 1 hour. The solution was passed through TALON affinity resin (Clontech), washed with denaturation buffer containing 10 mM imidazole, and then eluted with the denaturation buffer containing 100 mM imidazole to purify the solution. The purified product was dialyzed against PBS for 3 days with exchanging the buffer every other night to obtain the protein mBSSP2-His.

Example 4

Expression of novel serine protease mature protein encoded by mBSSP2 gene by using pFBTrypSigTag/BSSP2

(1) Construction of pFBTrypSigTag/BSSP2

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The sequences represented by SEQ ID NOS: 11 and 12 were subjected to annealing and digested with NheI and BamHI. The resultant fragment was inserted into pSecTag2A (Invitrogen) to obtain pSecTrypHis. Twenty units of BAmHI was added to 5 µg of pSecTrypHis vector and the vector was cleaved at 37°C over 4 hours. Then, 6 units of mung bean nuclease (TAKARA) was added thereto and reacted at room temperature (25°C) for 30 minutes to blunt the terminal ends. Further, the 3'-terminus side of the cloning site was digested cleaved with 20 units of XhoI, 1 unit of bacterial alkaline phosphatase (TAKARA) was added thereto and the reaction was carried out at 65°C for 30 minutes.

According to the same manner as that described in JP 9-149790 A or Biochim. Biophys. Acta, 1350, 11, 1997, prepared from COLO201 cells and was synthesized to obtain the plasmid pSPORT/neurosin. region of neurosin was obtained from active an pSPORT/neurosin by PCR using primers having the sequences represented by SEQ ID NOS: 13 and 14. Ten units of XhoI was reacted with the PCR product at 37°C for 3 hours to cleave XhoI site at the 3'-side thereof. This was inserted pSecTrypHis by TAKARA ligation kit to pSecTrypHis/neursoin (Fig. 3).

Amplification was carried out by using the primers having the sequences represented by SEQ ID NOS: 15

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and 16 so that the peptide of Leu-Val-His-Gly was present at the C-terminus of the part from trypsin signal to the enterokinase recognition site of pSecTrypHis/neurosin. This was inserted between NheI and HindIII sites of pSecTag2A to construct the plasmid pTrypSig.

One µg (0.1 µl) of the plasmid pSecTab2A was treated with the restriction enzymes NheI and BamHI to completely remove a region encoding the leader sequence of IgGk. One hundred pmol portions of DANs represented by SEQ ID NOS: 40 and 41 were added to the resultant solution and the mixture was heated at 70°C for 10 minutes and subjected to annealing by allowing to stand at room temperature for 30 minutes. Two µl of I solution of DNA ligation kit Ver. 2 (TAKARA) was added to 1 µl portions of His secretory signal sequence treated by NheI and BamHI and pSecTag2A and the reaction was carried out at 16°C for 30 minutes.

To the reaction mixture was add 0.1 ml of *E. coli* competent cell XL1-Blue (STRATAGENE) and reacted on ice for 30 minutes. Then, the reaction mixture was subjected to heat shock at 42°C for 60 seconds. After standing on ice for 2 minutes, 0.9 ml of SOC culture medium (Toyo Boseki K.K.) was added thereto and the mixture was shaken with a shaker at 37°C for 1 hour. The mixture was centrifuged at 5,000 r.p.m. for 1 minutes and the supernatant was discarded. The precipitated competent cells were suspended

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in the liquid remained in the centrifuge tube and the suspension was applied to 2 ampicillin LB plates containing  $100~\mu g/ml$  of ampicillin in the ratio of 1:10. The plates were incubated at  $37\,^{\circ}\text{C}$  for one night. Among the colonies formed, a colony into which DNA of His secretory signal was inserted was selected by PCR to obtain pTrypHis.

A sequence of about 200 bp containing His Tag region of pTrypHis was amplified by using primers having the sequence represented by SEQ ID NOS: 16 and 17 and a fragment of about 40 bp containing His Tag and enterokinase recognizing site formed by digestion of HindIII and BamHI was inserted into pTrypSig to construct pTrypSigTag (Fig. 4A).

cDNA was prepared by PCR of the sequence from trvpsin signal to enterokinase recognizing site pTrypSigTag using primers having the sequences represented by SEO ID NOS 14 and 18 and cut out by digestion with BglII and BamHI. It was inserted into BamHI site of pFastBAC1. The insertion direction was confirmed by PCR using primers having the sequences represented by SEQ ID NOS: 14 and 19. A clone into which the cDNA was inserted in the direction toward transcription and translation was selected to obtain pFBTrypSigTag.

Twenty units of BamHI was added to 5  $\mu g$  of pFBTrypSigTag vector and the vector was cleaved at 37°C

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over 4 hours, followed by addition of 6 units of mung bean nuclease (TAKARA) and reaction at room temperature (25°C) for 30 minutes to blunt the terminal ends. Further, the 3'-side of the cloning site was cleaved by 20 units of EcoRI, followed by addition of 1 unit of bacterial alkaline phosphatase (TAKARA). The reaction was carried out at 65°C for 30 minutes.

cDNA of the active region of mBSSP2 was obtained PCR according to a conventional manner using by or pCRII/mBSSP2 prepared from E. pTrcHis/mBSSP2 pTrcHis/mBSSP2 (accession No. FERM P-17033). The resultant into pFBTrypSigTag to was inserted CDNA pFBTrypSigTag/mBSSP2 (Fig. 4B). At this time, correct insertion of mBSSP2 was confirmed by determining the sequence.

Bacmid DNA was transformed PFBTrypSigTag/mBSSP2 according to a protocol of Gibco BRL BAC-TO-BAC baculovirus expression system to prepare a recombinant bacmid having chimera BSSP2 fused trypsinogen signal peptide, HisTag and enterokinase recognizing site. When this was expressed in Sf-9 cell according to a manual of BAC-TO-BAC baculovirus expression system, it was secreted in the culture supernatant from 2 days after infection of the virus.

(2) Determination of enzyme activity

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The recombinant fused protein mSSP2 obtained in the culture supernatant was passed through a chelate column to purify it and, after dialysis, its enzyme activity was determined. First, the culture supernatant was applied to a chelate column (Ni-NTA-Agarose, Qiagen) with PBS buffer and eluted stepwise with a solution of imidazole (Wako Pure Chemical Industries, Ltd.) dissolved in PBS. The resultant imidazole-eluted fraction was applied to a PD-10 column (Pharmacia) to exchange to PBS buffer. Fifty µl of this sample was mixed with 10  $\mu$ l of enterokinase (1 U/1  $\mu$ l, Invitrogen) and the reaction was carried out at room temperature for 60 minutes. Each of various synthetic substrates (Peptide Laboratory, Boc-Gln-Ala-Arg-MCA, Boc-Phe-Ser-Arg-MCA, Bz-Arg-MCA, Boc-Val-Leu-Lys-MCA, Pyr-Gly-Arg-MCA, Pro-Phe-Arg-MCA, Boc-Val-Pro-Arg-MCA, MCA, Arg-MCA, Z-Phe-Arg-MCA) was dissolved in DMSO and diluted with 1 M Tris-HCl (pH 8.0) to obtain a substrate Fifty µl of 0.2 M substrate solution was added thereto and further the reaction was carried out at 37°C. (7-amino-4fluorescence of AMC After one hour, the enzymatic reaction methylcoumalin) formed by the measured at 380 nm of excitation wavelength and 460 nm of fluorescence wavelength to determine the activity.

As a result, the recombinant fused protein mBSSP2 has been shown to have serine protease activity.

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Example 5

Cloning of hBSSP2 gene

Reverse transcription of 1 µg of mRNA of human fetus brain (Clontech) was carried out by using Superscript and oligo dT-Not Ι primer BRL) II (Gibco GGCCACGCGTCGACTAGTA C(T)₁₇ 3') to obtain cDNA. By using a template, PCR was carried out with primes as prepared from mBSSP2 nucleotide sequence and represented by SEO ID NOS: 30 and 31 to obtain a cDNA fragment of hBSSP2. Namely, 5 µl of the template, 5 µl of 10 x ExTaq buffer (TAKARA), 5  $\mu$ l of dNTPs, 10 pmol portions of the above primers and 0.5 µl of ExTaq (TAKARA) were adjusted to 50 µl with sterilized water and PCR was carried out by repeating a cycle of heating at 94°C for 0.5 minute, at 55°C for 0.5 minute and then at 72°C for 1 minute, 35 times. reactions described hereinafter were carried out according to the same manner as the above composition and conditions except the template and primers. The PCR product was mixed with pGEM-T Easy vector (Promega) and Takara Ligation Solution I (TAKARA) and the reaction was carried out at 16°C for 2 hours. Then, according to the same manner, E. coli JM109 was transformed and applied to a LB (Amp⁺) plate. A plasmid was extracted from each colony formed according to a conventional manner and its nucleotide sequence was determined by dideoxy method. As for a clone having

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homology to mBSSP2, full length cDNA was obtained by 5' and 3' RACE and its sequence was determined PCR was carried out by using the above described above. a template and primers having the sequences represented by SEQ ID NOS: 30 and 37. 3' RACE was carried out by PCR using a 1/100 dilution of the above PCR product as a template and primers having the sequences represented by SEQ ID NOS: 32 and 37. As for 5' RACE, cDNA for RACE was prepared from human fetal brain mRNA (Clontech) by using Superscript II and SMART RACE cDNA amplification kit PCR of this cDNA was carried out by using a primer of 10 x Universal Primer Mix (attached to the kit) and a primer having the sequence represented by SEQ ID NO: 33. Further, PCR was carried out by using the 1/100 dilution of the latter PCR product, a template, Nested PCR (attached to the kit) and a primer having the sequence represented by SEQ ID NO: 34. The finally obtained PCR product was subjected to TA cloning as described above and the nucleotide sequence was determined to obtain the upstream and downstream regions of the above clone. In addition, primers for amplifying the full length cDNA as represented by SEQ ID NOS: 35 and 36 were prepared based on the resultant nucleotide sequence and PCR was carried out by using the above synthetic cDNA as a template. This PCR product was cloned into pGEM-T Easy vector to

obtain the plasmid pGEM-TE/hBSSP2 containing the full length cDNA clone. The DNA sequence contained in this plasmid is shown in SEQ ID NO: 9 and hBSSP2 protein deduced from the nucleotide sequence is shown in SEQ ID NO: 10.

E. coli containing this plasmid was designated E. coli pGEM-TE/hBSSP2 and deposited at National Institute of Bioscience and Human-Technology (NIBH), Agency of Industrial Science & Technology of 1-1-3 Higashi, Tsukubashi, Ibaraki-ken, Japan on July 27, 1999 under the accession numbers of FERM P-17487.

Table 2

SEQ Name of

	ID	primer	tion			
	NO:					
15	30	BSSP2SPF	Forward	ACTGCTGCCCACTGCATG	for	part
	31	BSSP2SPR	Reverse	CAGGGGTCCCCCGCTGTCTCC	for	part
	32	hBSSP2F11	Forward	GCTCTCAACTTCTCAGACAC		RACE
	33	hBSSP2R12	Reverse	ACTCAGCTACCTTGGCGTAG		RACE
	34	hBSSP2R11	Reverse	CCTGGAGCATATCCGAGCTG		RACE
20	35	hBSSR2F12	Forward	GCTTTACAACAGTGCTAC		WB*
	36 h	BSSP2R13/E	Reverse	TGGAATTCGAGGAAACAGCAGGACT	'CAG	WB*
	37			TACTAGTCGACGCGTGGCC		

Direc- Sequence

Use

Example 6

Detection of hBSSP2 mRNA by northern blotting

^{*:} whole body

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Poly A + RNA extracted from respective tissues of adults and fetuses were blotted on a membrane (Clontech) and the membrane was subjected to northern hybridization with a hBSSP2 probe. The probe was labeled by Takara BcaBEST random labeling kit (TAKARA) according to random priming method using a cDNA fragment which was amplified by using the full length of hBSSP2 as a template and the sequences represented by SEQ ID NOS: 34 and 35 as primers. The hybridization was carried out at overnight and the filter was finally washed with 0.1 x SSC and 0.1% SDS. The radioactivity was detected by FLA-2000 (Fuji Film). The signal corresponding to the adult brain was recognized at about 2.4 kb, the signal corresponding to the adult skeletal muscle was recognized at 7 kb and 1.3 kb and further the signal of the fetus liver was recognized at The signal of the adult brain is considered 7 kb (Fig. 5). to correspond to the exact nucleotide sequence and the others are considered to correspond to polymorphic forms resulted from the difference in splicing.

20 Example 7

Detection of hBSSP2 mRNA by RT-PCR

mRNAs of human tissues purchased from Clontech were subjected to RT-PCR against hBSSP2 by using Ready To RT-PCR Beads (Amersham-Pharmacia) according to protocol attached to the kit. Expression of hBSSP2 was

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recognized in brain and skeletal muscle (Fig. 6). No clear band was obtained in pancreas due to the combination of primers. This is considered to be non-specific amplification by a large amount of a serine protease present in pancreas.

Example 8

Expression of hBSSP2 by baculovirus system

The signal sequence of human trypsinogen 2 and (His) 6 Tag and a sequence encoding the cleavage site of enterokinase were inserted into pFastBac1 (Gibco BRL) obtain the plasmid pFBTrypSigTag. The mature form of hBSSP2 was inserted into the plasmid pFBTrypSigTag so that it was located in the flame (Fig. 4B). The mature form of hBSSP2 amplified by the sequences represented by SEQ ID NOS: 38 and 36 was cleaved by EcoRI and, according to the same manner as described with respect to mBSSP2, it was inserted into pFBTrySigTag to construct pFastBacTrypSigTag/hBSSP2. At this time, correct insertion BSSP2 was confirmed by determining the nucleotide οf using the fluorescent sequence by labeled represented by SEQ ID NO: 39. Bacmid DNA was transformed with PFBTrypSigTag/hBSSP2 according to a protocol of Gibco BRL BAC-TO-BAC baculovirus expression system to prepare a recombinant bacmid having chimera BSSP2 fused with peptide, trypsinogen signal HisTag and enterokinase recognizing site. When this was expressed in Sf-9 cell according to a manual of BAC-TO-BAC baculovirus expression system and the culture supernatant from 3 days after infection of the virus subjected to western blot technique with anti-DDDDK antibody, a specific band was detected to confirm expression of hBSSP2 (Fig. 7).

Table 3

SEQ Name of Direc-

Sequence

Use

ID primer tion

10 NO:

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- 38 hBSSP2F13 Forward ACTGCTGCCCACTGCATG for part
- 39 FBTrypSigTagF5 GCGCTAGCAGATCTCCATGAATCTACTCCTGATCC NS*
- *: nucleotide sequence

## INDUSTRIAL UTILITY

According to the present invention, there are provided isolated human and mouse serine protease (hBSSP2 and mBSSP2) polynucleotides, their homologous forms, mature forms, precursors and polymorphic variants. Further, according to the present invention, there are provided hBSSP2 and mBSSP2 proteins as well as compositions containing hBSSP2 and mBssP2 polynucleotides and proteins, their production and use.

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SEQ ID NO: 11: Designed oligonucleotide to construct plasmid pSecTrypHis.

SEQ ID NO: 12: Designed oligonucleotide to construct plasmid pSecTrypHis.

SEQ ID NO: 13: Designed oligonucleotide primer to amplify neurosin-encoding sequence.

SEQ ID NO: 14: Designed oligonucleotide primer to amplify neurosin-encoding sequence.

SEQ ID NO: 15: Designed oligonucleotide primer to amplify a portion of plasmid pSecTrypHis/Neurosin.

SEQ ID NO: 16: Designed oligonucleotide primer to amplify a portion of plasmid pSecTrypHis/Neurosin.

SEQ ID NO: 17: Designed oligonucleotide primer to amplify a portion of plasmid pTrypHis.

SEQ ID NO: 18: Designed oligonucleotide primer to amplify a portion of plasmid pTrypSigTag.

SEQ ID NO: 19: Designed oligonucleotide primer to amplify a portion of plasmid pFBTrypSigTag.

SEQ ID NO: 20: Designed oligonucleotide primer to amplify conserved region of serine proteases-encoding sequence; n is a, c, g or t.

SEQ ID NO: 21: Designed oligonucleotide primer to amplify conserved region of serine proteases-encoding sequence; n is a, c, g or t.

SEQ ID NO: 22: Designed oligonucleotide primer designated as mBSSP2.0 for RACE for mBSSP2 (forward).

SEQ ID NO: 23: Designed oligonucleotide primer designated as mBSSP2.1 for RACE for mBSSP2 (forward).

SEQ ID NO: 24: Designed oligonucleotide primer designated as mBSSPF4 for RACE for mBSSP2 (forward).

SEQ ID NO: 25: Designed oligonucleotide primer designated as mBSSP2F5 to amplify mature mBSSP2-encoding region (forward).

SEQ ID NO: 26: Designed oligonucleotide primer designated as mBSSPF7 to amplify full-length mBSSP2-encoding mRNA (forward).

SEQ ID NO: 27: Designed oligonucleotide primer designated as mBSSP2.2 for RACE for mBSSP2 (reverse).

SEQ ID NO: 28: Designed oligonucleotide primer designated as mBSSP2R2 for RACE for mBSSP2 (reverse).

SEQ ID NO: 29: Designed oligonucleotide primer designated as mBSSP2R5/E to amplify full-length mBSSP2-encoding mRNA (reverse).

SEQ ID NO: 30: Designed oligonucleotide primer designated as BSSP2SPF to amplify a portion of hBSSP2 (forward).

SEQ ID NO: 31: Designed oligonucleotide primer designated as BSSP2SPR to amplify a portion of hBSSP2 (reverse).

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SEQ ID NO: 32: Designed oligonucleotide primer designated as hBSSP2F11 for RACE for hBSSP2 (forward).

SEQ ID NO: 33: Designed oligonucleotide primer designated as hBSSP2R12 for RACE for hBSSP2 (reverse).

5 SEQ ID NO: 34: Designed oligonucleotide primer designated as hBSSP2R11 for RACE for hBSSP2 (reverse).

SEQ ID NO: 35: Designed oligonucleotide primer designated as hBSSP2F12 to amplify full length hBSSP2 (forward).

SEQ ID NO: 36: Designed oligonucleotide primer designated as hBSSP2R13/E to amplify full length hBSSP2 (reverse).

SEQ ID NO: 37: Designed oligonucleotide primer for RACE for hBSSP2.

SEQ ID NO: 38: Designed oligonucleotide primer designated as hBSSP2F13 to amplify a portion of hBSSP2 (forward).

SEQ ID NO: 39: Designed oligonucleotide primer designated as FBTrpsigtagF5 to detect hBSSP2.

20 SEQ ID NO: 40: Designed oligonucleotide to construct plasmid pTrypHis.

SEQ ID NO: 41: Designed oligonucleotide to construct plasmid pTrypHis.

#### What is claimed is:

- 1. A protein having the amino acid sequence of 238 amino acids represented by SEQ ID NO: 2; or a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 2 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 2; or a modified derivative thereof.
- 2. A nucleotide sequence represented by the 1st to 714th bases of SEQ ID NO: 1; a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 2; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 2.
- 3. A protein having the amino acid sequence of 273 amino acids represented by SEQ ID NO: 4; or a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 4 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 4; or a modified derivative thereof.

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- 4. A nucleotide sequence represented by the 247th to 1065th bases of SEQ ID NO: 3; a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 4; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 4.
- 5. A protein having the amino acid sequence of 311 amino acids represented by SEQ ID NO: 6; or a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 6 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 6; or a modified derivative thereof.
- 6. A nucleotide sequence represented by the 516th to 1448th bases of SEQ ID NO: 5; a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 6; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 6.
  - 7. A protein having the amino acid sequence of

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455 amino acids represented by SEQ ID NO: 8; or a protein having an amino acid sequence derived from the amino acid sequence represented by SEQ ID NO: 8 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 8; or a modified derivative thereof.

- 8. A nucleotide sequence represented by the 116th to 1450th bases of SEQ ID NO: 7; a nucleotide sequence encoding the amino acid sequence represented by SEQ ID NO: 8; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by SEQ ID NO: 8.
- 9. A protein having the amino acid sequence of 240 amino acids represented by the 1st to 240th amino acids of SEQ ID NO: 10; or a protein having an amino acid sequence derived from the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10; or a modified derivative thereof.

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10. A nucleotide sequence represented by the 807th to 1526th bases of SEQ ID NO: 9; a nucleotide sequence encoding the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the 1st to 240th amino acids of SEQ ID NO: 10.

11. A protein having the amino acid sequence of 457 amino acids represented by the -217th to 240th amino acids of SEQ ID NO: 10; or a protein having an amino acid sequence derived from the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10; or a modified derivative thereof.

12. A nucleotide sequence represented by the 156th to 1526th bases of SEQ ID NO: 9; a nucleotide sequence encoding the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10; or a nucleotide sequence hybridizable with a nucleotide sequence

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which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the -217th to 240th amino acids of SEQ ID NO: 10.

- 13. A protein having the amino acid sequence of 217 amino acids represented by the -217th to -1st amino acids of SEQ ID NO: 10; or a protein having an amino acid sequence derived from the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10 by deletion, substitution or addition of one to several amino acids and having the same property as that of the protein having the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10; or a modified derivative thereof.
- 14. A nucleotide sequence represented by the 156th to 806th bases of SEQ ID NO: 9; a nucleotide sequence encoding the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein having the amino acid sequence represented by the -217th to -1st amino acids of SEQ ID NO: 10.

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- 15. A nucleotide sequence represented by SEQ ID NO: 1; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 1.
- NO: 3; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 3.
- NO: 5; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 5.
- 18. A nucleotide sequence represented by SEQ ID NO: 7; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding

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a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 7.

- NO: 9; or a nucleotide sequence hybridizable with a nucleotide sequence which is complementary to the above nucleotide sequence under stringent conditions and encoding a protein having the same property as that of the protein encoded by the nucleotide sequence represented by SEQ ID NO: 9.
  - 20. A vector comprising the nucleotide sequence according to any one of claims 2, 4, 6, 8, 10, 12 and 14-19.
  - 21. Transformed cells having the nucleotide sequence according to any one of claims 2, 4, 6, 8, 10, 12 and 14-19 in an expressible state.
  - 22. A process for producing a protein which comprises culturing cells transformed with the nucleotide sequence according to any one of claims 2, 4, 6, 8, 15-18, and collecting mBSSP2 produced.
- 23. A process for producing a protein which comprises culturing cells transformed with the nucleotide sequence according to any one of claims 10, 12, 14 or 19, and collecting hBSSP2 produced.
  - 24. The process according to claim 22 or 23, wherein the cells are *E. coli* cells, animal cells or insect

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cells.

- 25. A non-human transgenic animal whose expression level of BSSP2 gene has been altered.
- 26. The non-human transgenic animal according to claim 25, wherein BSSP2 gene is cDNA, genomic DNA or synthetic DNA encoding BSSP2.
- 27. The non-human transgenic animal according to claim 25, wherein the expression level has been altered by mutating a gene expression regulatory site.
- 28. A knockout mouse whose BSSP2 gene function is deficient.
- 29. An antibody against the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 or a fragment thereof.
- 30. The antibody according to claim 29 which is a polyclonal antibody, a monoclonal antibody or a peptide antibody.
- antibody against the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 or a fragment thereof which comprises administering the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13 or a fragment thereof to a warm-blooded animal other than a human being, selecting the animal whose antibody titer is recognized, collecting its spleen or lymph node, fusing the antibody producing cells

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contained therein with myeloma cells to prepare a monoclonal antibody producing hybridoma.

- 32. A method for determining the protein according to any one of claims 1; 3, 5, 7; 9, 11 and 13 or a fragment thereof in a specimen which is based on immunological binding of an antigen against the protein or a fragment thereof to the protein or a fragment thereof in the specimen.
- 33. A method for determining hBSSP2 or a fragment thereof in a specimen which comprises reacting a monoclonal antibody or a polyclonal antibody against the protein according to any one of claims 9, 11 and 13 or a fragment thereof and a labeled antibody with hBSSP2 or a fragment thereof in the specimen to detect a sandwich complex produced.
- 34. A method for determining hBSSP2 or a fragment thereof in a specimen which comprises reacting a monoclonal antibody or a polyclonal antibody against the protein according to any one of claims 9, 11 and 13 and a fragment thereof with labeled hBBSP2 and hBSSP2 or a fragment thereof in the specimen competitively to detect an amount of hBSSP2 or a fragment thereof in the specimen based on an amount of the labeled hBBSP2 reacted with the antibody.
- 35. The method according to any one of claims

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32-34, wherein the specimen is a body fluid.

- 36. A diagnostic marker for diseases in tissues comprising the protein according to any one of claims 1, 3, 5, 7, 9, 11 and 13.
- 37. The marker according to claim 36 to be used for diagnosis of Alzheimer's disease or epilepsy in brain.
- 38. The marker according to claim 36 to be used for diagnosis of cancer or inflammation of brain, prostate or testicle.
- 39. The marker according to claim 36 to be used for diagnosis of sterility in semen or sperms
- 40. The marker according to claim 36 to be used for diagnosis of prostatic hypertrophy in prostate.

## Abstract of the disclosure:

There are provided proteins having the amino acid sequences represented by SEQ ID NOS: 2, 4, 6, 8 and 10; proteins having amino acid sequences derived from these amino acid sequences by deletion, substitution or addition of one to several amino acids; and nucleotide sequences encoding the same; transgenic non-human animals with altered expression level of a serine protease BSSP2; an antibody against BSSP2; and a method for detecting BSSP2 in a specimen by using the antibody.

# mBSSP-2

	Fetus				Αf	te	er	birth					
	9th day	13th day	15th day	18th day	20th day	5-day-old	10-day-old	14-day-old	18-day-old	30-day-old	3-month-old	7-month-old	1-year-old
28S <del>-</del> 18S <b>-</b>													一種 大学学

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Fig. 2



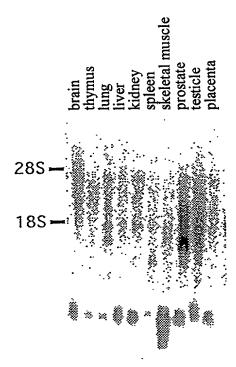
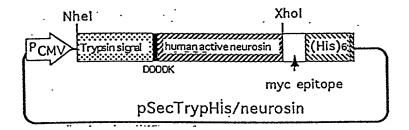
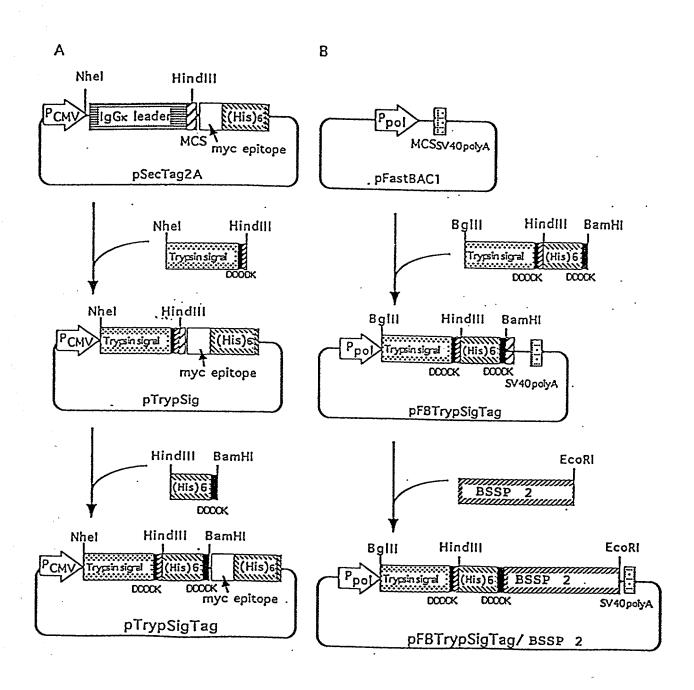


Fig. 3



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Fig. 4



5/7

Fig. 5

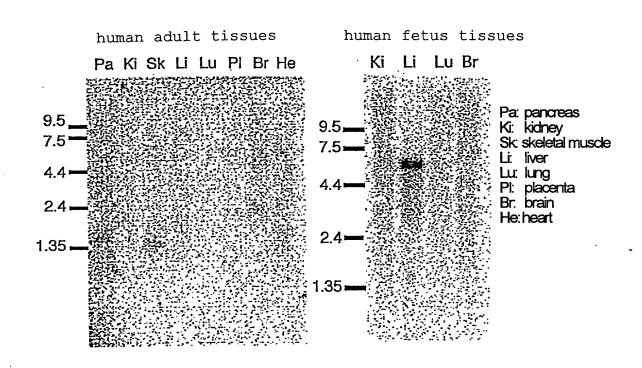


Fig. 6

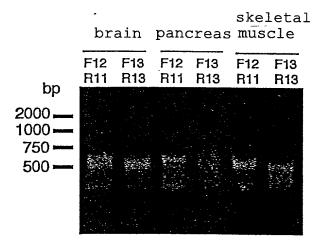


Fig. 7

kDa

97 —

50 ___

36 —

27.5

## Combined Declaration for Patent Application and Power of Attorney

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name; and that I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of

		for which a patent in PROTEASE	s sought on the inven BSSP2	tion entitled			
the specification of v	vhich (check one)						
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amendment referred information known b I hereby claim fore inventor's certificate	I to above; and I by me to be material ign priority benefit c, or prior PCT app	acknowledge the d to patentability as o s under 35 U.S.C. lication(s) designati	identified specifications to to disclose to the defined in 37 C.F.R. §§ 119 and 365 of any a country other the tion having a filing	he Patent and §1.56. any prior foreignan the U.S., lis	Trademark gn applicati sted below	office (PTO)  ion(s) for pater  with the "Yes"	nt or
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		the following regi	stered practioners to	prosecute this	application	and to transac	et all

All of the practioners associated with Customer Number 001444

Direct all correspondence to the address associated with Customer Number 001444; i.e.,

BROWDY AND NEIMARK, P.L.L.C. 624 Ninth Street, N.W. Washington, D.C. 20001-5303 (202) 628-5197

The undersigned hereby authorizes the U.S. Attorneys or Agents appointed herein to accept and follow instructions from AOYAMA & PARTNERS as to any action to be taken in the U.S. Patent and Trademark Office regarding this application without direct communication between the U.S. Attorneys or Agents and the undersigned. In the event of a change of the persons from whom instructions may be taken, the U.S. Attorneys or Agents appointed herein will be so notified by the undersigned.

Title: NOVEL SERINE PRO U.S. Application filed PCT Application filed Nov. 19, 1999  I hereby further declare that all statements made here information and belief are believed to be true; and the statements and the like so made are punishable by fine of false statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements made here information and belief are believed to be true; and the statements made here information and belief are believed to be true; and the statements made here information and belief are believed to be true; and the statements made here information and belief are believed to be true; and the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements made here information and the statements made her	, Serial No, Serial No. PCT/JP99/06 ein of my own knowledge are at these statements were made or imprisonment, or both, under	true and that all with the knowle 18 U.S.C. §1001 n.	dae that willful
PCT Application filed Nov. 19, 1999  I hereby further declare that all statements made here information and belief are believed to be true; and the statements and the like so made are punishable by fine of false statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements made here information and the statements made are punishable by fine of false statements may jeopardize the validity of the application and the statements may jeopardize the validity of the application and the statements may jeopardize the validity of the application and the statements may jeopardize the validity of the application and the statements made here information and the statemen	ein of my own knowledge are at these statements were made or imprisonment, or both, under cation or any patent issued thereous inventor's signature	true and that all with the knowle 18 U.S.C. §1001 n.	dae that willful
I hereby further declare that all statements made here information and belief are believed to be true; and the statements and the like so made are punishable by fine of false statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements may jeopardize the validity of the application of the statements made here information and the statements made here information and the statements made here information and the statements and th	ein of my own knowledge are at these statements were made or imprisonment, or both, under cation or any patent issued thereo	true and that all with the knowle 18 U.S.C. §1001 n.	dae that willful
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FULL NAME OF SECOND JOINT INVENTOR	INVENTOR'S SIGNATURE	Pan	T :
Akira OKUL			Apr. 17, 20
RESIDENT	akira Okii		142. 17, 20
Yamatokoriyama-shi, Nara Ja	nan TOY	CITIZENSHIP	
		Japan	
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FULL NAME OF THIRD JOINT INVENTOR	INVENTOR'S SIGNATURE	ia-siii, Na	DATE
Katsuya KOMINAMI	1.		Apr. 17, 20
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FULL NAME OF FOURTH JOINT INVENTOR			
Nozomi YAMAGUCHI	INVENTOR'S SIGNATURE	0	DATE
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FULL NAME OF FIFTH JOINT INVENTOR	INVENTOR'S SIGNATURE.		DATE
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FULL NAME OF SEVENTH JOINT INVENTOR	INVENTOR'S SIGNATURE		DATE
RESIDENT .		CITIZENSHIP	i

ALL INVENTORS MUST REVIEW APPLICATION AND DECLARATION BEFORE SIGNING. ALL ALTERATIONS MUST BE INITIALED AND DATED BY ALL INVENTORS PRIOR TO EXECUTION. NO ALTERATIONS CAN BE MADE AFTER THE DECLARATION IS SIGNED. ALL PAGES OF DECLARATION MUST BE SEEN BY ALL INVENTORS.

## SEQUENCE LISTING

<110> Fuso Pharmaceutical Industries Ltd.

5 <120> Novel serine protease BSSP2

<130> 661638

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<160> 41

<210> 1

15 <211> 717

ja#

<212> DNA

<213> mouse

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> 1 5 10 15

gtg atg ctt ggc tcc cgg cac acg tgt ggg gcc tct gtg ttg gca cca cac 102 Val Met Leu Gly Ser Arg His Thr Cys Gly Ala Ser Val Leu Ala Pro His

25 20 25 30

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	Trp	Val	Val	Thr	Ala	Ala	His	Cys	Met	Tyr	Ser	Phe	Arg	Leu	Ser	Arg	Leu	
	35					40					45					50		
	tcc	agc	tgg	cgg	gtt	cat	gca	ggg	ctg	gtc	agc	cat	ggt	gct	gtc	cga	caa	204
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210

215

220

gta gca gag ttc ctg gac tgg atc cat gac act gtg cag gtc cgc tagccga 1455 Val Ala Glu Phe Leu Asp Trp Ile His Asp Thr Val Gln Val Arg

225

230

235

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<212> PRT

<213> mouse

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	Leu Cys Asn Se	r Ser Cys Met	Tyr Ser Gly	Ala Leu Thr His	Arg Met Leu
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	Ser Trp Gly Ar	g Gly Cys Ala	Glu Pro Asn	Arg Pro Gly Val	Tyr Ala Lys
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	gag att cgg tg	c acg gaa gag	ggt gct ggg	cct ggg atc ttc	aga atg gag 169
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		-185		-180	-175	
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	Gln Arg (	Gly Cys Val	Ile Leu Gly	Val Leu Gly Leu	Leu Ala Gly	Ala Gly
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	Ile Ala S	Ser Trp Leu	Leu Val Leu	Tyr Leu Trp Pro	Ala Ala Ser	Pro Ser
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	Ile Ser (	Gly Thr Leu	Gln Glu Glu	Glu Met Thr Leu	Asn Cys Pro	Gly Val
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	Trp Leu L	Leu Val Cys	His Glu Gly	Trp Ser Pro Ala	Leu Gly Met	His Ile
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	Cys Lys S	Ser Leu Gly	His Ile Arg	Leu Thr Gln His	Lys Ala Val	Asn Leu
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	Ser	Asp	ı Ile	Lys	Leu	Asn	Arg	g Ser	G1n	Glu	Phe	Ala	G1n	Leu	Ser	·Ala	Arg	5
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	G1 y	Arg	Ile	Val	Ser	Leu	Lys	Cys	Ser	Glu	Cys	Gly	Ala	Arg	Pro	Leu	Ala	
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	Pro	His	Trp	Val	Val	Thr	Ala	Ala	His	Cys	Met	Tyr	Ser	Phe	Arg	Leu	Ser	
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	atc	aac	ttc	tca	gac	acc	gtg	gac	gct	gtg	tgc	ttg	ccg	gcc	aag	gag	cag	1087
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	Tyr	Phe	Pro	Trp	Gly	Ser	Gln	Cys	Trp	Val	Ser	Gly	Trp	Gly	His	Thr	Asp	
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235

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The first that the first that	10	tagaggatgc cctgtctcga gagttactag gcagataagg ttaaggttgg acaagctcag	1860
. Term		gtaaaggcac ggaagtcaag atcccctctc ccccgtgcgg tcctgttctg aggtaagcta	1920
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-195

-190

-200

225

220

25

-205

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	Gln	Arg	Gly	Cys	s Val	Ile	Leu	Gly	Val	Leu	Gly	Leu	Leu	ı Ala	Gly	Ala	Gly
			-170	)				-165	;				-160	)			
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Ann Ama						105					110					115		
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The States specific Stands				120	<b></b>		~	~	125		_			130				
e e	1 E		Ser	His	Thr	His		Ser	Asp	Thr	Leu	-	Asp	Thr	Met	Val		Leu
rei rei	15	135	Son	The	шіс	Lou	140	A an	Som	Com	Crra	145	Т	C	C1	41 -	150	T1 -
		Leu	Set	1111	155	Leu	Cys	ASII	ser	160	Cys	Met	туг	ser	165	А1а	Leu	ınr
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	20	Gly		Ser	Gly	Gly	Pro		Val	Cys	Pro	Ser		Asp	Thr	Trp	His	
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atg	agc	ctg	atg	ctg	gat	gac	caa	ccc	cct	atg	gag	gcc	cag	tat	gca	gag	206
Met	Ser	Leu	Met	Leu	Asp	Asp	Gln	Pro	Pro	Met	Glu	Ala	Gln	Tyr	Ala	Glu	
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Pro	Ile	Ser	Gln	Ala	Val	Cys	Trp	Arg	Ser	Met	Arg	Arg	G1y	Cys	Ala	Val	
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Leu		gcc	ctg		Leu		gcc	ggt		G1y		ggc	tca		Leu		359
Leu -	Gly -165	gcc Ala	ctg Leu	Gly	Leu -	Leu -160	gcc Ala	ggt Gly	Ala	Gly -	Val -155	ggc Gly	tca Ser	Trp	Leu -	Leu	

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	Leu	Leu	Glu	Ala	Gln	Val	Arg	Asp	Gln	Pro	Arg	Trp	Leu	Leu	Val	Cys	His	
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	Ser	Ser	Gln	Glu	Phe	Ala	Gln	Leu	Ser	Pro	Arg	Leu	G1y	G1y	Phe	Leu	Glu	,
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	Glu	Ala	Trp	Gln	Pro	Arg	Asn	Asn	Cys	Thr	Ser	G1y	Gln	Val	Val	Ser	Leu	
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	Arg	Cys	Ser	Glu	Cys	Gly	Ala	Arg	Pro	Leu	Ala	Ser	Arg	Ile	Val	G1y	Gly	
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	G1n	Ser	Val	Ala	Pro	G1y	Arg	Trp	Pro	Trp	Gln	Ala	Ser	Val	Ala	Leu	Gly	
	5					10					15					20		
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5	Phe	Arg	His	Thr	Cys	G1y	G1y	Ser	Val	Leu	Ala	Pro	Arg	Trp	Val	Val	Thr	
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	Ala	Ala	His	Cys	Met	His	Ser	Phe	Arg	Leu	Ala	Arg	Leu	Ser	Ser	Trp	Arg	
		40					45					50					55	
10	gtt	cat	gcg	ggg	ctg	gtc	agc	cac	agt	gcc	gtc	agg	ccc	cac	caa	ggg	gct	1022
•	Val	His	Ala	G1y	Leu	Val	Ser	His	Ser	Ala	Val	Arg	Pro	His	G1n	Gly	Ala	
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	Leu	Val	Glu	Arg	Ile	Ile	Pro	His	Pro	Leu	Tyr	Ser	Ala	Gln	Asn	His	Asp	
15			75				-	80					85					
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	Tyr	Asp	Val	Ala	Leu	Leu	Arg	Leu	Gln	Thr	Ala	Leu	Asn	Phe	Ser	Asp	Thr	
	90					95					100					105		
	gtg	ggc	gct	gtg	tgc	ctg	ccg	gcc	aag	gaa	cag	cat	ttt	ccg	aag	ggc	tcg	1175
2.0	Val	G1y	Ala	Val	Cys	Leu	Pro	Ala	Lys	Glu	Gln	His	Phe	Pro	Lys	G1y	Ser	
				110					115					120				
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	Arg	Cys	Trp	Val	Ser	Gly	Trp	Gly	His	Thr	His	Pro	Ser	His	Thr	Tyr	Ser	
		125					130					135					140	
25	tcg	gat	atg	ctc	cag	gac	acg	gtg	gtg	ссс	ttg	ttc	agc	act	cag	ctc	tgc	1277

Ser Asp Met Leu Gln Asp Thr Val Val Pro Leu Phe Ser Thr Gln Leu Cys

	145	150	155
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	Asn Ser Ser Cys Val Tyr	Ser Gly Ala Leu Thr	Pro Arg Met Leu Cys Ala
5	160	165	170
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	Gly Tyr Leu Asp Gly Arg	Ala Asp Ala Cys Gln	Gly Asp Ser Gly Gly Pro
	175 180	185	190
	cta gtg tgc cca gat ggg	gac aca tgg cgc cta	gtg ggg gtg gtc agc tgg 1430
10 [.]	Leu Val Cys Pro Asp Gly	Asp Thr Trp Arg Leu	Val Gly Val Val Ser Trp
	195	200	205
	ggg cgt gcg tgc gca gag	ccc aat cac cca ggt	gtc tac gcc aag gta gct 1481
, . [	Gly Arg Ala Cys Ala Glu	Pro Asn His Pro Gly	Val Tyr Ala Lys Val Ala
	210	215	220 225
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	Glu Phe Leu Asp Trp Ile	His Asp Thr Ala Gln	Asp Ser Leu Leu
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	ccagcagete cactaatgga g	gagaggcag tagcctccga	cacagaacgc atggacctcc 1646
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	tecetgeece tgtgtgagte t	tttagggag ggtgactggg a	aggtgeeece egteecacet 1946
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	ctgggcagat gggg	tcaagg ctgggcca	gt cccagatgaa gcc	ctatggga gtcaggac	cc 2066								
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	-200	-195	-190	-185	5								
	Pro Ile Ser Gli	n Ala Val Cys T	rp Arg Ser Met Ar	g Arg Gly Cys Ala	a Val								
	Pro Ile Ser Gla		rp Arg Ser Met Ar -175	g Arg Gly Cys Ala	a Val								
	-180	)	-175										
20	-180	)	-175	-170 1 Gly Ser Trp Leu									
20	-180 Leu Gly Ala Leu -165	) 1 Gly Leu Leu A -160	-175 la Gly Ala Gly Va -15	-170 1 Gly Ser Trp Leu	ı Leu -150								
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20	-180 Leu Gly Ala Leu -165 Val Leu Tyr Leu	) 1 Gly Leu Leu A -160 1 Cys Pro Ala A -145	-175 la Gly Ala Gly Va -15 la Ser Gln Pro Il -140	-170 1 Gly Ser Trp Leu 5 e Ser Gly Thr Leu	ı Leu -150 ı Gln								

Leu Pro Ala Leu Pro Lys Thr Val Ser Phe Arg Ile Asn Ser Glu Asp Phe

		-115				-	-110				-	-105				-	-100	
		Leu	Leu	Glu	Ala	Gln	Val	Arg	Asp	Gln	Pro	Arg	Trp	Leu	Leu	Val	Cys	His
	٠				-95					-90					-85			
		Glu	G1y	Trp	Ser	Pro	Ala	Leu	Gly	Leu	G1n	Ile	Cys	Trp	Ser	Leu	Gly	His
5			-80					-75					-70					-65
		Leu	Arg	Leu	Thr	His	His	Lys	Gly	Val	Asn	Leu	Thr	Asp	Ile	Lys	Leu	Asn
						-60					-55					-50		
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				-45					-40					-35				
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		-30				-	-25				-	-20				-	-15	
		Arg	Cys	Ser	Glu	Cys	G1y	Ala	Arg	Pro	Leu	Ala	Ser	Arg	Ile	Val	Gly	G1y
					-10					-5				-1	1			
	• .	Gln	Ser	Val	Ala	Pro	G1y	Arg	Trp	Pro	Trp	Gln	Ala	Ser	Val	Ala	Leu	G1y
15		5					10					15					20	
		Phe	Arg	His	Thr	Cys	Gly	Gly	Ser	Val	Leu	Ala	Pro	Arg	Trp	Val	Val	Thr
					25					30					35			
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			40					45					50					55
20		Val	His	Ala	Gly	Leu	Val	Ser	His	Ser	Ala	Val	Arg	Pro	His	Gln	Gly	Ala
						60					65					70		
		Leu	Val	Glu	Arg	Ile	Ile	Pro	His	Pro	Leu	Tyr	Ser	Ala	Gln	Asn	His	Asp
				75					80					85				
		Tyr	Asp	Val	Ala	Leu	Leu	Arg	Leu	G1n	Thr	Ala	Leu	Asn	Phe	Ser	Asp	Thr
25		90					95					100					105	

	Val	G1y	Ala	Val	Cys	Leu	Pro	Ala	Lys	Glu	Gln	His	Phe	Pro	Lys	Gly	Ser
				110					115					120			
	Arg	Cys	Trp	Val	Ser	G1y	Trp	G1y	His	Thr	His	Pro	Ser	His	Thr	Tyr	Ser
		125					130					135					140
5	Ser	Asp	Met	Leu	G1n	Asp	Thr	Val	Val	Pro	Leu	Phe	Ser	Thr	Gln	Leu	Cys
					145					150					155		
	Asn	Ser	Ser	Cys	Val	Tyr	Ser	Gly	Ala	Leu	Thr	Pro	Arg	Met	Leu	Cys	Ala
			160					165					170				
	G1y	Tyr	Leu	Asp	G1y	Arg	Ala	Asp	Ala	Cys	G1n	Gly	Asp	Ser	G1y	Gly	Pro
10	175					180					185					190	
	Leu	Val	Cys	Pro	Asp	G1y	Asp	Thr	Trp	Arg	Leu	Val	G1y	Val	Val	Ser	Trp
				195					200					205			
	Gly	Arg	Ala	Cys	Ala	Glu	Pro	Asn	His	Pro	G1y	Val	Tyr	Ala	Lys	Val	Ala
		210					215					220					225
15	G1u	Phe	Leu	Asp	Trp	Ile	His	Asp	Thr	Ala	G1n	Asp	Ser	Leu	Leu		
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<211> 99

20 <212> DNA

<213> Artificial Sequence

<220>

 $\ensuremath{\texttt{\langle 223\rangle}}$  Designed oligonucleotide to construct plasmid pSecTrypHis

25 <400> 11

10

15

20

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                                                          99
tttgacgacg atgacaagga tccgaattc
<210> 12
<211> 99
<212> DNA
<213> Artificial Sequence
<220>
<223> Designed oligonucleotide to construct plasmid pSecTrypHis
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gtagattcat ggtgttgcta gccaagctt
                                                          99
⟨210⟩ 13
<211> 15
<212> DNA
<213> Artificial Sequence
<220>
\langle 223 \rangle Designed oligonucleotide primer to amplify neurosin-encoding sequence
⟨400⟩ 13
ttggtgcatg gcgga
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25 <210> 14

<211> 27

<212> DNA

<213> Artificial Sequence

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25

<220>

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<220>
     5
            <223> Designed oligonucleotide primer to amplify neurosin-encoding sequence
            ⟨400⟩ 14
            tcctcgagac ttggcctgaa tggtttt
                                                                                    27
A. The the time that the the
    10
            <210> 15
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   15
            <223>
                    Designed oligonucleotide primer to amplify
                                                                            portion of
            pSecTrypHis/Neurosin
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                                                                                    35
   20
            ⟨210⟩ 16
            <211> 29
            <212> DNA
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 $\ensuremath{\texttt{\langle 223\rangle}}$  Designed oligonucleotide primer to amplify a portion of plasmid pSecTrypHis/Neurosin

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5 tgaagettge catggaccaa ettgteate

29

<210> 17

<211> 26

<212> DNA

10 <213> Artificial Sequence

<220>

<223> Designed oligonucleotide primer to amplify a portion of plasmid pTrypHis

<400> 17

15 ccaagettea ceateaceat caceat

26

⟨210⟩ 18

<211> 17

<212> DNA

20 <213> Artificial Sequence

<220>

<223> Designed oligonucleotide primer to amplify a portion of plasmid
pTrypSigTag

25 <400> 18

gcacagtcga ggctgat

<400> 20

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20
        gtgctcacng cngcbcaytg
        <210> 21
 5
        ⟨211⟩ 20
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        <220>
        <223> Designed oligonucleotide primer to amplify conserved region of serin
        proteases-encoding sequence
        <220>
        <221> UNSURE
        <222> 12, 15
        \langle 223 \rangle n is a, c, g or t.
        <400> 21
                                                                              20
        ccvctrwsdc cnccnggcga
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25
        (forward)
```

**<400> 22** 

<210> 25

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<211> 20
        <212> DNA
 5
        <213> Artificial Sequence
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        mBSSP2-encoding region (forward)
10
         <400> 25
                                                                                 20
         atagttggcg gccaagctgt
         <210> 26
         <211> 20
15
         <212> DNA
         <213> Artificial Sequence
         <220>
         \ensuremath{\texttt{\langle 223\rangle}} Designed oligonucleotide primer designated as mBSSPF7 to amplify full-
         length mBSSP2-encoding mRNA (forward)
20
         <400> 26
                                                                                 20
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         <210> 27
25
         ⟨211⟩ 20
```

<212> DNA

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<213> Artificial Sequence
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        \langle 223 \rangle Designed oligonucleotide primer designated as mBSSP2.2 for RACE for mBSSP2
 5
         (reverse)
        <400> 27
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10
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15
         (reverse)
         <400> 28
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20
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         <213> Artificial Sequence
25
         <220>
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<223> Designed oligonucleotide primer designated as mBSSP2R5/E to amplify full-length mBSSP2-encoding mRNA (reverse)

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27

<210> 30

<211> 18

<212> DNA

10 <213> Artificial Sequence

<220>

<223> Designed oligonucleotide primer designated as BSSP2SPF to amplify a
portion of hBSSP2 (forward)

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18

<210> 31

<211> 21

20 <212> DNA

<213> Artificial Sequence

<220>

<223> Designed oligonucleotide primer designated as BSSP2SPR to amplify a
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25

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<400> 31
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21

<210> 32

5 〈211〉 20

<212> DNA

<213> Artificial Sequence

<220>

 $\langle 223 \rangle$  Designed oligonucleotide primer designated as hBSSP2F11 for RACE for

10 hBSSP2 (forward)

<400> 32

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20

15 <210> 33

⟨211⟩ 20

<212> DNA

<213> Artificial Sequence

<220>

20 <223> Designed oligonucleotide primer designated as hBSSP2R12 for RACE for hBSSP2 (reverse)

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20

<210> 34

<211> 20

```
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 5
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        \ensuremath{\texttt{\langle 223\rangle}} Designed oligonucleotide primer designated as hBSSP2R11 for RACE for
        hBSSP2 (reverse)
         ⟨400⟩ 34
                                                                                   20
10
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         <211> 18
         <212> DNA
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15
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         \langle 223 \rangle Designed oligonucleotide primer designated as hBSSP2F12 to amplify full
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20
         <400> 35
                                                                                    18
         gctttacaac agtgctac
         <210> 36
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         <212> DNA
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<213> Artificial Sequence

A The State Com Said See State

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        \langle 223 \rangle Designed oligonucleotide primer designated as hBSSP2R13/E to amplify full
        length hBSSP2 (reverse)
 5
        <400> 36
                                                                                 28
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        <211> 19
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        \langle 223 \rangle Designed oligonucleotide primer for RACE for hBSSP2
15
         <400> 37
                                                                                  19
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20
         <211> 18
         <212> DNA
         <213> Artificial Sequence
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         \langle 223 \rangle Designed oligonucleotide primer designated as hBSSP2F13 to amplify a
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25
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<400> 38

```
18
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 5
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        <211> 35
        <212> DNA
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10
        hBSSP2
        <400> 39
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        <210> 40
        <211> 117
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        <213> Artificial Sequence
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         <223> Designed oligonucleotide to construct plasmid pTrypHis
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25
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⟨210⟩ 41

<211> 117

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<220>

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<400> 41

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